Shiksha Mandal's Bajaj College of Science, Wardha

COLLEGE OR

WARDHA

(Formerly known as Jankidevi Bajaj College of Science)

4th Cycle: Assessment and Accreditation by NAAC

Criterion III Research, Innovations

and Extension

QnM 3.4.4

Number of books and chapters in edited volumes / books published per teacher

Establishment Year : 1962 AISHE Code : C-18638 Shiksha Mandal's

Uni. College No. 652 Jr. College No. 07-01-003



Bajaj College of Science, Wardha

(Formerly known as Jankidevi Bajaj College of Science) An Autonomous College affiliated to RTM Nagpur University, Nagpur NAAC Reaccredited 'A' with CGPA of 3.21 (A Linguistic Minority College)



Date : 2/7 DEC 2022

Prof. Pradip. V. Tekade Principal (Offg.)

Email : jbsciencewardha@yahoo.co.in pradiptekade@gmail.com

Ref. No. BCS/ /20 -20

DECLARATION

This is to declare that the information and data furnished as supporting/additional document in the metric 3.4.4 Number of books and chapters in edited volumes / books published per teacher are true to the best of my knowledge and is verified by IQAC.

Dr. M. R. Chandrakar IQAC Co-ordinator

AJ E OF SC

Prof. R.V. Tekade

Principal (Offg.) Offi. Principal Bajaj College of Science WARDHA

BIOGENIC SUSTAINABLE NANOTECHNOLOGY

Trends and Progress

Edited by Raghvendra Pratap Singh Alok R. Rai Ahmed Abdala Ratiram G. Chaudhary



Micro & Nano Technologies Series

Elsevier Radarweg 29, PO Box 211, 1000 AE Amsterdam, Netherlands The Boulevard, Langford Lane, Kidlington, Oxford OX5 1GB, United Kingdom 50 Hampshire Street, 5th Floor, Cambridge, MA 02239, United States

Copyright @ 2022 Elsevier Inc. All rights reserved.

No part of this publication may be reproduced or transmitted in any form or by any means, electronic or mechanical, including photocopying, recording, or any information storage and retrieval system, without permission in writing from the publisher. Details on how to seek permission, further information about the Publisher's permissions policies and our arrangements with organizations such as the Copyright Clearance Center and the Copyright Licensing Agency, can be found at our website: www.elsevier.com/permissions.

This book and the individual contributions contained in it are protected under copyright by the Publisher (other than as may be noted herein).

Notices

Knowledge and best practice in this field are constantly changing. As new research and experience broaden our understanding, changes in research methods, professional practices, or medical treatment may become necessary.

Practitioners and researchers must always rely on their own experience and knowledge in evaluating and using any information, methods, compounds, or experiments described herein. In using such information or methods they should be mindful of their own safety and the safety of others, including parties for whom they have a professional responsibility.

To the fullest extent of the law, neither the Publisher nor the authors, contributors, or editors, assume any liability for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products, instructions, or ideas contained in the material herein.

ISBN: 978-0-323-88535-5

For information on all Elsevier publications visit our website at https://www.elsevier.com/books-and-journals

Publisher: Matthew Deans Acquisitions Editor: Sabrina Webber Editorial Project Manager: Clodagh Holland-Borosh Production Project Manager: Sojan P. Pazhayattil Cover Designer: Greg Harris



Typeset by MPS Limited, Chennai, India



Contents

List of contributors xvii Preface xxiii

1.	Building nanomaterials with microbial factories	
	PABLO E. ANTEZANA, SOFIA MUNICOY AND MARTIN F. DESIMONE	
	1.1 Introduction	1
	1.2 Mechanisms of metal nanoparticles synthesis by bacteria	3
	1.3 Nanoparticle biosynthesis	7
	1.3.1 Silver nanoparticles	7
	1.3.2 Gold nanoparticles	- 11
	1.3.3 Magnetite nanoparticles	14
	1.3.4 Copper nanoparticles	19
	1.3.5 Selenium nanoparticles	21
	1.3.6 Quantum dots	24
	1.4 Conclusion	
	1.5 Future prospects	29
	Acknowledgments	30
	References	30
2.	Phytofabrication of nickel-based nanoparticles: focus on environmental benign technology and therapeutic perspectives	41
	KANHAIYA M. DADURE, DEBARSHI KAR MAHAPATRA, ANIMESHCHANDRA G.M. HALDAR, RATIRAM G. CHAUDHARY AND AJAY K. POTBHARE	00(3).
	2.1 Introduction	41
	2.2 Current status of Ni-based nanoparticles	44
		v

vi Contents

	2.3 Fabrication of Ni-based nanoparticles	47
	2.3.1 Fabrication of NiO nanoparticles using plant extracts	47
	2.3.2 Fabrication of NiO nanoparticles using microbes	47
	2.4 Conclusion and future perspectives of Ni-based nanoparticles	51
	References	51
3.	Bacterial cellular mechanisms for synthesis of green nanostructured compounds	59
	SIMPAL KUMARI, ZHI FENG LI AND MIAN NABEEL ANWAR	
	3.1 Introduction	60
	3.2 Microorganism involved in the synthesis of nanoparticles	60
	3.2.1 Bacteria	61
	3.2.2 Fungi	62
	3.2.3 Cyanobacteria	64
	3.2.4 Others	64
	3.3 Synthesis of bacterial nanoparticles by using cellular mechanism	65
	3.3.1 Extracellular mechanism	65
	3.3.2 Intracellular mechanism	67
	3.4 Application of biologically synthesized nanoparticles	67
	3.4.1 Food	68
	3.4.2 Agriculture	68
	3.4.3 Environment	68
	3.4.4 Biomedical	68
	3.4.5 Textiles	69
	3.4.6 Renewable energy	69
	3.4.7 Electronics	69
	3.5 Conclusion	69
	References	69

4.	Ecofriendly microorganism assisted fabrication of metal nanoparticles and their applications	77
	SUDIP MONDAL, MANJIRI S. NAGMOTE, SURAJ V. KOMBE, BARUN K. DUTTA, TRIMURTI L. LAMBAT, PRASHANT B. CHOUKE AND ANIRUDDHA MONDAL	
	4.1 Introduction	77
	4.2 Bacteria-mediated synthesis	79
	4.2.1 Copper nanoparticles synthesis by bacterial font	79
	4.2.2 Silver nanoparticles synthesis by bacterial font	81
	4.2.3 Gold nanoparticles synthesis using different bacterial font	86
	4.3 Fungi-mediated synthesis	91
	4.3.1 Copper nanoparticles synthesis by fungi font	91
	4.3.2 Silver nanoparticles synthesis by fungi font	94
	4.3.3 Gold nanoparticles synthesis by fungi font	98
	4.4 Conclusion	99
	References	100
5.	Herbal spices and nanotechnology for the benefit of human health	107
	SHALINI JIWAN CHAHANDE, RASHMI JACHAK, RAGINI CHAHANDE AND PALLAVI PANTAWANE	
	5.1 Introduction	107
	5.2 Complementary role of spices and nanotechnology in development of herbal medicine	108
	5.3 Journey of spices for the betterment of human life	111
	5.3.1 Spices from kitchen to clinic	111
	5.4 Ancient to current status of the use of herbal spices and nanotechnology	112
	5.5 Use of spices as a source of natural color	115
	5.6 Use as a natural source of antioxidant and antimicrobial agents	116
	5.7 Need for bioprospection of herbs and spices	117

5.7.1 Medicinal bioprospecting	118
5.7.2 Bioprospection of essential oils for medicinal uses	118
5.7.3 Bioprospection of products from herbs and spices	119
5.7.4 Bioprospecting of spices and herbs for drug discovery	121
5.8 Issues and challenges with herbal nanomedicines	121
5.9 Conclusion and future perspectives	123
References	123
Nanoparticles for sustainable agriculture: innovative	
potential with current and future perspectives	131
SUBHASH RUPCHAND SOMKUWAR, RATNNADEEP C. SAWANT, PRASHANT P. INGALE, DHANRAJ T. MASRAM AND RUPALI RAMESH CHAUDHARY	
6.1 Introduction	131
6.2 Nanopesticides: agro-based formulations for pest control	132
6.3 Nanofertilizers: recent trends and prospect in agriculture system	134
6.4 Nanoparticles: uptake, translocations, and plant growth	136
6.5 Recent advances in nanoparticles for plant protection	137
6.6 Nanomaterials as agents to smart monitoring	138
6.7 Nanoparticles for managing the agricultural postharvest waste	139
6.8 Future perspective	140
6.9 Conclusion	140
References	140
Fabrications and applications of	
polymer-graphene nanocomposites for sustainability	149
AJAY K. POTBHARE, TRUPTI S. SHRIRAME, VIDYASAGAR DEVTHADE, SACHIN T. YERPUDE, MAYURI S. UMEKAR, RATIRAM G. CHAUDHARY AND GANESH S. BHUSARI	
7.1 Introduction	150
7.2 History background of polymer-graphene nanocomposites	152
	 5.7.2 Bioprospection of essential oils for medicinal uses 5.7.3 Bioprospecting of products from herbs and spices 5.7.4 Bioprospecting of spices and herbs for drug discovery 5.8 Issues and challenges with herbal nanomedicines 5.9 Conclusion and future perspectives References Nanoparticles for sustainable agriculture: innovative potential with current and future perspectives SUBHASH RUPCHAND SOMKUWAR, RATINNADEEP C. SAWANT, PRASHANT P. INGALE, DHANRAU T. MASRAM AND RUPALI RAMESH CHAUDHARY 6.1 Introduction 6.2 Nanopesticides: agro-based formulations for pest control 6.3 Nanofertilizers: recent trends and prospect in agriculture system 6.4 Nanoparticles: uptake, translocations, and plant growth 6.5 Recent advances in nanoparticles for plant protection 6.6 Nanomaterials as agents to smart monitoring 6.7 Nanoparticles for managing the agricultural postharvest waste 6.8 Future perspective 6.9 Conclusion References Fabrications and applications of polymer – graphene nanocomposites for sustainability AJAY K. POTBHARE, TRUPTI S. SHRIRAME, VIDYASAGAR DEVTHADE, SACHIN T. YERPUDE, MAYURI S. UMEKAR, RATIRAM G. CHAUDHARY AND GANESH S. BHUSARI 7.1 Introduction

7.3 Overview of polymer-graphene nanocomposites	154
7.4 Preparation methods polymer-graphene nanocomposites	155
7.4.1 Solution cast technique	155
7.4.2 Melt mixing technique	156
7.4.3 In situ polymerization	157
7.4.4 Electrospinning technique	157
7.4.5 Electrodeposition	158
7.5 Modification techniques for graphene and graphene oxide	158
7.5.1 Grafting	158
7.5.2 Atom transfer radical polymerization	158
7.5.3 Radical polymerization techniques	159
7.5.4 Condensation techniques	159
7.6 Interactions of graphene oxide and graphene with polymers	160
7.6.1 Interactions of graphene oxide in polymer matrices	160
7.6.2 Interactions of graphene in polymer matrices	160
7.7 Natural polymers nanocomposites	161
7.7.1 Chitosan/graphene/graphene oxide nanocomposites	161
7.7.2 Cellulose/graphene/graphene oxide nanocomposites	162
7.8 Synthetic polymers nanocomposites	162
7.8.1 Polyvinylidene fluoride/graphene/graphene oxide nanocomposites	162
7.8.2 Polyurethane/graphene/graphene oxide nanocomposites	163
7.9 Conductive polymers nanocomposites	164
7.9.1 Polypyrrole/graphene/graphene oxide nanocomposites	164
7.9.2 Polyaniline/graphene/graphene oxide nanocomposites	165
7.10 Applications of graphene/polymer nanocomposites	166
7.10.1 Antibacterial activity	166
7.10.2 Sensors	167

x Contents

	7.10.3 Energy storage devices	168
	7.10.4 High-performance materials	169
	7.10.5 Drug delivery	170
	7.10.6 Biomedical	171
	7.10.7 Water purification	171
	7.11 Conclusion	173
	References	174
8.	Phytofabrication of metal oxide/iron-based and their therapeutic and their therapeutic potentials: in-depth insights into the recent progress ANIMESHCHANDRA G.M. HALDAR, DEBARSHI KAR MAHAPATRA,	185
	KANHAIYA M. DADURE AND RATIRAM G. CHAUDHARY	
	8.1 Introduction	186
	8.1.1 Different ways to define NPs	186
	8.1.2 Development from ancient to scientific age	187
	8.2 Methods for nanoparticles fabrication	187
	8.2.1 Mechanical grinding/milling	189
	8.2.2 Laser ablation	189
	8.2.3 Electro-explosion	190
	8.2.4 Chemical vapor deposition	190
	8.2.5 Sol-gel process	190
	8.2.6 Biological fabrication	190
	8.3 Biofabrication of NPs	191
	8.4 Phytofabrication of NPs	192
	8.4.1 Stem-based phytofabrication	193
	8.4.2 Fruit-based phytofabrication	193
	8.4.3 Seed/seed coats-based phytofabrication	193
	8.4.4 Flower-based phytofabrication	194
	8.4.5 Root-based phytofabrication	194

	8.4.6 Leaves-based phytofabrication	194
	8.5 Mechanism of phytofabrication of NPs	194
	8.6 Therapeutic potentials of iron-based NPs	197
	8.7 Conclusion	204
	References	204
9.	Highlights of decade long progress of nano-selenium fabricated from plant biomass: insights into techniques and mechanisms	217
	DEBARSHI KAR MAHAPATRA, ANIMESHCHANDRA G.M. HALDAR AND KANHAIYA M. DADURE	
	9.1 Introduction	217
	9.2 Selenium nanoparticles	219
	9.3 Synthesis	219
	9.4 Mechanism of formation of SeNPs	220
	9.5 Recent reports of SeNPs formation	221
	9.6 Applications in SeNPs in food packing	222
	9.7 Toxicity of SeNPs	223
	9.8 Conclusion	224
	References	224
10.	Strategies of nanotechnology as a defense system in plants	227
	RASHMI JACHAK, SHALINI CHAHANDE, JAYSHREE THAWARE AND RUPALI MAHAKHODE	
	10.1 Introduction	227
	10.2 Nanotechnology in plant defense mechanism	229
	10.2.1 Nanobiosensors	229
	10.2.2 Nanoencapsulation	231
	10.2.3 Metal-based nanoparticles	232

	10.2.4 Nanohybrid	234
	10.2.5 Nanoantioxidant mechanism	236
	10.3 Nanotoxicity and nanobusiness	237
	10.3.1 Nanotoxicity—monitored toxicity and potential health risks of nanomaterials	237
	10.3.2 Nanobusiness and its risky path	239
	10.4 Conclusion	239
	Future line of work	240
	References	240
	Further reading	248
11.	Nanocomposites for dye remediation from aqueous solutions	249
	N.B. SINGH, N.P. SINGH, A.K. SINGH AND LELLOUCHE JEAN-PAUL	
	11.1 Introduction	249
	11.2 Dyes	251
	11.3 Nanocomposites	251
	11.3.1 Magnetic nanocomposites	253
	11.3.2 Metal/metal oxide-based nanocomposites	253
	11.3.3 Polymer nanocomposites	255
	11.3.4 Hydroxyapatite nanocomposites	257
	11.3.5 Carbon-based nanocomposites	257
	11.3.6 Ash-based nanocomposites	258
	11.3.7 Hydrogel-based nanocomposites	258
	11.3.8 Chitosan-based nanocomposites	261
	11.3.9 Other types of nanocomposites	261
	11.4 Photocatalytic degradation of dyes	261
	11.5 Conclusion	265
	References	265

12.	Sustainable hybrid nanomaterials for environmental remediation and agricultural advancement	267
	PRERNA KHAGAR, SANGESH ZODAPE, UMESH PRATAP, ATUL MALDHURE, GAYATRI GAIKVIAD AND ATUL WANKHADE	267
	12.1 Introduction	267
	12.1.1 Hybrid nanomaterials	269
	12.1.2 Designing strategy and properties of hybrid nanomaterials	269
	12.2 Applications of hybrid nanomaterials	272
	12.2.1 Polymer-based hybrid nanomaterial	272
	12.2.2 Metal-organic framework	274
	12.2.3 Phytochemical-based hybrid nanomaterials	279
	12.3 Future aspects	283
	12.4 Concluding remarks	283
	References	283
13.	Bacterial synthesis of zinc oxide nanoparticles	
	and their applications	293
	LEKSHMI GANGADHAR, NALLURI ABHISHEK, MADUTHURI VENKATESH. V.V.S. PRASAD, PENTAKOTA SURYA NAGENDRA, MADAKKA MEKAPOGU, AMAR P. GARG AND SIVA SANKAR SANA	
	13.1 Introduction	294
	13.2 Synthesis of nanoparticles	294
	13.2.1 Top-down approach	294
	13.2.2 Bottom-up approach	295
	13.3 Classification of nanomaterials	296
	13.3.1 Based on source	296
	13.3.2 Based on dimension	296
	13.3.3 Based on chemical composition	297
	13.3.4 Based on toxicity	297

	13.4 Green nanotechnology	297
	13.5 Scheming of green nanomaterials	297
	13.5.1 Approaches for green nanomaterial synthesis	299
	13.6 Zinc oxide	300
	13.7 Applications of zinc oxide nanoparticles	301
	13.8 Biosynthesis of nanoparticles	303
	13.9 Bacterial synthesis of ZnO nanoparticles and its applications	304
	13.10 Conclusions	310
	References	310
14.	Environmental impact on toxicity of nanomaterials	315
	J. PRAKASH ARUL JOSE, LAITH A. YOUNUS, KESAVAN BHASKAR REDDY, SIVA SANKAR SANA, LEKSHMI GANGADHAR, TIANYU HOU, ARGHYA CHAKRAVORTY AND PREETAM BHARDWAJ	
	14.1 Introduction	316
	14.2 A brief walk to nanomaterials and their properties	318
	14.3 The history of nanomaterials and their creation	318
	14.4 Nanomaterial sources	320
	14.5 Types and classification of nanomaterials	321
	14.5.1 Nanomaterials-based categories	321
	14.6 Applications of nanoparticles	324
	14.6.1 Applications in drugs and medications	324
	14.6.2 Fabrication and materials applications	326
	14.6.3 Applications in the environment	326
	14.6.4 Applications in electronics	327
	14.6.5 Applications in energy harvesting	327
	14.6.6 Applications in mechanical industries	328
	14.7 Nanomaterial regulations	328
	14.8 Nanomaterials problems and risk valuation	329
	14.8.1 Nanomaterial toxicity	329

	14.8.2 Toxici	ty of nanoparticles	330
		al for interactions between nanoparticles and as sources and health effects of nanoparticles	331
	14.10 Mechanism	as of nanoparticle toxicity	332
	14.11 Nanopartic	les in living systems – the surface effects	336
	14.12 Toxicology	of nanoparticles	338
	14.13 Nanomater	ials of different substances and their toxicity	341
	14.14 Solving tox	ic problem	346
	14.15 Conclusion		346
	References		347
15.	Sustainable nan resource develo	otechnology for human pment	357
	DIPTI SINGH AND R	AGHVENDRA PRATAP SINGH	
	15.1 Introduction	C.	358
	15.2 The nano-ag	proparticles	358
	15.3 Nanotechno	logy for sustainable practice	359
	15.3.1 Chitos	an in crop production	359
		an prevents deficiency of micronutrient in d crops	360
	15.3.3 Chitos	an vector for gene delivery	360
		echnology to improve the water quality for table agriculture	361
	15.3.5 Nano-	oligodynamic metal particles	361
	15.3.6 Nanot	echnology for crop yield enhancement	362
	15.3.7 Applic	ations of nanotechnology in food industries	363
	15.4 Chitosan nanoparticles synthesis		365
	15.4.1 Ionotr	opic gelation	365
	15.4.2 Coace	rvation	365
	15.4.3 Copre	cipitation	366
	15.4.4 Micro	emulsion method	366

	15.4.5 Spray drying method	366
	15.5 How to load active principle into chitosan nanoparticles	367
	15.6 Function of chitosan nanoparticles	367
	15.7 Conclusion and future perspectives	367
	References	369
16.	Rationale and trends of applied nanotechnology	373
	RAGHVENDRA PRATAP SINGH, ALOK R. RAI, RAJSHREE B. JOTANIA, RATIRAM G. CHAUDHARY AND AHMED ABDALA	
	16.1 Introduction	373
	16.2 Rules and regulations for nanotechnology	374
	16.3 Global nanotechnology sectors	376
	16.3.1 Nanotechnology industry in the world	377
	16.4 Types of nanotechnology	379
	16.4.1 Materials nanotechnology	379
	16.4.2 Green nanotechnology	380
	16.5 Nanotechnology applications	383
	16.6 Societal acceptance of nanotechnology	385
	References	385

Index 391



List of contributors

Ahmed Abdala Chemical Engineering Program, Texas A&M University at Qatar, Doha, Qatar

Nalluri Abhishek Department of Materials Science and Engineering, Huazhong University of Science and Technology, Wuhan, Taiyuan, P.R. China

Pablo E. Antezana Universidad de Buenos Aires, Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET). Instituto de Química y Metabolismo del Fármaco (IQUIMEFA), Facultad de Farmacia y Bioquímica, Buenos Aires, Argentina

Mian Nabeel Anwar Department of Civil and Environmental Engineering, University of Alberta, Edmonton, AB, Canada

Preetam Bhardwaj School of Electronics Engineering, Vellore Institute of Technology University, Vellore, Tamil Nadu, India

Ganesh S. Bhusari Research and Development Division, Solar Industries India Limited, Nagpur-440023, Maharashtra, India

Ragini Chahande Department of Biochemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Shalini Chahande Department of Biochemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce. Kamptee, Maharashtra, India

Arghya Chakravorty School of Bio Sciences and Technology, Vellore Institute of Technology, Vellore, Tamil Nadu, India

Ratiram G. Chaudhary Post Graduate Department of Chemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Rupali Ramesh Chaudhary Department of Botany, Sant Gadge Maharaj College Hingna, Nagpur, Maharashtra, India

Prashant B. Chouke Post Graduate Department of Chemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

xvii

xviii List of contributors

Kanhaiya M. Dadure Department of Chemistry, Bajaj College of Science, Wardha, Maharashtra, India

Martin F. Desimone Universidad de Buenos Aires, Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET). Instituto de Química y Metabolismo del Fármaco (IQUIMEFA), Facultad de Farmacia y Bioquímica, Buenos Aires, Argentina

Vidyasagar Devthade Material Science and Engineering Department, Kyungpook National University, Daegu, Republic of Korea

Barun K. Dutta Department of Chemistry and Chemical Engineering, Xiamen University, Xiamen, P.R. China

Gayatri Gaikwad Department of Chemistry, Priyadarshini J. L. College of Engineering, Nagpur, Maharashtra, India

Lekshmi Gangadhar Department of Nanotechnology, NanoDot Research Private Limited, Nagercoil, Tamil Nadu, India

Amar P. Garg Shobhit Institute of Engineering & Technology, Meerut, Uttar Pradesh, India

Animeshchandra G.M. Haldar Department of Applied Chemistry, Priyadarshini Bhagwati College of Engineering, Nagpur, Maharashtra, India

Tianyu Hou School of Chemical Engineering and Technology, North University of China, Taiyuan, P.R. China

Prashant P. Ingale Department of Zoology, Saibaba College Parseoni, Nagpur, Maharashtra, India

Rashmi Jachak Department of Botany. Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Lellouche Jean-Paul Department of Chemistry & Institute of Nanotechnology & Advanced Materials (BINA), Bar-Ilan University, Israel

Shalini Jiwan Chahande Department of Biochemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Rajshree B. Jotania Department of Physics, Electronics and Space science, University school of sciences, Gujarat University, Ahmedabad, India Prerna Khagar Department of Chemistry, Visvesvaraya National Institute of Technology, Nagpur, Maharashtra, India

Suraj V. Kombe Post Graduate Department of Microbiology, S. K. Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Simpal Kumari Dr. Shakuntala Misra National Rehabilitation University, Lucknow, Uttar Pradesh, India

Trimurti L. Lambat Department of Chemistry, Manoharbhai Patel College of Arts, Commerce and Science, Deori, India

Zhi Feng Li State Key Laboratory of Microbial Technology, Shandong University, Jinan, P.R. China

Rupali Mahakhode Department of Botany, Shri Shivaji Science College, Congress Nagar, Nagpur, Maharashtra, India

Debarshi Kar Mahapatra Department of Pharmaceutical Chemistry, Dadasaheb Balpande College of Pharmacy, Nagpur, Maharashtra, India

Atul Maldhure Water Technology and Management Division, CSIR-National Environmental Engineering Research Institute, Nagpur, Maharashtra, India

Dhanraj T. Masram Department of Chemistry, University of Delhi, Delhi, India

Madakka Mekapogu Department of Biotechnology and Bioinformatics, Yogi Vemana University, Kadapa, Andhra Pradesh, India

Aniruddha Mondal Division of Materials Science, Department of Engineering Sciences and Mathematics, Lulea University of Technology, Lulea, Sweden

Sudip Mondal Post Graduate Department of Chemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Sofia Municoy Universidad de Buenos Aires, Consejo Nacional de Investigaciones Científicas y Técnicas (CONICET). Instituto de Química y Metabolismo del Fármaco (IQUIMEFA), Facultad de Farmacia y Bioquímica, Buenos Aires, Argentina

Pentakota Surya Nagendra Department of Marine Engineering, Andhra University College of Engineering, Visakhapatnam, Andhra Pradesh, India

xx List of contributors

Manjiri S. Nagmote Post Graduate Department of Chemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Pallavi Pantawane Dr. Ambedkar College, Nagpur, Maharashtra, India

Ajay K. Potbhare Post Graduate Department of Chemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

J. Prakash Arul Jose Department of Civil Engineering, Paavai Engineering College, Namakkal, Tamil Nadu, India

V.V.S. Prasad Department of Marine Engineering, Andhra University, Visakhapatnam, Andhra Pradesh, India

Umesh Pratap Department of Chemistry, Visvesvaraya National Institute of Technology, Nagpur, Maharashtra, India

Alok R. Ral Department of Microbiology, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India

Kesavan Bhaskar Reddy Centre for Pharmaceutical Nanotechnology, Sri Venkateswra College of Pharmacy, Chittoor, Andhra Pradesh, India

Siva Sankar Sana School of Chemical Engineering and Technology, North University of China, Taiyuan, Taiyuan, P.R. China

Ratnnadeep C. Sawant Department of Chemistry, Dr. Ambedkar College Deekshabhoomi, Nagpur, Maharashtra, India

Trupti S. Shrirame Post Graduate Department of Chemistry, Seth Kesarimal Porwal College of Arts and Science and Commerce, Kamptee, Maharashtra, India: Department of Chemistry, Shri Shivaji Science and Arts College, Chikhli, India

A.K. Singh Department of Chemistry, Govt. VYTPG Autonomous College, Durg, Chhattisgarh, India

Dipti Singh Department of Microbiology, VBS Purvanchal University, Jaunpur, India

N.B. Singh Department of Chemistry and Biochemistry, SBSR and, Research Development Cell, Sharda University, Greater Noida, Uttar Pradesh, India



Preface

With the emergence of interest in the development of eco-friendly, green-synthesized, costeffective, and straightforward methods, the nanotechnology has emerged as a foremost research topic that has glimmered extensive curiosity. Nanotechnology signifies a radical path for high-tech expansion of living as well as nonliving material at the nanometer scale (reduced to billion times from a meter).

It comprehends the assembly and application of biological, chemical, and physical systems ranging from single atoms or molecules to submicron dimensions, and further an active and sustainable integration of it into the larger systems.

The driving force of this interest is the possibility to avoid hazardous chemicals and lower energy consumption. In green synthesis methods different materials from biological origins such as microorganism, cells, plants, or their enzymes or extract are employed with the focus on a greener environment minimizing waste generation and implementing sustainable processes. The green synthesis of nanostructured material is an extremely challenging approaches for all the researchers due to the existence of phytochemical agents in the extracts such as various sugars, flavones, saponins, proteins, amino acids, chromone, steroids, phytol, and terpenoids. The phytochemicals present in the plant extracts play a key role in the improvement of reduction rate, size, and stabilization by acting as good reducing surfactants and structure directing and capping agents. Therefore this themed issue is a compilation of articles reviewing the green synthesis of nanomaterials with various biological systems, especially the emphasis is placed on the mechanisms of nanomaterial synthesis, spectroscopic characterization, and their applications in different fields. These synthesized nanomaterials have wide potentiality to change our perceptions and expectations by providing us the competence to resolve comprehensive mankind issues.

Nowadays, nanotechnology is a vital part of our research and reality. A sundry of domains of human resource activities has reached from food production to its conservation, medicine production to its efficient application, biology to biotechnology, computing to communications, transport to space investigation. The chemical mediated nanotechnology inevitably contributes to hazardous ecological contamination. Hence, the editors have planned to upsurge the facts and flaws of nanotechnology and tried to display the radical solution with the compilation of global intellectual ideas in a book.

This book expresses the cumulative efforts of handpicked specific scientist and professors having expertise in nanotechnology, biotechnology, and microbiology and the noble effort of them has been summarized as the biogenic nanotechnology or green nanomaterials and its related areas.

The authored chapters precisely touch upon facets such as microbial factory-based environment-friendly nanosynthesis and their characterization and the use of various biological items such as plants and its part, bacteria, fungi, etc. to synthesize an efficient and high functional materials compared to traditional chemical synthesis of nanoparticles. Moreover, the historical backgrounds and future prospectives have also been elaborated nicely and finally the economic aids of green nanotechnology to human resource development have been shown.

Editors are highly grateful to all the eminent authors for their outstanding contribution, who are to be recognized not only for their contribution but, most notably, for keeping their promise timely which has facilitated us to finalize this project in the given timeline.

We would also like to thank Cloe Holland-Borosh, Editorial Project Manager at Elsevier, who fortified us to commence editing of the present project; Rachel Pomery, Editorial Project Manager; Emily loy Grace Thomson, Senior Editorial Project Manager of the book project; and the entire Elsevier team participated into its publication, for their firm work, skillful handling, sustenance, and patience. Finally, we thank the researchers and readers, who perceived this book, read, and found it motivating for nanoresearch. Editors are always welcome for any type of comments regarding its content and presentation manners.

> Raghvendra Pratap Singh Alok R. Rai Ratiram G. Chaudhary Ahmed Abdala



ScienceDirect



Biogenic Sustainable Nanotechnology

Trends and Progress

A volume in Micro and Nano Technologies

Book • 2022 Edited by: Raghvendra Pratap Singh, Alok R. Rai, ... Ratiram G. Chaudhary

Browse book content

About the book

Search in this book

Search in this book

Table of contents

Full text access
 Title page, Copyright, Contents, List of contributors, Preface

Book chapter \circ Abstract only 1 - Building nanomaterials with microbial factories

Pablo E. Antezana, Sofia Municoy and Martin F. Desimone Pages 1-39

🍸 Purchase 🛛 View abstract 🗸

Book chapter O Abstract only

2 - Phytofabrication of nickel-based nanoparticles: focus on environmental benign technology and therapeutic perspectives

Kanhaiya M. Dadure, Debarshi Kar Mahapatra, ... Ajay K. Potbhare Pages 41-57

🍸 Purchase 🛛 View abstract 🗸

Book chapter • Abstract only 3 - Bacterial cellular mechanisms for synthesis of green nanostructured compounds

FEEDBACK 🖵

Simpal Kumari, Zhi Feng Li and Mian Nabeel Anwar Pages 59-76

🕑 Purchase 🛛 View abstract 🗸

Book chapter O Abstract only

4 - Ecofriendly microorganism assisted fabrication of metal nanoparticles and their applications

Sudip Mondal, Manjiri S. Nagmote, ... Aniruddha Mondal Pages 77-105

🍸 Purchase 🛛 View abstract 🗸

Book chapter O Abstract only

5 - Herbal spices and nanotechnology for the benefit of human health

Shalini Jiwan Chahande, Rashmi Jachak, ... Pallavi Pantawane Pages 107-129

🕑 Purchase 🛛 View abstract 🗸

Book chapter O Abstract only

6 - Nanoparticles for sustainable agriculture: innovative potential with current and future perspectives

Subhash Rupchand Somkuwar, Ratnnadeep C. Sawant, ... Rupali Ramesh Chaudhary Pages 131-148

Purchase View abstract 🗸

Book chapter O Abstract only

7 - Fabrications and applications of polymer–graphene nanocomposites for sustainability

Ajay K. Potbhare, Trupti S. Shrirame, ... Ganesh S. Bhusari Pages 149-184

🕑 Purchase 🛛 View abstract 🗸

Book chapter O Abstract only

8 - Phytofabrication of metal oxide/iron-based and their therapeutic and their therapeutic potentials: in-depth insights into the recent progress

Animeshchandra G.M. Haldar, Debarshi Kar Mahapatra, ... Ratiram G. Chaudhary Pages 185-216

🍸 Purchase 🛛 View abstract 🗸

Book chapter O Abstract only

9 - Highlights of decade long progress of nano-selenium fabricated from plant biomass: insights into techniques and mechanisms

Debarshi Kar Mahapatra, Animeshchandra G.M. Haldar and Kanhaiya M. Dadure Pages 217-226

🍸 Purchase 🛛 View abstract 🗸

Book chapter O Abstract only

02/12/2022, 08:38 Biogenic Sustainable Nanotechnology | ScienceDirect 10 - Strategies of nanotechnology as a defense system in plants Rashmi Jachak, Shalini Chahande, ... Rupali Mahakhode Pages 227-248 Purchase View abstract 🗸 Book chapter O Abstract only 11 - Nanocomposites for dye remediation from aqueous solutions N.B. Singh, N.P. Singh, ... Lellouche Jean-Paul Pages 249-266 Purchase View abstract 🗸 Book chapter O Abstract only 12 - Sustainable hybrid nanomaterials for environmental remediation and agricultural advancement Prerna Khagar, Sangesh Zodape, ... Atul Wankhade Pages 267-292 Purchase View abstract 🗸 Book chapter O Abstract only 13 - Bacterial synthesis of zinc oxide nanoparticles and their applications Lekshmi Gangadhar, Nalluri Abhishek, ... Siva Sankar Sana Pages 293-313 Purchase View abstract 🗸 Book chapter O Abstract only 14 - Environmental impact on toxicity of nanomaterials J. Prakash Arul Jose, Laith A. Younus, ... Preetam Bhardwaj Pages 315-355 Purchase View abstract 🗸 Book chapter O Abstract only 15 - Sustainable nanotechnology for human resource development Dipti Singh and Raghvendra Pratap Singh Pages 357-372 Purchase View abstract 🗸 Book chapter O Abstract only 16 - Rationale and trends of applied nanotechnology Raghvendra Pratap Singh, Alok R. Rai, ... Ahmed Abdala Pages 373-389 Purchase View abstract 🗸 Book chapter O Full text access

➡ Download PDF

About the book

Description

Biogenic Sustainable Nanotechnology: Trends and Progress focuses on the green synthesis of nanomaterials with various biological systems, emphasizing the mechanisms of nanomaterial synthesis, spectroscopic characterizations, and applications in a variety of industrial sectors.

Show more \checkmark

Key Features

Outlines the major synthesis methods used to create environmentally-friendly bionanomaterials for biomedical applications

Explores how environmentally-friendly bionanomaterials are used for a variety of industry sectors

Show more \checkmark

Details

ISBN

978-0-323-88535-5

Language

English

Published

2022

Copyright

Copyright © 2022 Elsevier Inc. All rights reserved.

Imprint

Elsevier

DOI

https://doi.org/10.1016/C2020-0-03506-1

You currently don't have access to this book, however you can purchase separate chapters directly from the table of contents or buy the full version.

Purchase the book ↗

Editors

Raghvendra Pratap Singh

Department of Research & Development, Biotechnology, Uttaranchal University, Dehradun, Uttarakhand, India

Alok R. Rai

Department of Microbiology, Seth Kesarimal Porwal College of Arts, Commerce, and Science, Nagpur, Maharashtra, India

Ahmed Abdala

Chemical Engineering Program, Texas A&M University at Qatar, Doha, Qatar

Ratiram G. Chaudhary

Department of Chemistry, Seth Kesarimal Porwal College of Arts, Science and Commerce, Kamptee, Maharashtra, India

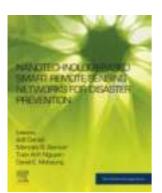
Related publications

(i) Why related?



Journal

Materials Today: Proceedings



Vijay L. Maheshwari Ravindra H. Patil *Editors*

Natural Products as Enzyme Inhibitors

An Industrial Perspective



Editors Vijay L. Maheshwari School of Life Sciences KBC North Maharashtra University Jalagon, Maharashtra, India

Ravindra H. Patil Department of Microbiology and Biotechnology R. C. Patel Arts, Commerce and Science College Shirpur, India

ISBN 978-981-19-0931-3 ISBN 978-981-19-0932-0 (eBook) https://doi.org/10.1007/978-981-19-0932-0

 ${\ensuremath{\mathbb C}}$ The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd. 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Singapore Pte Ltd. The registered company address is: 152 Beach Road, #21-01/04 Gateway East, Singapore 189721, Singapore

Contents

1	Fourth-Generation Allosteric EGFR Tyrosine Kinase Inhibitorsto Combat the Drug Resistance Associated with Non-smallCell Lung Cancer (NSCLC)Iqrar Ahmad, Rahul Pawara, Asama Pathan, and Harun Patel	1
2	Plant Peptides as Protease Inhibitors for Therapeutic andAgricultural ApplicationsRanjit S. Barbole, Nidhi Saikhedkar, and Ashok Giri	25
3	Bioactive α-Amylase Inhibitors: Sources, Mechanism of Action, Biochemical Characterization, and Applications Sainath S. Kasar, Vijay L. Maheshwari, and Pankaj K. Pawar	59
4	Recent Updates on In Silico Screening of Natural Products as Potential Inhibitors of Enzymes of Biomedical and Pharmaceutical Importance	105
5	H ⁺ /K ⁺ -ATPase Inhibitors from Plants: A Potential Source for Drug Discovery	125
6	Use of Protease Inhibitors as a Promising Alternative for Pest Control	137
7	Pancreatic Lipase (PL) Inhibitors from Medicinal Plants and Their Potential Applications in the Management of Obesity Samadhan Patil, Mohini Patil, Vijay L. Maheshwari, and Ravindra H. Patil	153

Contents

8	Bioactive Peptides and Polysaccharides: Setting a New Trendin Replacing Conventional Angiotensin-Converting EnzymeInhibitorsMuhammad Hakimin Shafie, Pei Gee Yap, and Chee-Yuen Gan	169
9	Natural Protease Inhibitors and Their Therapeutic PotentialsAgainst SARS-CoV-2Nilesh Chandrabhan Vadnere and Nitinkumar P. Patil	205
10	Telomerase and its Inhibitor in Cancer Therapeutics:Current Status and Future ProspectiveVivek Srivastava, Saleha Siddiqui, Akanksha Dhondiyal,Pakhi Gupta, and Ankush Yadav	227
11	Metal Nanomaterials as Enzyme Inhibitors and Their Applications in Agriculture and Pharmaceutics	251
12	$\alpha\text{-}Glucosidase$ Inhibitors for Diabetes/Blood Sugar Regulation Aditi Bhatnagar and Abha Mishra	269

Chapter 11 Metal Nanomaterials as Enzyme Inhibitors and Their Applications in Agriculture and Pharmaceutics



Satish V. Patil, Kiran R. Marathe, Hemant P. Borase, and Bhavana V. Mohite

Abstract As enzymes play a vital role in all biological systems, their regulation is also an important mechanism to drive various systems. In modern agriculture practices, the use of enzyme inhibitors to control pests and to regulate soil microbial activity is becoming an essential practice. The synthetic and natural organic chemicals are already documented for their use in pest control and fertilizers as a protectant. But these chemicals have reported some drawbacks like sensitivity to physical factors like temperature, pH, development of resistance and phytotoxicity, etc. The current review focuses on the potential and the use of metal nanomaterials as enzyme inhibitors in important agriculture practices. Various nanomaterials like lead, copper, gold, etc. were reported for their enzyme inhibition potential such as proteases, ureases, nitrate reductase, acetylcholine esterases, etc. Their potential with some conjugates as important agrochemicals such as pest control agents, fertilizers additives is briefly described in this review which will induce researchers for designing future agro formulations.

Keywords Enzyme inhibitors · Pests · Nanomaterials · Fertilizer additives · Agriculture practices

S. V. Patil $(\boxtimes) \cdot K$. R. Marathe

H. P. Borase

B. V. Mohite Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India

School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India

Department of Ophthalmology and Visual Sciences, University of Illinois, Chicago, Illinois, USA

11.1 Introduction

Enzymes are the biocatalyst of the living system. After nucleic acid, enzymes are considered to be the most vital biomolecules. Around 5000 biochemical reactions are carried out by enzymes (Ahmed et al. 2016). The biological functions of enzymes are diverse ranging from metabolism, nerve impulse transmission, control of biological activities such as replication, translation, organ specialization, blood coagulation, breathing, reproduction, generation of resistance, and diseases to name a few (Katsimpouras and Stephanopoulos 2021; Gomez-Fabra Gala and Vögtle 2021).

Owing to the essential role of enzymes not only in humans but also in microbes and insects, a class of molecules known as "enzyme inhibitors" are researched as a promising drug candidate in diabetes, Alzheimer's, and cancer and as a biocidal agent to kill plant pathogens and pests (Chen et al. 2017). Enzyme inhibitors act on enzymes because of the association of enzymes with life-supporting processes and diseases such as metabolic, neurological, cardiovascular, etc. Inhibition of enzymes can be lethal to pathogens, and therefore, enzyme inhibitors can be used as pesticides (Copland 2005). Enzyme inhibitor binds to an enzyme's active site and partially or completely inhibits its activity. However, they need not cover the complete binding site.

A variety of molecules such as paracetamol, rivastigmine, mupirocin, fosinopril, ritonavir, atorvastatin inhibit target enzymes such as acetylcholinesterase, cyclooxvgenase, HIV protease, HMG-CoA reductase and, therefore, are used in the treatment of bacterial and viral infections, pain, cancer, hypertension, hypercholesteremia, etc. (Copeland et al. 2007; Ouertani et al. 2019). However, most of the currently available enzyme inhibitors are organic and have several drawbacks such as instability, degradation, and low catalytic activity (Cha et al. 2015; Ouertani et al. 2019). Moreover, organic enzyme inhibitors work on relatively few major types of inhibitory mechanisms such as competitive, irreversible, or allosteric inhibition with several limitations (Ouertani et al. 2019). There is a need to find enzyme inhibitors having a broad inhibition spectrum.

Global pharma-biotech-agro industries such as Ranbaxy Laboratories, AstraZeneca, Merck, Pfizer, Roche are investing a big amount of money in enzyme inhibitor research (Cision 2017). The global market for enzyme inhibitors was estimated to be more than US\$ 95 billion in 2006 and is predicted to grow at an average rate of around 8% per annum (BCC Research 2018). Hence, there is a plenty of space to find versatile and robust enzyme inhibitors with unconventional chemical structures, desirable features, degradation resistance, and diverse inhibitory effects.

11.2 Potential of a Metal Nanomaterial as an Enzyme Inhibitor

According to the national nanotechnology initiative (United States), nanotechnology is science, engineering, and technology conducted at the nanoscale (1–100 nm) and utilizing products obtained from this unique phenomenon in diverse fields from chemistry and physics to medicine and engineering (NNI 2020). Pieces of evidence of the use of nanomaterials by humans trace back to the fourth century AD in the time of Greek and Democritus and can be found in historical objects like the Lycurgus cup and medieval church windows (Bayda et al. 2020). Nanotechnology is holding great economical potential as evidenced by its tremendous industrial applications and incorporation in day-to-day consumer goods (Janković and Plata 2019). Nanoforms of silver, gold, copper, zinc, and other metals are successfully applied in the pharmaceutical and agriculture industry (Khan et al. 2019; Singh et al. 2021).

There are two approaches for the synthesis of metal nanoparticles, viz. top-down and bottom-up. Extensive research was done on chemical, physical, and biological methods of nanoparticles synthesis, but the growing demand for nanomaterials is fulfilled by chemical and physical methods despite growing toxicity issues. On the other hand, biological methods utilizing microbes and plants present a promising approach but are still not feasible for large-scale nano synthesis (Patil et al. 2016; Borase et al. 2014, 2021; Ahmed Mohamed et al. 2020).

Pharmaceutics and agriculture are expected to gain tremendous benefits from nanotechnology interventions (Singh et al. 2021; Contera et al. 2020). Current agricultural practices in most of the world rely on high doses of agrochemicals such as fertilizers, pesticides that adversely affect soil rhizospheric microbiome, causing water pollution and biomagnification, affecting food quality and supply (Singh et al. 2021).

Plant growth is largely affected by various biotic and abiotic factors such as disease-causing pathogens, genetic traits, moisture availability, and soil fertility (Lahiani et al. 2013). Nanoscience is an innovative platform that involves the development of approaches to a range of inexpensive nanotech applications for enhanced seed germination, plant growth, development, and acclimation to environments. In this regard, an extensive number of studies have shown that the application of nanomaterials has positive effects on germination as well as plant growth and development. Likely, the application of multiwalled carbon nanotubes (MWCNTs) positively influences seed germination of different crop species including tomato, corn, soybean, barley, wheat, maize, peanut, and garlic (Khodakovskaya and Biris 2010; Srivastava and Rao 2014). Nanoparticles as enzyme inhibitors are promising due to several properties such as the high surface area to mass ratio, nano size, different shape, chemical functionalization, resistance to degradation in environmental conditions, etc. (Maccormack et al. 2012; Chen et al. 2017). Nanoparticles are expected to act as broad-spectrum enzyme inhibitors due to the above characteristics (Benelli 2018).

11.3 Nanoparticles Inhibiting Vital Enzymes (Some Examples)

Most of the commercially available antibiotics and agrochemicals act on pathogens and insects by targeting important enzymes. However, due to the widespread issue of resistance emergence, nanoparticles offer a promising alternative to the traditional arsenal of enzyme inhibitors (Ahmed et al. 2016; Ali et al. 2018). Nanoparticles have useful properties including shape, size tenability, binding of multiple ligands on the surface, and diverse enzyme inhibitory strategies.

Penicillin inhibits enzyme transpeptidase, which is essential for bacterial cell wall synthesis. Thus, the β -lactam-mediated inhibition of transpeptidation leads to cell lysis. However, after the introduction of penicillin, within a few years, various bacterial strains started showing drug resistance by naturally producing an enzyme, penicillinase (Drawz et al. 2014).

Two-dimensional molybdenum disulfide (2D-MoS₂) nanomaterials were reported to inhibit the β -lactamase enzyme. The negatively charged ligand functionalized MoS₂ materials exhibit electrostatic and other non-covalent interactions between enzyme and inhibitor leading to competitive inhibition of the latter (Ali et al. 2018).

Trypsin (223 amino acid residue protein) is another important target enzyme involved in human diseases and also helps insects to neutralize biocidal agrochemicals (Schnebli and Braun 1986). It was observed that absorption of trypsin on TiO₂ NPs decreased the β sheet content. TiO₂ NP-trypsin interaction altered the secondary structure due to electrostatic force, van der Waals force, and hydrogen bonding (Wang et al. 2011). In another study, gold nanoparticles were also found to inhibit trypsin. Electrostatic and hydrophobic interactions, along with covalent interactions between Cys-58, 42, Lys-60, and gold nanoparticles, might be involved in the binding and inhibition process (Zhang et al. 2014).

Urease (nickel-containing metalloenzymes) is an important factor in peptic ulcers, and commercially available urease inhibitors have multiple side effects such as antibiotic resistance (Naz et al. 2019). Silver nanoparticles functionalized with N-substituted methyl 5-acetamido- β -resorcylate (AgL) with an average size of 20 nm were found to be stable at variable temperatures and pH. AgL showed enhanced urease inhibition as compared to the standard drug (thiourea), N-substituted methyl 5-acetamido- β -resorcylate, and silver nanoparticles. Interestingly, AgL is inactive against other metalloenzymes like xanthine oxidase and carbonic anhydrase II as well as for non-metallic enzymes such as α -chymotrypsin and acetylcholinesterase. Therefore, AgL is very selective to inhibit urease only (Benelli 2018).

Silver nanoparticles fabricated using *Cassia fistula* fruit pulp extract inhibits the fourth instar larvae of *Aedes albopictus* and *Culex pipienspallens* (Coquilett) with a substantial decrease of acetylcholine esterase and α - and β -carboxylesterase activities. Similar to the above study, silver nanoparticles prepared using salicylic acid and

3,5-dinitrosalicylic acid inhibit *Ae. albopictus*, with the decrease of total proteins, esterase, acetylcholine esterase, and phosphatase enzymes.

Multivalent nanoconstructs are another new advanced technology involving the conjugation of multiple copies of enzyme inhibitors on nanosurface to improve inhibitory potency and selectivity. Nanometer-sized fluorescent hybrid silica (NFHS) particles (size 150 nm) loaded with fluorescent and sulfonamide carbonic anhydrase inhibit carbonic anhydrase activity several-fold because of higher silica nanoparticles adsorption on the border of carbonic anhydrase (Touisni et al. 2015).

Detoxification enzymes (GST, Catalase, SOD) of insects act as the first line of defense against chemical-induced stress and hence are an attractive target for designing insecticidal and pesticide formulations. Several other enzymes (aspartate aminotransferase (AST) and alanine aminotransferase (ALT)) serve an important role in protein and carbohydrate metabolism (Borase et al. 2021). Insects characteristically lack important detoxification enzymes glutathione peroxidase and catalase known to have little affinity with hydrogen peroxide (H₂O₂). Hence, ascorbate peroxidase (APOX) plays a major role in the clearance of (H₂O₂). Beetles species *Blapspolychresta* were treated with 26.27 \pm 4.43 nm Nickel (II) oxide nanoparticles (NiO NPs) at sub-lethal concentrations of 0.02 mg/g. NiO NPs were found to cause a significant decrease in the activity of APOX as compared to the untreated group (El-Ashram et al. 2021).

Borase et al. (2019, 2020, 2021) evaluated the effects of the most widely used metal nanoparticles (silver nanoparticles (20 and 40 nm), gold nanoparticles (30 nm), and zinc oxide particles (250 and 500 nm)) on important enzymes of aquatic organism *Moina macrocopa*. Interestingly all nanoparticles under investigation inhibit key enzymes including acetylcholinesterase and digestive enzymes (trypsin, amylase) and β -galactosidase. It was found that the size of nanoparticles is a crucial factor during their interaction with enzymes along with the resistance/ sensitivity of the organism. Although *M. macrocopa* is an aquatic organism but finding from the above studies can be a reference for developing enzyme inhibitors for pharma and agriculture sectors.

11.4 Future of Nanoparticles as an Enzyme Inhibitor

Although several studies put forward the superiority of metal nanoparticles as a new class of enzyme inhibitor (BCC Research 2018), more research is needed to study the complete life cycle of nanomaterials including nonspecific interactions with other proteins normally present in biological fluid. Toxicity and accumulation in non-target host and environmental consequences are also unanswered questions.

11.4.1 Nanoparticles as Protease Inhibitors in Pest Management

Nanoparticles have various applications in different fields of agriculture like pest management, vector-pest management, herbicide delivery, and nanosensors for pest detection (Scrinis and Lyons 2007; Rahman et al. 2009). The role of nanoparticles as a protease inhibitor, their mechanism of action in insects, and their possible applications in agriculture are summarized below.

Proteases or peptidases are hydrolytic enzymes that selectively catalyze the cleavage of peptide bonds in proteins. Peptidases participate in various cellular physiological processes and irreversible proteolytic reactions whose control is essential for cell functions. The proteolytic activity of proteases is controlled by regulating secretions, specific degradation, and also by inhibition. Several natural specific and selective protease inhibitors are now known as major regulating proteins to control proteolytic activity in all life forms (Umezawa 1982). These characteristics make protease inhibitors good diagnostic and therapeutic tools for the treatment of various microbial (hepatitis, herpes, AIDS, aspergillosis) and mortal (arthritis, muscular dystrophy, malaria, cancer, obesity), neurodegenerative, and cardiovascular diseases (Karthik et al. 2014). Until now, several protease inhibitors are identified from plants, animals, and microbes for each mechanistic class of proteases, e.g., serine, cysteine, aspartyl, and metalloproteases (Lorito et al. 1994; Joshi et al. 1999; Bijina et al. 2011). Although the role of protease inhibitors have been identified, they are not widely used in agriculture due to ability of insects to produce insensitive proteases or degradation of inhibitors to neutralize their effects. Hence, there is the necessity to look for alternative solutions to overcome protease inhibitor resistance developed by insects. With the emergence of widespread antibiotic, enzyme, and insecticides resistance, a better alternative is the use of different metal nanoparticles. Nanoparticles possess several advantages over conventional protease inhibitors in respect of their stability, reproducibility, and reactivity with other chemicals (Friedman et al. 1993). The different biologically synthesized metal nanoparticles, their size, applications, and enzyme inhibition pattern are summarized in Table 11.1.

The biosynthesized chromium nanoparticles (Cr_2O_3 NPs) from *Hyphaene thebaica* showed antiviral activity against the poliovirus by inhibiting the protein kinase enzymes (Khalil et al. 2018). Similarly, Iqbal et al. (2020) reported the Cr_2O_3 NPs from *Rhamnus virgata* leaves extract showed the inhibition of protein kinase and α -amylase enzymes with biopotential against the fungi and bacterial species. The silver, aluminum oxide, zinc oxide, and titanium dioxide NPs have been successfully used as insecticides to manage different pests, most of these NPs showed detrimental effects on treated insects by inhibiting the gut proteases enzymes (Table 11.1). Of these, silver nanoparticles are the most studied and utilized nanoparticles for biosystem due to their strong inhibitory and bactericidal effects as well as a broad spectrum of antimicrobial activities (Becker et al. 2005). Feng et al. (2000) reported that silver ions interact strongly with the thiol groups of vital enzymes, causing their inactivation. Patil et al. (2016) observed the

Tablé	Table 11.1 Some examples of metal nand	particles a	es of metal nanoparticles as an enzyme inhibitor	lbitor		
Sr.		Size				
No.	Metal nanoparticles	(um)	Shape	Target enzyme	Major finding	Reference
	Gold nanoparticles (AuNPs)	20-50	Triangular, hexagonal	Protease	Serum protease of A. aegypt, H. armigera, C. maculatus inhibited due to multiple bonding between enzyme and AuNPs	Patil et al. (2016)
5	Zinc oxide nanoparticles (ZnO NPs)	20	Pyramids, plates, and spheres	β-Galactosidase	Shape-dependent denaturation, electrostatic attraction	Cha et al. (2015)
б	Silica nanoparticles	4, 20 and 100	Spherical	Chicken egg lysozyme	Loss in alpha-helix content. Strong adsorp- tion for large NPs	Vertegel et al. (2004)
4	Copper oxide nanoparticles	<50	Spherical	Nitrate reductase and nitric oxide reductase	Nitrate reductase and nitric Inhibition of electron transport chain oxide reductase	Zhao et al. (2020)
S	Silver nanoparticles (AgNPs)	10	I	Urease, acid phosphatase, arylsulfatase, β-glucosidase	AgNPs negatively affect soil exoenzyme activities, with the urease activity especially sensitive to AgNPs due to non-competitive inhibitor by blocking sulfhydryl groups	Shin et al. (2012)
9	Silver nanoparticles (AgNPs)	20	Spherical	Catalase (CAT) and superoxide dismutase	Disturbing α -helicalsecondary structure of enzymes	Liu et al. (2020)
٢	Silver-moxifloxacin nanoparticles	50-60	Spherical	Urease	250 times better compared to moxifloxacin	Nisar et al. (2015)
×	Two-dimensional molybdenumdisulfide (2D-MoS2) nanomaterials	I		β-Lactamase	Electrostatic, non-covalent interactions and steric obstruction. Useful in β -lactamase resistance	Ali et al. (2018)
						(continued)

(continued)	
11.1	
Fable	

Table	Table 11.1 (continued)					
Sr. No.	Sr. Metal nanoparticles	Size (nm)	Shape	Target enzyme	Major finding	Reference
6	Lead oxide (PbO) nanoparticles	27		Acetylcholinesterase	Potential use in the treatment of Alzheimer's Khalil et al. (2018)	Khalil et al. (2018)
10	10 Gold nanoparticles (AuNPs)	5-70	5–70 Triangular	Cytochrome P450	AuNPs likely block the substrate pocket on Ye et al. (2014) the CYP surface	Ye et al. (2014)

protease-inhibiting properties of latex fabricated AuNPs, particularly with regard to trypsin when mixed with insect serum. Protease activity was found to be inhibited by 66% in the serum of A. aegypti mosquito larvae and pests including Helicoverpa armigera, C. maculatus, C. chinensis, and M. hirsutus. Kantrao et al. (2017) also reported the inhibition of gut protease activity of H. armigera due to biosynthesized AgNPs. Raj et al. (2017) reported that increased oral dosages of AgNPs in the larval stage of *Drosophila melanogaster* alter the protein, carbohydrate, and lipid levels by impairing the metabolic activity, which may be due to the inhibition of gut proteases. Similarly, the silver nanoparticles lower the activity of copper-dependent superoxide dismutase and tyrosinase enzymes in Drosophila melanogaster which is involved in the antioxidant activity and pigment production, respectively (Posgai et al. 2011; Armstrong et al. 2013; Ávalos et al. 2015). The biosynthesized AgNPs using the fruits of *Cassia fistula* showed a significant decrease in the activity of AChE, a-and b-carboxylesterases, phosphatase enzymes, and total proteins in Aedes albopictus and Culex pipienspallens larvae and pupa (Fouad et al. 2018). Yasur and Rani (2015) also reported the insecticidal activity of AgNPs against the Spodoptera litura F. and Achaea janata L. due to the accumulation of nanoparticles in the gut of larvae which decreases larval and pupal weight. Also, the conjugated silica nanoparticles with plant protease inhibitors showed excellent larvicidal activity against the Helicoverpa armigera by inhibiting gut proteinase activity (Bapat et al. 2020; Khandelwal et al. 2015). Marathe et al. (2021) also investigated the mode of action of AgNPs and suggested that antifungal activity could be due to inhibition of ergosterol biosynthesis leading to severe damage to the cell membrane. A summary of the effects of different metal nanoparticles on various pests is presented in Table 11.2.

11.4.2 Nano Metals as Urease Inhibitors

Urease is the nickel metalloenzyme produced by various microbes such as bacteria and fungi, which are the major component of soil. They are the key component of urea loss by gaseous ammonia and other minerals of nitrogen. Although very efficient urea inhibitors are available, they have some negative impacts on crop health, i.e., phytotoxicity and get rapidly hydrolyzed by the other soil microbes. They also lose their urease inhibiting potential by various environmental factors. Leaf scorches and necrosis of leaf margin are common examples of phytotoxicity of these synthetic inhibitors. It was also proved that N-(n-butyl) thiophosphoric triamide (NBPT), taken by peas and spinach roots and translocated to leaves, also inhibits endogenous urease of roots and leaves plants or crops (Artola et al. 2011). The glutamate synthetase and amino acid levels were found to be reduced by NBPT (Cruchaga et al. 2011).

These drawbacks led the scientific community to find other nonorganic urease inhibitors. Various metals have already been identified to possess potent urease inhibition potential. Some already identified metals (silver and mercury) forming insoluble sulfides act as potent urease inhibitors by reacting with sulfhydryl groups.

Sr. No.	Nanoparticles	Size (nm)	Application	Mechanism of action	References
1	Cr ₂ O ₃ -NPs	25–38	Antiviral, antioxidant	Inhibits the protein kinases in polio virus	Khalil et al. (2018)
2	Cr ₂ O ₃ -NPs	28	Antibacterial, antioxidant, antifungal	Inhibition of protein kinase and alpha amylases	Iqbal et al. (2020)
3	AgNPs synthe- sized with Ficus extracts	20	Insecticidal activity against <i>H. armigera</i>	Inhibits larval gut pro- teases of <i>H. armigera</i>	Kantrao et al. (2017)
4	AgNPs	20–100	Insecticidal activity against Drosophila melanogaster	Altered the protein, car- bohydrate, lipid metabolism	Raj et al. (2017)
5	AgNPs uncoated and polysaccharide- coated	10 and 60	Insecticidal activity against Drosophila melanogaster	Depigmentation by inhibiting the tyrosinase enzymes	Posgai et al. (2011)
6	AgNPs	10–50	Insecticidal activity against Drosophila melanogaster	Depigmentation, impaired movement, compromised fertility	Armstrong et al. (2013)
7	AgNPs	4.7	Insecticidal activity against Drosophila melanogaster	Depigmentation, impaired movement, compromised fertility	Ávalos et al (2015)
8	AgNPs PVP coated	-	Insecticidal activity against <i>Spodoptera</i> <i>litura</i> F. and <i>Achaea janata</i> L.	Larval and pupal weight decreased due to the accumulation of nanoparticles in gut	Yasur and Rani (2015)
9	AgNPs synthe- sized with Cas- sia fistula fruit	148–900	Aedes albopictus and Culex pipienspallens larvae and pupa	Inhibition of acetylcho- linesterase and a- and b-carboxyl esterases activity	Fouad et al. (2018)
10	AgNPs synthe- sized with <i>Streptomyces</i> spp.	20–40	Antifungal activ- ity against <i>fusar-</i> <i>ium verticilloides</i>	Inhibition of ergosterol biosynthesis and mem- brane damage	Marathe et al. (2021)
11	AuNPs synthe- sized with plant latex J. gossypifolia	20	Insecticidal activity against <i>H. armigera and</i> <i>A. aegypti</i>	Inhibition of larval gut protease	Patil et al. (2016)
11	SiNPs conju- gated with soyabean tryp- sin inhibitor (SiNPs-STI)	20–100	Insecticidal activity against <i>H. armigera</i>	Inhibition of gut pro- teinase activity (HGP)	Bapat et al. 2020)

 Table 11.2
 A summary of effect of metal nanoparticles against different pest

(continued)

Sr.		Size			
No.	Nanoparticles	(nm)	Application	Mechanism of action	References
12	Silica nanoparticles conjugated with plant pro- tease inhibitors	240	Insecticidal activity against <i>H. armigera</i>	Inhibition of gut pro- teinase activity	Khandelwal et al. (2015)
13	Copper nanoparticles (CuNPs)	10–70	Insecticidal activity against Tribolium castaneu	Body deformation throughout the life cycle of insect	El-Saadony et al. (2020)
14	Copper oxide nanoparticles (CuO NPs)	10–70	Insecticidal activity against Spodoptera littoralis	Reduced pupation and adult emergence, adult malformation, adult fecundity, and egg hatchability	Shaker et al. (2016)
15	Zinc nanoparticles (ZnO NPs)	25–50	Insecticidal activity against Spodoptera frugiperda	Effect on fecundity, fer- tility and longevity of insect	Pittarate et al. (2021)
16	ZnO nanoparticles (ZnO NPs)	50-60	Insecticidal activity against Aedes aegypti	Programmed cell death (apoptosis)	Banumathi et al. (2017) Gunathilaka et al. (2021)
17	Silica nanoparticles (SiO ₂ NPs)	50–150	Insecticidal activity against <i>Bombus</i> <i>terrestris</i>	Midgut epithelial injury	Mommaerts et al. (2012)
18	Graphene oxide nanoparticles	1–10	Insecticidal activity against Acheta domesticus	Increased enzymatic activity of catalase and glutathione peroxidases, as well as heat shock protein (HSP 70) and total antioxidant capac- ity level	Dziewięcka et al. (2016)
19	TiO ₂ nanoparticles	<100	Insecticidal activity against Bombyx mori	Upregulation of signal- ing pathway and downregulation of development and molting period	Li et al. (2014)
20	Alumina nanoparticle	30-60	Insecticidal activity against <i>Sitophilus oryzae</i> (L.)	Binding to beetle cuticle resulting in insect dehydration	Stadler et al. (2017)

 Table 11.2 (continued)

The general potential of metal ions as urease inhibitors is in the order of $Ag^{2+} > Hg^{2+}$ $Cu^{2+} > Ni^{2+} > CO^{2+} > Fe^{2+} > Mn^{2+}$ (Upadhyay 2012). These data may lead to the use of the above metals/non-metals as urease inhibitors. Although no commercial use of nanomaterial as urease inhibitor for soil application is available as yet, but

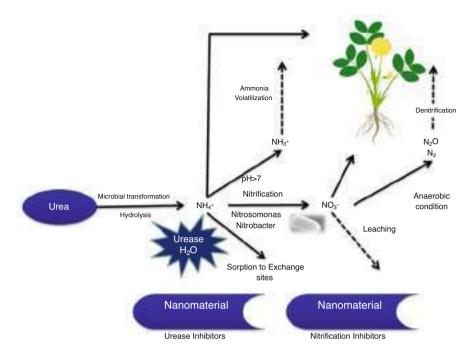


Fig. 11.1 Possible mechanism of urease regulations by metal nanomaterials

various reports of these metal nanomaterials as urease inhibitors make them potential candidates as the future component of nitrogenous fertilizers for controlled release of nitrogen, i.e., ammonia as per the requirement of the crop by controlling urease activity (Fig. 11.1).

Some reports on the use of nanometals as urease inhibitors also indicate their future potential. It was documented that PVP(polyvinyl alcohol)-based silver nanoparticles (AgNPs) having 15 nm size inhibiting the ammonia-oxidizing capacity of soil bacteria after direct application in soil (Huang et al. 2018). Similarly, Vandevoort and Arai (2018) and Simonin et al. (2018) found that copper nanometals, less than 50 nm in size, significantly decreased conversions of gaseous ammonia in nitrogen cycling.

Size-dependent reduction in ammonia oxidation potential of Nitrosomonas eutropea by zero-valent silver polyvinyl alcohol (PVA) composite was also reported (Yan et al. 2020). Yuan et al. (2013) revealed that Na₂ ATP-doped silver nanoparticle composite retains 58.2% AgNPs after 24 h while PVA-coated AgNPs remain or retain 9.9%; hence, the latter is more efficient in inhibiting NH₃ oxidation. It was also found that this reduction may be due to cell wall damage and disintegration of nuclei. Silver nanomaterial with 20 nm size functionalized with N-substituted methyl acetoamide B-resocyclate was found to be more stable at the wide range of pH, temperature, and salt concentration with significantly increased urease inhibition. It was also proved that Ag-5-Amino-β-resorcylic acid hydrochloride dihydrate (AR) had significantly greater urease inhibition than the thiourea. The observed inhibitory activity of the latter was recorded at 11–18 times lower concentration than the former. Similarly, silver nanoconjugates of 5-amino-beta-resorcylic acid hydrochloride dihydrate were found to possess significant in vitro inhibitory potential against important enzymes like urease, xanthin oxidases, cholinesterase, and chymotrypsin which are targeted in various nitrogen fertilizer protection and agricultural pest management (Naz et al. 2014).

Zheng et al. (2017) reported that AgNPs inhibit the nitrification rate which was found to be inversely related to the concentration of AgNPs. Borase et al. (2015) reported significant urease inhibition by plant latex-mediated AgNPs of 21 nm size. Similarly, Jadon et al. (2018) advocated urea coated with 4% neem, 4% pine oleoresins, 35% rock phosphate, and 2% nano zinc particles (ZnO) for reducing the ammonia volatilization by 27%, 41%, 26% and 35% respectively.

11.5 Conclusion

The present literature points to the enzyme inhibitory and entomologic properties of metal nanoparticles. Especially, the silver nanoparticles showed the maximum inhibitory activity against the gut proteases, acetylcholine esterase, and phosphatases. This review suggests the possible use of metal nanoparticles as a biocontrol agent in agriculture, which can withstand alkaline conditions of an insect's gut. Moreover, with the commencement of insecticides and antibiotic resistance, nanoparticle-based enzyme inhibitory strategies could open up a new tool in therapeutics and agriculture as nano-insecticides in pest management and revolutionize insect and phytopathogen control which can benefit humans in the long run.

References

- Ahmed Mohamed HE, Afridi S, Khalil AT, Zohra T, Ali M, Alam MM, Ikram A, Shinwari ZK, Maaza M (2020) Phyto-fabricated Cr₂O₃ nanoparticle for multifunctional biomedical applications. Nanomedicine 15:1653–1669
- Ahmed KB, Raman T, Veerappan A (2016) Future prospects of antibacterial metal nanoparticles as enzyme inhibitor. Mater Sci Eng C 68:939–947
- Ali SR, Pandit S, De M (2018) 2D-MoS₂-based β-lactamase inhibitor for combination therapy against drug-resistant bacteria. ACS Appl Biol Mater 1(4):967–974
- Armstrong N, Ramamoorthy M, Lyon D, Jones K, Duttaroy A (2013) Mechanism of silver nanoparticles action on insect pigmentation reveals intervention of copper homeostasis. PLoS One 8:e53186
- Artola E, Cruchaga S, Ariz I, Moran JF, Garnica M, Houdusse F, Mina JM, Irigoyen I, Lasa B, Aparicio-Tejo PM (2011) Effect of N-(n-butyl) thiophosphorictriamide on urea metabolism and the assimilation of ammonium by *Triticum aestivum* L. Plant Growth Regul 63:73–79

- Ávalos A, Haza AI, Drosopoulou E, Mavragani-Tsipidou P, Morales P (2015) In vivo genotoxicity assessment of silver nanoparticles of different sizes by the somatic mutation and recombination test (SMART) on Drosophila. Food Chem Toxicol 85:114–119
- Banumathi B, Vaseeharan B, Ishwarya R, Govindarajan M, Alharbi NS, Kadaikunnan S, Khaled JM, Benelli G (2017) Toxicity of herbal extracts used in ethno-veterinary medicine and green encapsulated ZnO nanoparticles against Aedes aegypti and microbial pathogens. Parasitol Res 116:1637–1651
- Bapat G, Zinjarde S, Tamhane V (2020) Evaluation of silica nanoparticle mediated delivery of protease inhibitor in tomato plants and its effect on insect pest *Helicoverpa armigera*. Colloids Surf B Biointerfaces 193:111079
- Bayda S, Adeel M, Tuccinardi T, Cordani M, Rizzolio F (2020) The history of nanoscience and nanotechnology from chemical-physical applications to nanomedicine. Molecules 25:112
- BCC Research (2018). https://www.bccresearch.com/market-research/biotechnology/global-mar kets-for-enzyme-inhibitors-report.html
- Becker C, Pradhan A, Pakstis L, Pochan DJ, Shah SI (2005) Synthesis and antibacterial properties of silver nanoparticles. J Nanosci Nanotechnol 5(2):44–249
- Benelli G (2018) Mode of action of nanoparticles against insects. Environ Sci Pollut Res 25(13): 12329–12341
- Bijina B, Chellappan S, Basheer SM, Elyas KK, Bahkali AH, Chandrasekaran M (2011) Protease inhibitor from *Moringa oleifera* leaves: isolation, purification and characterization. Process Biochem 46:2291–2300
- Borase HP, Patil CD, Salunkhe RB, Suryawanshi RK, Salunke BK, Patil SV (2014) Mercury sensing and toxicity studies of novel latex fabricated silver nanoparticles. Bioprocess Biosyst Eng 37(11):2223–2233
- Borase HP, Salunkhe RB, Patil CD, Suryawanshi RK, Salunke BK, Wagh ND, Patil SV (2015) Innovative approach for urease inhibition by Ficus carica extract–fabricated silver nanoparticles: an in vitro study. Buitechnol Appl Biochem 62(6):780–784
- Borase HP, Patil SV, Singhal RS (2019) Moina macrocopa as a non-target aquatic organism for assessment of ecotoxicity of silver nanoparticles: effect of size. Chemosphere 1(219):713–723
- Borase HP, Muley AB, Patil SV, Singhal RS (2021) Enzymatic response of Moina macrocopa to different sized zinc oxide particles: an aquatic metal toxicology study. Environ Res 194:110609
- Cha SH, Hong J, McGuffie M, Yeom B, VanEpps JS, Kotov NA (2015) Shape-dependent biomimetic inhibition of enzyme by nanoparticles and their antibacterial activity. ACS Nano 9(9):9097–9105
- Chen M, Zeng G, Xu P, Lai C, Tang L (2017) How do enzymes 'meet' nanoparticles and nanomaterials? Trends Biochem Sci 42(11):914–930
- Cision (2017). https://www.prnewswire.com/news-releases/1799-billion-enzyme-inhibitors-mar kets-2022-300573064.html
- Contera S, Bernardino de la Serna J, Tetley TD (2020) Biotechnology, nanotechnology and medicine. Emerg Top Life Sci 4(6):551–554
- Copeland RA, Harpel MR, Tummino PJ (2007) Targeting enzyme inhibitors in drug discovery. Expert Opin Ther Targets 11(7):967–978
- Copland RA (2005) Evaluation of enzyme inhibitors in drug discovery: a guide for medicinal chemists and pharmacologists. Wiley, Hoboken
- Cruchaga S, Artola E, Lasa B, Ariz I, Irigoyen I, Moran JF, Aparicio-Tejo PM (2011) Short term physiological implications of NBPT application on the N metabolism of *Pisum sativum* and *Spinacea oleracea*. J Plant Physiol 168:329–336
- Drawz SM, Papp-Wallace KM, Bonomo RA (2014) New β-lactamase inhibitors: a therapeutic renaissance in an MDR world. Antimicrob Agents Chemother 58(4):1835–1846
- Dziewięcka M, Karpeta-Kaczmarek J, Augustyniak M, Majchrzycki Ł, Augustyniak-Jabłokow MA (2016) Evaluation of in vivo graphene oxide toxicity for Acheta domesticus in relation to nanomaterial purity and time passed from the exposure. J Hazard Mater 305:30–40

- El-Ashram S, Ali AM, Osman SE, Huang S, Shouman AM, Kheirallah DA (2021) Biochemical and histological alterations induced by nickel oxide nanoparticles in the ground beetle Blapspolychresta (Forskl, 1775) (Coleoptera: Tenebrionidae). PLoS One 16:1–21
- El-Saadony MT, El-Hack A, Mohamed E, Taha AE, Fouda MM, Ajarem JS, Elshaer N (2020) Ecofriendly synthesis and insecticidal application of copper nanoparticles against the storage pest Tribolium castaneum. Nanomaterials (Basel) 10(3):587
- Feng QL, Wu J, Chen GQ, Cui FZ, Kim TN, Kim JO (2000) A mechanistic study of the antibacterial effect of silver ions on Escherichia coli and Staphylococcus aureus. J Biomed Mater Res 52:662–668
- Fouad H, Hongjie L, Hosni D, Wei J, Abbas G, Ga'al H, Jianchu M (2018) Controlling Aedes albopictus and Culex pipienspallens using silver nanoparticles synthesized from aqueous extract of Cassia fistula fruit pulp and its mode of action. Artif Cells Nanomed Biotechnol 46:558–567
- Friedman SH, DeCamp DL, Sijbesma RP, Srdanov G, Wudl F, Kenyon GL (1993) Inhibition of the HIV-1 protease by fullerene derivatives: model building studies and experimental verification. J Am Chem Soc 115:6506–6509
- Gomez-Fabra Gala M, Vögtle FN (2021) Mitochondrial proteases in human diseases. FEBS Lett 595:1205–1222
- Gunathilaka UMTM, de Silva WAPP, Dunuweera SP, Rajapakse RMG (2021) Effect of morphology on larvicidal activity of chemically synthesized zinc oxide nanoparticles against mosquito vectors. RSC Adv 11(15):8857–8866
- Huang J, Chong CA, Runqing LI, Wenzhu GU (2018) Effects of silver nanoparticles on soil ammonia-oxidizing microorganisms under temperatures of 25 and 5°C. Pedosphere 28:607–616
- Iqbal J, Abbasi BA, Munir A, Uddin S, Kanwal S, Mahmood T (2020) Facile green synthesis approach for the production of chromium oxide nanoparticles and their different in vitro biological activities. Microsc Res Tech 83:706–719
- Jadon P, Selladurai R, Yadav SS, Coumar MV, Dotaniya ML, Singh AK, Bhadouriya J, Kundu S (2018) Volatilization and leaching losses of nitrogen from different coated urea fertilizers. J Soil Sci Plant Nutr 18(4):1036–1047
- Janković NZ, Plata DL (2019) Engineered nanomaterials in the context of global element cycles. Environ Sci Nano 6(9):2697–2711
- Joshi BN, Sainani MN, Bastawade KB, Deshpande VV, Gupta VS, Ranjekar PK (1999) Pearl millet cysteine protease inhibitor: evidence for the presence of two distinct sites responsible for antifungal and anti-feedent activities. Eur J Biochem 265:556–563
- Kantrao S, Ravindra MA, Akbar SM, Jayanthi PK, Venkataraman A (2017) Effect of biosynthesized silver nanoparticles on growth and development of Helicoverpa armigera (Lepidoptera: Noctuidae): interaction with midgut protease. J Asia Pac Entomol 20(2):583–589
- Karthik L, Kumar G, Keswani T, Bhattacharyya A, Chandar SS, Bhaskara Rao KV (2014) Protease inhibitors from marine actinobacteria as a potential source for antimalarial compound. PLoS One 9:1–13
- Katsimpouras C, Stephanopoulos G (2021) Enzymes in biotechnology: critical platform technologies for bioprocess development. Curr Opin Biotechnol 69:91–102
- Khalil AT, Ayaz M, Ovais M, Wadood A, Ali M, Shinwari ZK, Maaza M (2018) In vitro cholinesterase enzymes inhibitory potential and in silico molecular docking studies of biogenic metal oxides nanoparticles. Inorg Nano-Met Chem 48(9):441–448
- Khan I, Saeed K, Khan I (2019) Nanoparticles: properties, applications and toxicities. Arab J Chem 12(7):908–931
- Khandelwal N, Doke DS, Khandare JJ, Jawale PV, Biradar AV, Giri AP (2015) Bio-physical evaluation and in vivo delivery of plant proteinase inhibitor immobilized on silica nanospheres. Colloids Surf B Biointerfaces 130:84–92
- Khodakovskaya MV, Biris AS (2010) Method of using carbon nanotubes to affect seed germination and plant growth. 13/509,487. U.S. Patent Application. 2010

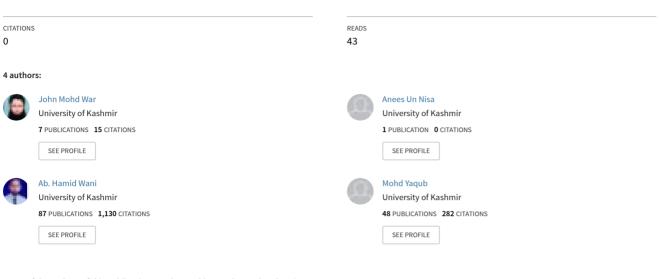
- Lahiani MH, Dervishi E, Chen J, Nima Z, Gaume A, Biris AS, Khodakovskaya MV (2013) Impact of carbon nanotube exposure to seeds of valuable crops. ACS Appl Mater Interfaces 5:7965– 7973
- Li F, Gu Z, Wang B, Xie Y, Ma L, Xu K, Ni M, Zhang H, Shen W, Li B (2014) Effects of the biosynthesis and signaling pathway of ecdysterone on silkworm (Bombyx mori) following exposure to titanium dioxide nanoparticles. J Chem Ecol 40:913–922
- Liu W, Worms I, Slaveykova VI (2020) Interaction of silver nanoparticles with antioxidant enzymes. Environ Sci Nano 7(5):1507–1517
- Lorito M, Peterbauer C, Hayes CK, Harman GE (1994) Synergistic interaction between fungal cell wall degrading enzymes and different antifungal compounds enhances inhibition of spore germination. Microbiology 140:623–629
- MacCormack TJ, Clark RJ, Dang MK, Ma G, Kelly JA, Veinot JG, Goss GG (2012) Inhibition of enzyme activity by nanomaterials: potential mechanisms and implications for nanotoxicity testing. Nanotoxicology 6(5):514–525
- Marathe K, Naik J, Maheshwari V (2021) Biogenic synthesis of silver nanoparticles using Streptomyces spp. and their antifungal activity against Fusarium verticillioides. J Clust Sci 32 (5):1299–1309
- Mommaerts V, Jodko K, Thomassen LC, Martens JA, Kirsch-Volders M, Smagghe G (2012) Assessment of side-effects by LudoxTMA silica nanoparticles following a dietary exposure on the bumblebee Bombus terrestris. Nanotoxicology 6:554–561
- National Nanotechnology Initiative (NNI) (2020) Available online: www.nano.gov (accessed on 22 July 2019)
- Naz SS, Shah MR, Islam NU, Khan A, Nazir S, Qaisar S, Alam SS (2014) Synthesis and bioactivities of silver nanoparticles capped with 5-amino- β -resorcylic acid hydrochloride dihydrate. J Nanobiotechnology 12:1–8
- Naz SS, Shah MR, Islam NU, Alam SS (2019) Enhanced urease inhibition activity of Ag nanomaterials capped with N-substituted methyl 5-acetamido-β-resorcylate. Prog Nat Sci Mater Int 29(2):129–137
- Nisar M, Khan SA, Shah MR, Khan A, Farooq U, Uddin G, Ahmad B (2015) Moxifloxacin-capped noble metal nanoparticles as potential urease inhibitors. New J Chem 39(10):8080–8086
- Ouertani A, Neifar M, Ouertani R (2019) Effectiveness of enzyme inhibitors in biomedicine and pharmacotherapy. Tissue Eng Regen Med 5(2):85–90
- Patil CD, Borase HP, Suryawanshi RK, Patil SV (2016) Trypsin inactivation by latex fabricated gold nanoparticles: a new strategy towards insect control. Enzyme Microb Technol 92:18–25
- Pittarate S, Rajula J, Rahman A, Vivekanandhan P, Thungrabeab M, Mekchay S, Krutmuang P (2021) Insecticidal effect of zinc oxide nanoparticles against Spodoptera frugiperda under laboratory conditions. Insects 12(11):1017
- Posgai R, Cipolla-McCulloch CB, Murphy KR, Hussain SM, Rowe JJ, Nielsen MG (2011) Differential toxicity of silver and titanium dioxide nanoparticles on Drosophila melanogaster development, reproductive effort, and viability: size, coatings and antioxidants matter. Chemosphere 85:34–42
- Rahman MF, Wang J, Patterson TA, Saini UT, Robinson BL, Newport GD, Murdock RC, Schlager JJ, Hussain SM, Ali SF (2009) Expression of genes related to oxidative stress in the mouse brain after exposure to silver-25 nanoparticles. Toxicol Lett 187:15–21
- Raj A, Shah P, Agrawal N (2017) Sedentary behaviour and altered metabolic activity by AgNPs ingestion in Drosophila melanogaster. Sci Rep 7:1–10
- Schnebli HP, Braun NJ (1986) Proteinase inhibitors as drugs. In: Barrett AJ, Salvesen A (eds) Proteinase inhibitors. Elsevier, Amsterdam, pp 613–627
- Scrinis G, Lyons K (2007) The emerging nano-corporate paradigm: nanotechnology and the transformation of nature, food and agri-food systems. Int J Soc Food Agric 15:22–44
- Shaker AM, Zaki Elham AH, Abdel-Rahim F, Khedr MH (2016) Novel CuO nanoparticles for pest management and pesticides photodegradation. Adv Environ Biol 10:274–283

- Shin YJ, Kwak JI, An YJ (2012) Evidence for the inhibitory effects of silver nanoparticles on the activities of soil exoenzymes. Chemosphere 88(4):524–529
- Simonin M, Cantarel AAM, Crouzet A, Gervaix J, Martins JMF, Richaume A (2018) Negative effects of copper oxide nanoparticles on carbon and nitrogen cycle microbial activities in contrasting agricultural soils and in presence of plants. Front Microbiol 9:3102. https://doi. org/10.3389/fmicb.2018.03102
- Singh RP, Handa R, Manchanda G (2021) Nanoparticles in sustainable agriculture: an emerging opportunity. J Control Release 329:1234–1248
- Srivastava A, Rao DP (2014) Enhancement of seed germination and plant growth of wheat, maize, peanut and garlic using multiwalled carbon nanotubes. Eur Chem Bull 3:502–504
- Stadler T, Lopez-Garcia GP, Gitto JG, Buteler M (2017) Nanostructured alumina: biocidal properties and mechanism of action of a novel insecticide powder. Bull Insectol 70:17–25
- Touisni N, Kanfar N, Ulrich S, Dumy P, Supuran CT, Mehdi A, Winum JY (2015) Fluorescent silica nanoparticles with multivalent inhibitory effects towards carbonic anhydrases. Chem A Eur J 21(29):10306–10309
- Umezawa H (1982) Low-molecular-weight enzyme inhibitors of microbial origin. Annu Rev Microbiol 36:75–99
- Upadhyay LS (2012) Urease inhibitors: a review. Indian J Biotechnol 11(1):81-388
- VandeVoort AR, Arai Y (2018) Macroscopic observation of soil nitrification kinetics impacted by copper nanoparticles: implications for micronutrient nanofertilizer. Nanomaterials (Basel) 8(11):927
- Vertegel AA, Siegel RW, Dordick JS (2004) Silica nanoparticle size influences the structure and enzymatic activity of adsorbed lysozyme. Langmuir 20(16):6800–6807
- Wang WR, Zhu RR, Xiao R, Liu H, Wang SL (2011) The electrostatic interactions between nano-TiO₂ and trypsin inhibit the enzyme activity and change the secondary structure of trypsin. Biol Trace Elem Res 142(3):435–446
- Yan C, Huang J, Cao C, Li R, Ma Y, Wang Y (2020) Effects of PVP-coated silver nanoparticles on enzyme activity, bacterial and archaeal community structure and function in a yellow-brown loam soil. Environ Sci Pollut Res 27:8058–8070
- Yasur J, Rani PU (2015) Lepidopteran insect susceptibility to silver nanoparticles and measurement of changes in their growth, development and physiology. Chemosphere 124:92–102
- Ye M, Tang L, Luo M, Zhou J, Guo B, Liu Y, Chen B (2014) Size-and timedependent alteration in metabolic activities of human hepatic cytochrome P450 isozymes by gold nanoparticles via microsomal coincubations. Nanoscale Res Lett 9(1):1–6
- Yuan Z, Li J, Cui L, Xu B, Zhang H, Yu CP (2013) Interaction of silver nanoparticles with pure nitrifying bacteria. Chemosphere 90:1404–1411
- Zhang H, Cao J, Wu S, Wang Y (2014) Mechanism of gold nanoparticles-induced trypsin inhibition: a multi-technique approach. Mol Biol Rep 41(8):4911–4918
- Zhao S, Su X, Wang Y, Yang X, Bi M, He Q, Chen Y (2020) Copper oxide nanoparticles inhibited denitrifying enzymes and electron transport system activities to influence soil denitrification and N₂O emission. Chemosphere 245:125394
- Zheng Y, Hou L, Liu M, Newell SE, Yin G, Yu C, Zhang H, Li X, Gao D, Gao J, Wang R (2017) Effects of silver nanoparticles on nitrification and associated nitrous oxide production in aquatic environments. Sci Adv 3(8):e1603229

See discussions, stats, and author profiles for this publication at: https://www.researchgate.net/publication/358684705

Microbial Food-borne Diseases Due to Climate Change

Chapter · February 2022 DOI: 10.1201/9781003189725-7



Some of the authors of this publication are also working on these related projects:

Bioprospecting of endophytic fungi from medicinal plants View project

-

Green synthesis of nanoparticles View project

Climate Change

and Microbes

Impacts and Vulnerability



Javid A. Parray | Suhaib A. Bandh | Nowsheen Shameem Editors

CLIMATE CHANGE AND MICROBES

Impacts and Vulnerability

Author Copy

Apple Academic Press

Author Copy

CLIMATE CHANGE AND MICROBES

Impacts and Vulnerability

Edited by

Nuthor Copy

Javid A. Parray, PhD Suhaib A. Bandh, PhD Nowsheen Shameem, PhD



First edition published 2022

Apple Academic Press Inc. 1265 Goldenrod Circle, NE, Palm Bay, FL 32905 USA 4164 Lakeshore Road, Burlington, ON, L7L 1A4 Canada

© 2022 Apple Academic Press, Inc.

IC Press

Apple Academ

CRC Press 6000 Broken Sound Parkway NW, Suite 300, Boca Raton, FL 33487-2742 USA 2 Park Square, Milton Park, Abingdon, Oxon, OX14 4RN UK

Apple Academic Press exclusively co-publishes with CRC Press, an imprint of Taylor & Francis Group, LLC

Reasonable efforts have been made to publish reliable data and information, but the authors, editors, and publisher cannot assume responsibility for the validity of all materials or the consequences of their use. The authors, editors, and publishers have attempted to trace the copyright holders of all material reproduced in this publication and apologize to copyright holders if permission to publish in this form has not been obtained. If any copyright material has not been acknowledged, please write and let us know so we may rectify in any future reprint.

Except as permitted under U.S. Copyright Law, no part of this book may be reprinted, reproduced, transmitted, or utilized in any form by any electronic, mechanical, or other means, now known or hereafter invented, including photocopying, microfilming, and recording, or in any information storage or retrieval system, without written permission from the publishers.

For permission to photocopy or use material electronically from this work, access www.copyright.com or contact the Copyright Clearance Center, Inc. (CCC), 222 Rosewood Drive, Danvers, MA 01923, 978-750-8400. For works that are not available on CCC please contact mpkbookspermissions@tandf.co.uk

Trademark notice: Product or corporate names may be trademarks or registered trademarks and are used only for identification and explanation without intent to infringe.

Library and Archives Canada Cataloguing in Publication

Title: Climate change and microbes : impacts and vulnerability / edited by Javid A. Parray, PhD, Suhaib A. Bandh, PhD, Nowsheen Shameem, PhD.

Names: Parray, Javid Ahmad, editor. | Bandh, Suhaib A., editor. | Shameem, Nowsheen, editor.

Description: First edition. | Includes bibliographical references and index.

Identifiers: Canadiana (print) 20210370149 | Canadiana (ebook) 20210370203 | ISBN 9781774637210 (hardcover) | ISBN 9781774637968 (softcover) | ISBN 9781003189725 (ebook)

Subjects: LCSH: Microbial ecology. | LCSH: Climatic changes. | LCSH: Microorganisms. | LCSH: Plants-Microbiology. Classification: LCC QR100 .C53 2022 | DDC 579/.17-dc23

Library of Congress Cataloging-in-Publication Data

CIP data on file with US Library of Congress

ISBN: 978-1-77463-721-0 (hbk) ISBN: 978-1-77463-796-8 (pbk) ISBN: 978-1-00318-972-5 (ebk)

Javid A. Parray, PhD

Javid A. Parray, PhD, is an Assistant Professor in the Higher Education Department at Government Degree College Eidgah, Srinagar, India, where he teaches the subject of environmental science. He has published many research articles in reputable refereed international and national journals. He had authored books on different themes, including Approaches to Heavy Metal Tolerance in Plants; Sustainable Agriculture: Biotechniques in Plant Biology; and Sustainable Agriculture: Advances in Plant Metabolome and Microbiome. He was awarded the "Emerging Scientist of Year Award" by the Indian Academy of Environmental Science for the year 2015 in addition to being awarded with an international travel grant for participating in international conferences. Dr. Parray completed a fast-track project entitled "Molecular characterization and metabolic potential of rhizospheric bacteria from Arnebia benthamii across North Western Himalaya" at CORD, University of Kashmir, India. He finished his postdoctorate research associateship on a DBT-funded project entitled "Tissue culture-based network programme on saffron." He earned a PhD in Environmental Science with a specialization in plant microbe interaction on the topic "Evaluating role of Rhizospheric bacteria in saffron culture" from the University of Kashmir, India.

Suhaib A. Bandh, PhD

Apple Academic Press

Suhaib A. Bandh, PhD, is an Assistant Professor in the Higher Education Department of Jammu and Kashmir at Government Degree College, DH Pora, Kulgam, India, where he teaches courses on environmental science and disaster management at the graduate level. He is the President of the Academy of EcoScience in addition to being a life member of the Academy of Plant Sciences, India, and the National Environmental Science Academy. He is a recipient of many awards and has participated in a number of national and international conferences organized by reputed scientific bodies in India and abroad. He has a number of scientific

publications to his credit, published in some highly reputed and impacted journals. Dr. Bandh has recently edited and authored a number of books, including *Freshwater Microbiology: Perspectives of Bacterial Dynamics in Lake Ecosystems; Environmental Management: Environmental Issues, Awareness and Abatement,* and *Environmental Perspectives and Issues.* Dr. Bandh has also edited a special issue on "Ecology and Biotechnological Applications of Biofilm" in *BioMed Research International.*

Nowsheen Shameem, PhD

Nowsheen Shameem, PhD, is an Assistant Professor in the Department of Environmental Science at Cluster University Srinagar, India. She has worked as a project associate on the DBT-sanctioned project on "Spawn production for the entrepreneurs of Kashmir Valley" at CORD, University of Kashmir, India. She has also worked as a group leader for drafting a number of research proposals and ideas. She has published many research articles in reputed, refereed international and national journals of Springer, Elsevier, and Hindawi. She has also presented her research work at international and national conferences. Dr. Shameem finished her doctorate through the University of Kashmir on "Phytochemical analysis and nutraceutical value of some wild mushrooms growing in Kashmir Valley" in 2017.

Apple Academic Press

Contents

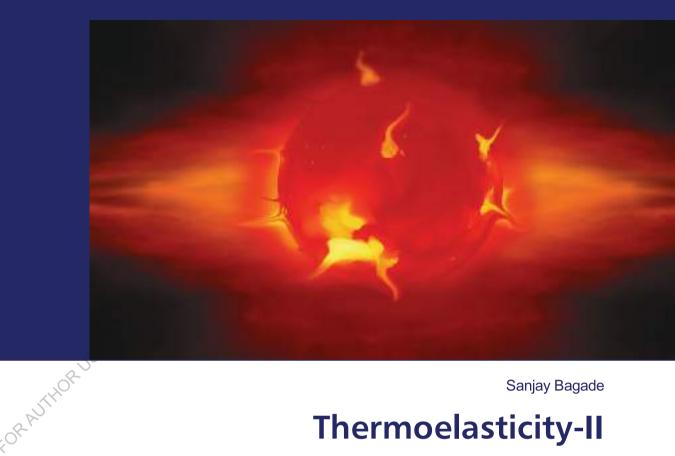
Apple Academic Press

Cor	ıtributorsix
Abk	previationsxi
Pre	facexiii
1.	Role of Microorganisms as Climate Engineers:
	Mitigation and Adaptations to Climate Change1
	Muhammad Mahroz Hussain, Zia Ur Rahman Farooqi, Fahad Rasheed, and Waqas Mohy Ud Din
2.	Climate Change-Induced Aggravations in Microbial Populations and Processes: Constraints and Remediations
	Waqas Mohy Ud Din, Muhammad Mahroz Hussain, and Zia Ur Rahman Farooqi
3.	The Vulnerability of Microbial Ecosystems in a
	Challenging Climate
	Basharat A. Bhat, Lubna Tariq, Rakeeb A. Mir, Ishfaq Majeed, and Maajid M. Bandh
4.	Climate Change and Its Influence on Microbial Diversity, Communities, and Processes
	Irteza Qayoom, Haika Mohi-ud-din, Aqsa Khursheed, Aashia Altaf, and Suhaib A. Bandh
5.	Climate Change and Its Impact on Plant-Microbe Interaction
	S. S. Khandare and M. G. Ingale
6.	Climate Change and Microbial Aquatic Life
	Tanzeela Rehman, Shahnaz Bashir, Mohazib Nabi, and Suhaib A. Bandh
7.	Microbial Food-borne Diseases Due to Climate Change
	John Mohd War, Anees Un Nisa, Abdul Hamid Wani, and Mohd Yaqub Bhat
8.	Algae as Indicators of Climate Change
	Christy B. K. Sangma and Sabira Sultana

Contents

9.	Regulation of Ethylene Levels with 1-Aminocyclopropane- 1-Carboxylate Deaminase Produced by Plant Growth	
	Promoting Rhizobacteria	267
	Aparna B. Gunjal	
10.	Core Microbiome of <i>Solanum Lycopersicum</i> for Sustainable Agroecosystems	279
	Anamika Chattopadhyay and G. Thiribhuvanamala	
Ind	lex	

This book deals with the concept of thermoelasticity which finds its roots and applications in the branch of Physics, Mathematics and Engineering. Thermoelasticity is a fast growing concept and today it is an integral part of engineering and structural designs, rocket science and space engineering. Mathematical aspects of thermoelastic problems in cylindrical and spherical structures have been dealt with and thermal displacements, thermal stress and strains in these structures have been discussed in detail. Functionally Graded Materials, which have gained popularity as high temperature and high thermal stress bearing materials and widely used in space engineering have been discussed. Magneto-thermoelastic stresses and magnetic field vector perturbations for FGM hollow cylinder, FGM hollow sphere have been deduced. Mathematical approach for thermoelastic problems may be of immense use for the researchers, teachers pursuing work on thermoelasticity.



Sanjay Bagade

Thermoelasticity-II



Dr. SANJAY H BAGADE M.Sc. (PHYSICS), Ph.D Bajaj College of Science, Wardha-442001 INDÍA He has 20 years of teaching experience and his research area of interest is Theoretical Physics.

Sanjay Bagade



Sanjay Bagade

Thermoelasticity-II

FORAUTHORUSEONIX

Sanjay Bagade

Thermoelasticity-II



LAP LAMBERT Academic Publishing

Imprint

Any brand names and product names mentioned in this book are subject to trademark, brand or patent protection and are trademarks or registered trademarks of their respective holders. The use of brand names, product names, common names, trade names, product descriptions etc. even without a particular marking in this work is in no way to be construed to mean that such names may be regarded as unrestricted in respect of trademark and brand protection legislation and could thus be used by anyone.

Cover image: www.ingimage.com

Publisher: LAP LAMBERT Academic Publishing is a trademark of Dodo Books Indian Ocean Ltd., member of the OmniScriptum S.R.L Publishing group str. A.Russo 15, of. 61, Chisinau-2068, Republic of Moldova Europe Printed at: see last page ISBN: 978-620-4-74445-2

Copyright © Sanjay Bagade Copyright © 2022 Dodo Books Indian Ocean Ltd., member of the OmniScriptum S.R.L Publishing group

THERMOELASTICITY-II

SANJAY H BAGADE

M.Sc. (PHYSICS), Ph.D.

Bajaj College of Science, Wardha-442001.India.

DEDICATED TO MY PARENTS

PREFACE

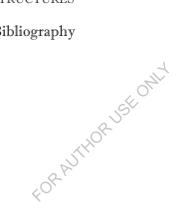
The book "THERMOELASTICITY- II" is second book on the topic dealing with thermal stresses, thermal strain and resulting displacements, the first one being THERMOELASTICITY- I. The mutual interactions between Physics, mathematics and Engineering have enriched the mankind in many ways. Thermoelasticity has been in human knowledge since two centuries, but studies related to it are recent developments. The principles of thermoelasticity have affected the way engineers design different structures and machinery. Thermoelastic investigations have now become an inevitable part of all design and construction mechanisms.

The book "THERMOELASTICITY-II "involves three chapters. The mathematical tools essential for this study have been discussed in chapter-I. Chapter-II discusses the thermoelasticity in cylindrical structures while chapter-III deals with thermoelastic behaviour of spherical structures. Thermal strain - stress in these bodies subjected to different heating conditions and boundary conditions are discussed here. Functionally Graded Materials(FGM) which has earned a reputation as the materials that can sustain more temperatures and thermal stresses and hence more useful in rocket science and aerospace engineering have also been discussed in this chapters. Hope that this book will be helpful to the students in understanding the mathematical aspects of thermoelastic problems, enabling them to make further research developments in this field.

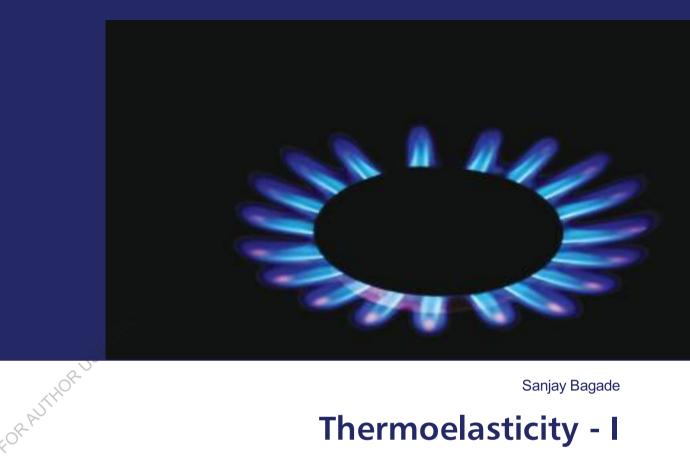
SANJAY H BAGADE

CONTENTS

Ι	INTRODUCTION	1-5
II	THERMOELASTICITY OF CYLINDRICAL STRUCTURES	7-93
III	THERMOELASTICITY OF SPHERICAL STRUCTURES	95-146
	Bibliography	147-158



Thermoelasticity is one of the streams in Science where the concepts and ideas in Physics, Mathematics and Engineering are extensively used. The fruitful interactions and applications between these branches have led to many technological advancements. This book deals with the concept of thermoelasticity which finds its roots and application in the above branches. Thermal response of few circular and rectangular geometrical structures subjected to different boundary and heating conditions are discussed here. Thermal stresses and strains under thermal variations and different heating conditions are elaborated. Topics discussed here should be helpful to students and researchers pursuing work on thermoelasticity.



Sanjay Bagade

Thermoelasticity - I

Thermal Stresses in Circular and Rectangular Structures



Dr. SANJAY H BAGADE. M.Sc. (PHYSICS), Ph.D. Bajaj College of Science, Wardha-442001.INDIA. Author has a teaching experience of about 20 years. His area of interest is Theoretical and Mathematical Physics.



Bagade



Sanjay Bagade

Thermoelasticity - I

FORAUTHORUSEONIX

Sanjay Bagade

Thermoelasticity - I

Thermal Stresses in Circular and Rectangular Structures



LAP LAMBERT Academic Publishing

Imprint

Any brand names and product names mentioned in this book are subject to trademark, brand or patent protection and are trademarks or registered trademarks of their respective holders. The use of brand names, product names, common names, trade names, product descriptions etc. even without a particular marking in this work is in no way to be construed to mean that such names may be regarded as unrestricted in respect of trademark and brand protection legislation and could thus be used by anyone.

Cover image: www.ingimage.com

Publisher: LAP LAMBERT Academic Publishing is a trademark of International Book Market Service Ltd., member of OmniScriptum Publishing Group 17 Meldrum Street, Beau Bassin 71504, Mauritius Printed at: see last page ISBN: 978-620-3-19308-4

Copyright © Sanjay Bagade Copyright © 2021 International Book Market Service Ltd., member of OmniScriptum Publishing Group

FORAUTHORUSEONI

THERMOELASTICITY-I

SANJAY H BAGADE

M.Sc. (PHYSICS), Ph.D.

Bajaj College of Science, Wardha-442001.India.

DEDICATED TO MY PARENTS

PREFACE

Physics has always played an important role in this fast growing world of Engineering and Technology. The concepts and discoveries from the field of Physics are harnessed by Engineers, and Engineering, in turn provides the incentives to Physics, paving a way for more concepts and discoveries. Such mutual interactions between Physics and Engineering have enriched the mankind in many ways. Thermoelasticity has been in human knowledge since two centuries, but studies related to it are recent developments. The principles of thermoelasticity have affected the way engineers design different structures and machinery. Thermoelastic investigations have now become an inevitable part of all design and construction mechanisms. This book on thermoelasticity is a small effort to acquaint the readers about the topic, making use of the concepts and tools in Physics and Mathematics.

The book "THERMOELASTICITY-I " consists of three chapters. Chapter-I deals with the basic introduction and the necessary mathematical tools used in this study. Chapter-II discusses the thermoelasticity in circular structures while chapter-III deals with thermoelastic behaviour of rectangular structures. Thermal strain -stress in these bodies subjected to different heating conditions and boundary conditions are discussed here. Hope that this book will be helpful to the students in understanding the mathematical aspects of thermoelastic problems, enabling them to make further research developments in this field.

SANJAY H BAGADE

CONTENTS

Ι	INTRODUCTION	1-8
II	THERMOELASTICITY OF CIRCULAR STRUCTURES	9-118
III	THERMOELASTICITY OF RECTANGULAR STRUCTURES	119-279
	Bibliography	281-292
	FORAUTHORUSEOMIX	

Springer Protocols

Natarajan Amaresan Prittesh Patel Dhruti Amin *Editors*

Practical Handbook on Agricultural Microbiology



Springer Protocols Handbooks

For further volumes: http://www.springer.com/series/7657 Springer Protocols Handbooks collects a diverse range of step-by-step laboratory methods and protocols from across the life and biomedical sciences. Each protocol is provided in the Springer Protocol format: readily-reproducible in a step-by-step fashion. Each protocol opens with an introductory overview, a list of the materials and reagents needed to complete the experiment, and is followed by a detailed procedure supported by a helpful notes section offering tips and tricks of the trade as well as troubleshooting advice. With a focus on large comprehensive protocol collections and an international authorship, Springer Protocols Handbooks are a valuable addition to the laboratory.

More information about this series at http://www.springer.com/series/8623

Practical Handbook on Agricultural Microbiology

Edited by

Natarajan Amaresan, Prittesh Patel, and Dhruti Amin

Uka Tarsadia University, Surat, Gujarat, India

💥 Humana Press

Editors Natarajan Amaresan Uka Tarsadia University Surat, Gujarat, India

Dhruti Amin Uka Tarsadia University Surat, Gujarat, India Prittesh Patel Uka Tarsadia University Surat, Gujarat, India

ISSN 1949-2448 ISSN 1949-2456 (electronic) Springer Protocols Handbooks ISBN 978-1-0716-1723-6 ISBN 978-1-0716-1724-3 (eBook) https://doi.org/10.1007/978-1-0716-1724-3

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Humana imprint is published by the registered company Springer Science+Business Media, LLC part of Springer Nature.

The registered company address is: 1 New York Plaza, New York, NY 10004, U.S.A.

Preface

Agricultural Microbiology is a part of the microbiology branch dealing with beneficial or harmful microbes associated with either plants or soil. This manual focuses on beneficial microbes dealing with soil fertility, microbial degradation of organic matter, soil nutrient transformations, and biocontrol agents. Nowadays, techniques involved in the study of beneficial microbes in agricultural microbiology toward enhancing global agricultural productivity are in trend. This manual covers a wide range of basic and advanced techniques associated with research on the isolation of agriculturally important microbes, identification, biological nitrogen fixation, microbe-mediated plant nutrient use efficiency, and biological control of plant diseases and pests. Introduction to each protocol explains the role/importance of chemicals involved, uniqueness, and protocol application. A proper understanding of the protocol helps the researchers to manipulate them as per their need.

This book is composed of seven parts with 52 protocol chapters. Parts I and II represent the importance, isolation, and purification methods of agriculturally important microbes and include mineral-solubilizing microbes. Part III deals with phytohormones quantitative protocols directly or indirectly associated with microbes. Parts IV and V provide deep insights into protocols for screening agriculturally important enzymes and compounds related to biocontrol activity. Part VI represents assessment methods of soil microbial activity by soil respiration. The final Part VII deals with protocols for selecting microbial strains for inoculant production and quality control ultimately representing commercial biofertilizers production criteria. This book will help postgraduate students, research scholars, postdoctoral fellows, and teachers belonging to different disciplines of Plant Microbiology and Pathology. Moreover, this manual may also serve as a textbook for undergraduate courses like Techniques on Plant-Microbe Interaction/Biological Control of Plant Diseases/Nutrient Use Efficiency.

Surat, Gujarat, India Surat, Gujarat, India Surat, Gujarat, India Natarajan Amaresan Prittesh Patel Dhruti Amin

Contents

	face stributors	v xi
Par	AT I ISOLATION AND IDENTIFICATION OF AGRICULTURALLY IMPORTANT MICROBES	
1	Methods for Isolation and Identification of Rhizobia	3
2	Isolation of Frankia from Casuarina Root Nodule Narayanasamy Marappa, Dhanasekaran Dharumadurai, and Thajuddin Nooruddin	15
3	Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum Bhavana V. Mohite and Satish V. Patil	25
4	Isolation and Identification of Azospirillum	35
5	Isolation and Identification of <i>Gluconacetobacter diazotrophicus</i>	41
6	Isolation and Identification of Nitrogen Fixing Bacteria: Azoarcus Species	47
7	Isolation and Identification of <i>Derxia</i> Species from the Soil Sample Harshida A. Gamit and Natarajan Amaresan	57
8	Isolation and Characterization of Enterobacter, Klebsiella,and ClostridiumV. Mageshwaran and K. Pandiyan	63
9	Isolation and Characterization of Genus Desulfotomaculum Mobini Pimpalse, Harshida A. Gamit, and Natarajan Amaresan	71
10	Isolation and Identification of Associative Symbiotic N ₂ Fixing Microbes: Desulfovibrio	77
11	Cyanobacteria Dhruti Amin, Abhishek Sharma, and Sanket Ray	85
12	Pseudomonas Sanket Ray and Harsh Patel	93
13	Isolation and Identification of Entomopathogenic <i>Bacillus</i> Species Preeti Parmar, B. K. Rajkumar, and Naresh Butani	99
14	Methylobacterium	111

viii	Contents

15	Isolation and Identification of <i>Beijerinckia</i>	119
16	Isolation of Streptomyces from Soil Sample Vishnu Raja Vijayakumar and Dharumadurai Dhanasekaran	127
17	Isolation and Identification of <i>Trichoderma</i> Spp. from Different Agricultural Samples	131
18	Extraction, Isolation and Culturing of <i>Mycorrhizal</i> Spores from Rhizospheric Soil	145
19	Isolation and Identification of <i>Metarhizium</i>	151
20	Isolation and Identification of Bacteriophage for Biocontrol Mitesh Dwivedi	161
21	Isolation of Bacterivorous Protozoan, Acanthamoeba Spp., as New-Age Agro Bio-Input Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil	173
Par	TT II ISOLATION OF MINERAL SOLUBILIZING MICROBES	
22	Isolation and Screening of Zinc Solubilizing Microbes: As Essential Micronutrient Bio-Inputs for Crops Satish V. Patil, Hemant P. Borase, Jitendra D. Salunkhe, and Rahul K. Suryawanshi	181
23	Isolation and Screening of Mineral Phosphate Solubilizing Microorganisms	187
24	Isolation and Identification of Potassium-Solubilizing Microbes Prittesh Patel and Swati Patel	193
25	Isolation and Identification of Sulfur-Oxidizing Bacteria Vimalkumar Prajapati, Swati Patel, Radhika Patel, and Vaibhavkumar Mehta	197
26	Isolation of Ammonia Oxidizing Bacteria Naresh Butani, Shruti Satashia, Hemanshi Kanpariya, and Preeti Parmar	203
27	Isolation and Identification of Nitrite-Oxidizing Microbes Prittesh Patel, Ami Naik, and Abhishek Sharma	219
28	Isolation and Identification of Iron-Oxidizing Microbes Ami Naik and Pooja Patel	223
29	Isolation and Identification of Phytin Mineralizing Microbes Swati Patel and Prittesh Patel	231
30	Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil	237

31	Isolation of Selenium Biotransforming Microbes as New Age Bioinputs Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil	243
Par	T III ESTIMATION OF PHYTOHORMONES BY BENEFICIAL MICROBES	
32	Auxin Dixita Panchal, Jemisha Mistry, and Dhruti Amin	251
33	Abscisic Acid Natarajan Amaresan, A. Sankaranarayanan, and Dhruti Amin	257
34	Cytokinins Jawahar Ganapathy, Jemisha Mistry, and Dhruti Amin	263
35	Ethylene	269
36	Gibberellin Dhruti Amin, Sanket Ray, and Abhishek Sharma	273
37	Brassinosteroids	277
38	Strigolactones: Extraction and Characterization	283
39	Estimation of Jasmonic Acid Using Non-pathogenic Microbes Jasmonic Acid Krutika S. Abhyankar and Monisha Kottayi	289
Par	TT IV SCREENING OF AGRICULTURALLY IMPORTANT ENZYMES	
40	Chitinase Purvesh B. Bharvad and Harsha J. Algotar	301
41	Glucanase Purvesh B. Bharvad and Harsha J. Algotar	309
42	Identification of Cellulase Enzyme Involved in Biocontrol Activity	317
43	Identification of Protease Enzymes Involved in Biocontrol Activity Vimalkumar Prajapati, Swati Patel, Sanket Ray, and Kamlesh C. Patel	323
44	Isolation and Screening of Naringinase Producing Microbes: As Industrial Inputs for Agro Waste Base Enzyme Industry Satish V. Patil, Jitendra D. Salunkhe, and Vishal Marathe	331
45	Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil	337

Par	тV	Identification of Compounds Involved in Biocontrol Activity	
46		rogen Cyanide ti Bhatt and Dhruti Amin	345
47		ophores sa Patel	351
48	for D	tion and Screening of ACC Deaminase-Producing Microbes Prought Stress Management in Crops	361
49		olysaccharides a Chandwani and Natarajan Amaresan	369
Par	тVI	Assessment of Soil Microbial Activity by Soil Respiration	
50		matic Analyses in Soils ar Bilen and Veysel Turan	377
Par	т VII	Selection of Microbial Strains for Inoculant Production and Quality Control	
51		tion of <i>Rhizobium</i> Strain for Inoculum Production	389
52		her Culture, Broth, and Peat Test uti Amin, Sanket Ray, and Vrushali Wagh	395
Ind	ex		405

Contributors

- KRUTIKA S. ABHYANKAR School of Science, Navrachana University, Vadodara, Gujarat, India
- HARSHA J. ALGOTAR . D. L. Patel Science College, Himatnagar, Gujarat, India
- NATARAJAN AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- DHRUTI AMIN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- HIMANSHU BARIYA Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- PURVESH B. BHARVAD . D. L. Patel Science College, Himatnagar, Gujarat, India
- KHYATI BHATT Post Graduate Department of Biosciences, Sardar Patel University, Gujarat, India
- SERDAR BILEN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Atatürk University, Erzurum, Turkey
- HEMANT P. BORASE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- NARESH BUTANI Department of Microbiology, Faculty of Science, Sarvajanik University, Surat, India
- SAPNA CHANDWANI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- SHREYA DESAI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- DHANASEKARAN DHARUMADURAI Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- MITESH DWIVEDI C. G. Bhakta Institute of Biotechnology, Faculty of Science, Uka Tarsadia University, Surat, Gujarat, India
- HARSHIDA A. GAMIT C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- JAWAHAR GANAPATHY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- TEJAS GOHIL Sabarmati Ashram Gaushala, Kheda, Gujarat, India
- HEMANSHI KANPARIYA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- MONISHA KOTTAYI School of Science, Navrachana University, Vadodara, Gujarat, India
- V. MAGESHWARAN ICAR-National Bureau of Agriculturally Important Microorganisms, Mau, Uttar Pradesh, India
- MAHADEVASWAMY University of Agricultural Sciences, Raichur, Karnataka, India
- NARAYANASAMY MARAPPA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- KIRAN MARATHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- VISHAL MARATHE N.E.S. Science College, Nanded, Maharashtra, India

- VAIBHAVKUMAR MEHTA Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- HARSH MISTRY Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- JEMISHA MISTRY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- BHAVANA V. MOHITE Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India
- AMI NAIK C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- VRUTUJA NAIK National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- PRADNYA B. NIKAM School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- THAJUDDIN NOORUDDIN Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- DIXITA PANCHAL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- KETANKUMAR J. PANCHAL Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, India
- K. PANDIYAN . ICAR-Central Institute for Research on Cotton Technology, Mumbai, India
- PREETI PARMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- HARSH PATEL P. G. Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, Gujarat, India
- KAMLESH C. PATEL PG Department of Biosciences, Sardar Patel University, Anand, Gujarat, India
- NAFISA PATEL Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- POOJA PATEL . Government Medical College, Surat, Gujarat, India
- PRITTESH PATEL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- RADHIKA PATEL PG Department of Biosciences, Sardar Patel University, Gujarat, India
- SWATI PATEL Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- TARUN KUMAR PATEL Department of Biotechnology, Sant Guru Ghasidas Government P.G. College, Dhamtari, Chattisgarh, India; Department of Biotechnology, Guru Ghasidas Vishwavidyalaya (a Central University), Bilaspur, Chattisgarh, India
- CHANDRASHEKHAR D. PATIL Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- SATISH V. PATIL School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India
- VIKAS S. PATIL University Institute of Chemical Technology, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- MOHINI PIMPALSE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, India
- VIMALKUMAR PRAJAPATI Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India

- PRAVEEN RAHI National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- VIJAYAKUMAR VISHNU RAJA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- B. K. RAJKUMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- SANKET RAY Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- NARENDRA SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- JITENDRA D. SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- A. SANKARANARAYANAN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- SHRUTI SATASHIA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- ABHISHEK SHARMA C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- K. SOWMIYA University of Agricultural Sciences, Raichur, Karnataka, India
- RAHUL K. SURYAWANSHI Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- RASHMI THAKOR Department of Life Sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- VEYSEL TURAN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Bingöl University, Bingöl, Turkey
- VRUSHALI WAGH Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India



Isolation and Screening of ACC Deaminase-Producing Microbes for Drought Stress Management in Crops

Satish V. Patil, Chandrashekhar D. Patil, and Bhavana V. Mohite

Abstract

The compound aminocylopropane-1-carboxylic acid (ACC) is the product of Yang cycle, which plays a vital role in drought response, salt stress, host–pathogen interactions, seed germination, flowering, and fruit ripening. While ACC is produced within the plant, some of it is exuded from the roots where it may alter the rhizosphere microbial composition. Within the plant, ACC may be converted to the phytohormone ethylene by the action of the plant enzyme ACC oxidase (ACO). When the rhizosphere or the plant endosphere contains microbes that produce ACC deaminase, some of the ACC is converted to alpha ketobutyric acid and ammonia. Thus, reducing ethylene production and decreasing the negative effects of ethylene including numerous drought-like symptoms, thereby reduces the deleterious effects of drought stress. ACC deaminase-producing microorganisms may readily be isolated by using minimal medium containing ACC as sole source of nitrogen and a pH indicator. The microbial cleavage of ACC changes the pH of the growth medium and causes an easily detectable color change around the colony. Subsequently, qualitative or quantitative assays for alpha ketoglutaric acid or ammonia may be used to assess the level of ACC deaminase activity.

Key words PGPR, ACC deaminase, Screening, pH dye, Phenol red

1 Introduction

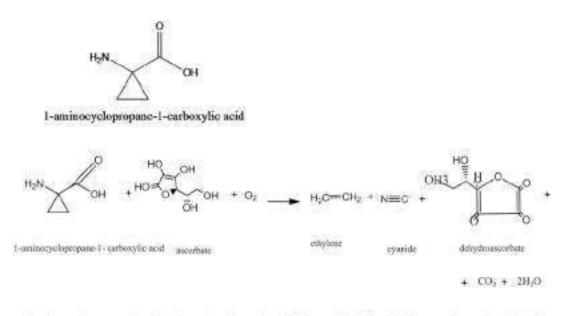
The studies of Yang and his colleagues pioneered the research to unlock the mystery of freshness of fruit, flowers, defoliations, the ripening of fruits, etc. through the Yang cycle [1]. Ethylene has a vital role in host–pathogen interactions, seed germination, flowering, fruit ripening, and the response of plants to both biotic and abiotic stress [2].

Yang's studies proved that the synthesis of S-adenosylmethionine is an intermediate compound which further converted to aminocylopropane-1-carboxylic acid (ACC) and then, by the action of the enzyme ACC oxidase to ethylene (Fig. 1).

Springer Protocols Handbooks, https://doi.org/10.1007/978-1-0716-1724-3_48,

© The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

Natarajan Amaresan et al. (eds.), Practical Handbook on Agricultural Microbiology,



(1-aminocyclopropane-I-carboxylate + Ascorbate + O₂ = Ethylene + Cyanide + dehydroascorbate + CO₂ + 2 H₂O)

Fig. 1 Conversion of ACC into ethylene

In plants, ethylene level has been found to be rapidly increased following the onset of drought and other stress conditions [2, 3]. This ethylene synthesis is a marker of the stress condition and is sometimes known as stress ethylene.

Glick et al. [4] described the role and importance of some plant growth promoting Rhizobacteria in management of drought stress and various physiological activities of plants. This work showed that the ACC is produced in greater quantities during drought stress; ACC is exudated outside by the root cells. The plant growth inducing bacteria around the roots are recognized for their various beneficial activities. These microbes utilize the plant exudates including ACC by using ACC deaminase which helps to maintain the balance between the inner and outer ACC level. As more of the ACC gets secreted outside the plant roots, it reduces the concentration of ACC that can be used for the synthesis of ethylene inside the plant cells. In this way by utilizing ACC, rhizospheric microbes help to reduce the ethylene production that occurs during drought conditions. This leads to less defoliation, more root elongation, increased nodulations, and increased transpiration activities [5] (Fig. 2).

Various rhizospheric microbes have been reported to have ACC deaminase-producing potential, e.g., Enterobacter cloacae, Pseudomonas putida, Pseudomonas alcaligenes, Hansenula spp., Rhizobium, Sinorhizobium spp., Pseudomonas chlororaphis, Rhizobium leguminosarum, Bacillus subtilis, Penicillium citrinum, etc. [6].

1.1 Aminocylopropane-1-Carboxylic Acid Deaminase and Its Role

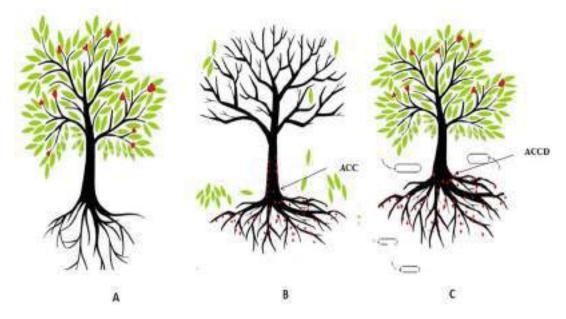


Fig. 2 ACC deaminase-producing microorganisms are attracted towards plants during stress conditions. (a) Normal plant, (b) Under the stress conditions the level of ACC increases and subsequent formation of ethylene causes defoliation and root growth inhibition. (c) The rhizospheric microorganisms utilize ACC in root exudates through the functioning of ACC deaminase and decrease the effects of the stress condition

ACC deaminase is a member of the oxidoreductase enzymes, a multimeric enzyme having monomeric subunit of molecular mass of 35–42 KDa [5]. ACC deaminase catalyzes the ACC to alpha ketobutyric acid and ammonia. It was reported that D-serine and D-cysteine can also act as substrates for ACC deaminase. ACC deaminase has a reported optimum pH and temperature of 8.5 and 30–35 °C, respectively [7–9].

This assay combines the two classical approaches as shown in Fig. 3, "Reproduced from Patil et al. [10] with permission from [Elsevier]" (1) Cultivation of test microorganisms on ACC-containing minimal medium [11] and (2) addition of pH indicator dyes phenol red and bromothymol blue.

The ACC deaminase-producing microbes utilize the synthetic substrate, ACC as a sole nitrogen source in minimal medium. The growing bacteria show a zone of clearance relative to its capacity for substrate utilization. The pH indicator dye shows a zone of color change around the colony due to the production of ammonia, a byproduct of the catalytic reaction of ACC deaminase.

1.2 Principle of Assay

Screening of ACC deaminase producers

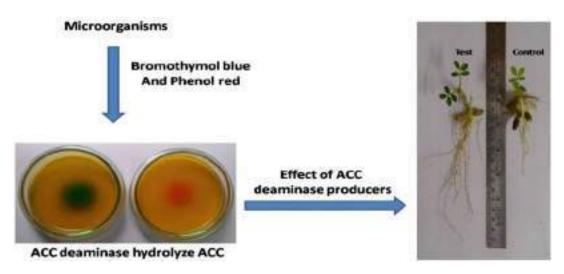


Fig. 3 Example of isolation and application of an ACCD producing microorganism and plant growth assay. ("Reproduced from Patil et al. [10] with permission from [Elsevier]")

2 Materials

2.1 Isolation	1. Rhizospheric soil of plants, especially from drought areas.
of Rhizobacteria	2. Sterile plastic screw cap bottle.
and Qualitative Estimation of ACC	3. Sterile saline tubes for serial dilution of the soil samples.
Deaminase Activity	4. Sterile modified minimal medium.
Dealiniaco neurity	5. Sterile Nutrient or LB agar.
	6. Modified Dworkin and Foster salt medium.
	7. Stock solution of trace elements.
2.2 ACC Deaminase	1. α-ketobutyrate.
Enzyme Assay	2. 0.1% 2, 4-dinitrophenyl hydrazine solution.
	3. 2 N sodium hydroxide.
	4. 0.1 M Tris (hydroxymethyl amino methane) buffer (pH 7.5).
	5. 0.56 N hydrochloric acid (HCl).
	6. Trichloroacetic acid.
	7. Nesslers Reagent.
	8. Ammonium sulfate.
	9. Potassium bromide.

3 Methods

1. Prepare 100 ml of medium by mixing sterile modified minimal 3.1 Stepwise medium and the sterilized agar-agar solution, mix well by Procedure rotating flask and pour solution in plates. for the Isolation and Screening of ACC 2. Allow the plates to solidify. Deaminase-Producing 3. Make serial dilutions of the test soil sample. Microbes from Soil 4. Streak the diluted soil suspension of dilutions 10^4 – 10^7 on solidified ACC plates, and incubate for 48 h at 37 $^\circ \mathrm{C}.$ 5. Observe the color change around the colonies (Fig. 3) [10]. 6. Subculture the pure colonies on nutrient or LB agar. 7. Make spot of diluted colony at the center of ACC plates and then incubate the plates for 48 h. 8. Measure the zone of color change as a function of time. 9. Compare the isolated colonies with the zone of color changes and note the colony characters. 3.2 Collection of Soil For the isolation of ACC deaminase producers, rhizosphere soil of plants, especially from drought areas, should be collected in sterile plastic bottles. In addition, other rhizosphere soils from different locations may also be useful. Collect 1 g of rhizosphere soil in a sterile plastic screw cap bottle, close the bottle, label it, and store at 4 °C or freeze until further processing. Dilute the collected soil sample up to 10^{-9} dilution in sterile 10 mL 3.3 Isolation tubes, mix well and incubate for 10 min at room temperature. Then of Rhizobacteria prepare modified Dworkin and Foster salt medium containing in and Qualitative g/l: Glucose, 2.0; KH₂PO₄, 4.0; Citric acid, 2.0; Gluconic acid, Estimation of ACC 2.0; MgSO₄ \cdot H₂O 0.2; Na₂HPO₄, 6.0; and 3 mM amino cyclo **Deaminase Activity** propane, 594 ml; Distilled water to 1 l; mix this solution well. Prepare stock solution of trace elements as follows: add $FeSO_4 \cdot 7H_2O$, 10 mg in 10 ml distilled water; 100 mg H_3BO_3 in 10 ml DW; 110 mg of MnSO₄ \cdot H₂O in 10 ml DW; 1240 mg $ZnSO_4 \cdot 7H_2O$ in 10 ml DW; 780 mg CuSO₄ $\cdot 5H_2O$ in 10 ml DW; and 100 mg MoO₃ in 10 ml DW. Add 1 ml of each trace solution into above prepared Modified DF medium; maintain final pH of the medium 6.8-7.2. Then add 1 ml of bromophenol blue or phenol red into the medium mix, plug with cotton, and wrap with paper. To a separate flask, add 30 g of agar powder in 400 ml DW,

mix well, plug the flask with cotton, wrap with paper and sterilize both preparations. After sterilization, allow the medium to cool to 40 °C and mix in agar solution into the modified DF medium and prepare plates.

3.4 ACC Deaminase Enzyme Assay	Preparation of standard curve of α -ketobutyrate: For quantitative assay, prepare stock of alpha-ketobutyrate (0.1–1.0 μ M) and add 0.3 ml of 0.1% 2, 4-dinitrophenyl hydrazine solution to it and incubate mixture at 30 °C, for 30 min. Thereafter add 2 ml of 2 N sodium hydroxide to the above mixture and measure the absorbance at 540 nm, and prepare a standard graph. For test organism capacity, estimate protein content of crude enzyme/fermented broth of isolate by the Bradford method and express activity by measuring the nano mole/1 μ M of alpha keto butyrate liberated per mg of cellular protein per hour.
3.4.1 ACC Deaminase Assay	For estimation of α -ketobutyrate, mix the enzyme extract (0.2 ml) and ACC (50 mM) and incubate in 0.2 ml of 0.1 M Tris (hydro- xymethyl amino methane) buffer (pH 7.5) at 30 °C for 30 min. Terminate the reaction by adding 1.8 ml of 0.56 N hydrochloric acid (HCl). Add 0.3 ml of 0.1% 2,4-dinitrophenyl hydrazine solu- tion and keep the mixture at 30 °C for 30 min. Thereafter, add 2 ml of 2 N sodium hydroxide in the above mixture and measure the absorbance at 540 nm. One unit of enzyme activity is defined as the "amount of enzyme liberating 1 μ M of α -ketobutyrate/min." Per- form the experiment at least in triplicate. (Reaction mixture can also be used for confirmation of activity by FTIR (<i>see</i> Notes 1 and 2).
3.5 Confirmatory Test for ACC Deaminase Production	In addition to α -ketobutyrate measurement, ACC deaminase production can be confirmed qualitatively and quantitatively by analyzing the release of ammonia from ACC. The procedure includes: Take a 100 µl mixture of positive bacteria and 400 µl 0.3 M ACC in Tris buffer solution and incubate at 37 °C for exactly 45 min. After the incubation add 250 µl trichloroacetic acid 24.5% (w/w) and centrifuge the mixture at 50,000 rpm. Collect the supernatant and add 250 µl of the supernatant to the Nesslers solution (2000 µl water plus 250 µl Nesslers reagent). Measure the optical density at 450 nm and compare the absorbance with an ammonium sulfate-Nesslers-standard curve based on daily calibration at concentrations of 0.05, 0.1, 0.25, 0.5, 1.0, and 2.5 mM. The units of enzyme activity are defined as micromoles of ammonia released per minute.
3.6 Quantitative Estimation	By using standard graph of α -ketobutyrate, extrapolate the reading of test sample (ACC deaminase-producing bacteria) and determine amount of α -ketobutyrate released where one unit of enzyme activ- ity measured is the amount of enzyme liberating 1 µg of α -ketobutyrate/min.

4 Notes

1. For confirmation of the α -ketobutyrate liberation after a deamination reaction by microbial ACC deaminase activity, the reaction mixture or broth may be used for potassium bromide

(KBr) cell pellet preparation and analysis with the help of Fourier-transform infrared spectroscopy (FTIR).

2. In FTIR spectra analysis results, observe peaks at 1689 and 3343 cm⁻¹, which confirm the presence of a ketonic group and amino functional group, respectively recognized as α -ketobutyrate [12].

Acknowledgments

We acknowledge *Prof. Bernard R. Glick, Professor, Department of Biology, University of Waterloo, Canada*, for inspiring us to work in agromicrobiology by his huge and legend contribution in agro biotechnology.

References

- Yang SF, Hoffman NE (1984) Ethylene biosynthesis and its regulation in higher-plants. Annu Rev Plant Physiol 35:155–189
- 2. Abeles FB, Morgan PW, Saltveit ME Jr (1992) Ethylene in plant biology, 2nd edn. Academic Press, San Diego
- Morgan PW, Drew MC (1997) Ethylene and plant response to stress. Physiol Plant 100:620–630
- Glick BR, Penrose DM, Li J (1998) A model for the lowering of plant ethylene concentrations by plant growth promoting bacteria. J Theor Biol 190:63–68
- Glick BR, Cheng Z, Czarny J (2007) Promotion of plant growth by ACC deaminaseproducing soil bacteria. Eur J Plant Pathol 119:329–333
- Glick BR (2014) Bacteria with ACC deaminase can promote plant growth and help to feed the world. Microbiol Res 169:30–39
- Jacobson CB, Pasternak JJ, Glick BR (1994) Partial purification and characterization of ACC deaminase from the plant growthpromoting rhizobacterium *Pseudomonas*

putida GR12-2. Can J Microbiol 40:1019–1025

- Honma M, Shimomura T (1978) Metabolism of 1-aminocyclopropane-1-carboxylic acid. Agric Biol Chem 42:1825–1831
- 9. Jia YJ, Kakuta Y, Sugawara M (1999) Synthesis and degradation of 1-aminocyclopropane-1carboxylicacid by *Penicillium citrinum*. Biosci Biotech Biochem 63:542–549
- Patil CD, Suryawanshi RK, Koli SH, Patil SV (2016) Improved method for effective screening of ACC (1-aminocyclopropane-1-carboxylate) deaminase producing microorganisms. J Microbiol Methods 131:102–104
- Penrose DM, Glick BR (2003) Methods for isolating and characterizing ACC deaminase contaiing plant growth-promoting rhizobacteria. Physiol Plant 118:10–15
- 12. Sarkar A, Ghosh PK, Pramanik K et al (2018) A halotolerant *Enterobacter* sp. displaying ACC deaminase activity promotes rice seedling growth under salt stress. Res Microbiol 169:20–32

Springer Protocols

Natarajan Amaresan Prittesh Patel Dhruti Amin *Editors*

Practical Handbook on Agricultural Microbiology



Springer Protocols Handbooks

For further volumes: http://www.springer.com/series/7657 Springer Protocols Handbooks collects a diverse range of step-by-step laboratory methods and protocols from across the life and biomedical sciences. Each protocol is provided in the Springer Protocol format: readily-reproducible in a step-by-step fashion. Each protocol opens with an introductory overview, a list of the materials and reagents needed to complete the experiment, and is followed by a detailed procedure supported by a helpful notes section offering tips and tricks of the trade as well as troubleshooting advice. With a focus on large comprehensive protocol collections and an international authorship, Springer Protocols Handbooks are a valuable addition to the laboratory.

More information about this series at http://www.springer.com/series/8623

Practical Handbook on Agricultural Microbiology

Edited by

Natarajan Amaresan, Prittesh Patel, and Dhruti Amin

Uka Tarsadia University, Surat, Gujarat, India

💥 Humana Press

Editors Natarajan Amaresan Uka Tarsadia University Surat, Gujarat, India

Dhruti Amin Uka Tarsadia University Surat, Gujarat, India Prittesh Patel Uka Tarsadia University Surat, Gujarat, India

ISSN 1949-2448 ISSN 1949-2456 (electronic) Springer Protocols Handbooks ISBN 978-1-0716-1723-6 ISBN 978-1-0716-1724-3 (eBook) https://doi.org/10.1007/978-1-0716-1724-3

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Humana imprint is published by the registered company Springer Science+Business Media, LLC part of Springer Nature.

The registered company address is: 1 New York Plaza, New York, NY 10004, U.S.A.

Preface

Agricultural Microbiology is a part of the microbiology branch dealing with beneficial or harmful microbes associated with either plants or soil. This manual focuses on beneficial microbes dealing with soil fertility, microbial degradation of organic matter, soil nutrient transformations, and biocontrol agents. Nowadays, techniques involved in the study of beneficial microbes in agricultural microbiology toward enhancing global agricultural productivity are in trend. This manual covers a wide range of basic and advanced techniques associated with research on the isolation of agriculturally important microbes, identification, biological nitrogen fixation, microbe-mediated plant nutrient use efficiency, and biological control of plant diseases and pests. Introduction to each protocol explains the role/importance of chemicals involved, uniqueness, and protocol application. A proper understanding of the protocol helps the researchers to manipulate them as per their need.

This book is composed of seven parts with 52 protocol chapters. Parts I and II represent the importance, isolation, and purification methods of agriculturally important microbes and include mineral-solubilizing microbes. Part III deals with phytohormones quantitative protocols directly or indirectly associated with microbes. Parts IV and V provide deep insights into protocols for screening agriculturally important enzymes and compounds related to biocontrol activity. Part VI represents assessment methods of soil microbial activity by soil respiration. The final Part VII deals with protocols for selecting microbial strains for inoculant production and quality control ultimately representing commercial biofertilizers production criteria. This book will help postgraduate students, research scholars, postdoctoral fellows, and teachers belonging to different disciplines of Plant Microbiology and Pathology. Moreover, this manual may also serve as a textbook for undergraduate courses like Techniques on Plant-Microbe Interaction/Biological Control of Plant Diseases/Nutrient Use Efficiency.

Surat, Gujarat, India Surat, Gujarat, India Surat, Gujarat, India Natarajan Amaresan Prittesh Patel Dhruti Amin

Contents

	face stributors	v xi
Par	AT I ISOLATION AND IDENTIFICATION OF AGRICULTURALLY IMPORTANT MICROBES	
1	Methods for Isolation and Identification of Rhizobia	3
2	Isolation of Frankia from Casuarina Root Nodule Narayanasamy Marappa, Dhanasekaran Dharumadurai, and Thajuddin Nooruddin	15
3	Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum Bhavana V. Mohite and Satish V. Patil	25
4	Isolation and Identification of Azospirillum	35
5	Isolation and Identification of <i>Gluconacetobacter diazotrophicus</i>	41
6	Isolation and Identification of Nitrogen Fixing Bacteria: Azoarcus Species	47
7	Isolation and Identification of <i>Derxia</i> Species from the Soil Sample Harshida A. Gamit and Natarajan Amaresan	57
8	Isolation and Characterization of Enterobacter, Klebsiella,and ClostridiumV. Mageshwaran and K. Pandiyan	63
9	Isolation and Characterization of Genus Desulfotomaculum Mobini Pimpalse, Harshida A. Gamit, and Natarajan Amaresan	71
10	Isolation and Identification of Associative Symbiotic N ₂ Fixing Microbes: Desulfovibrio	77
11	Cyanobacteria Dhruti Amin, Abhishek Sharma, and Sanket Ray	85
12	Pseudomonas Sanket Ray and Harsh Patel	93
13	Isolation and Identification of Entomopathogenic <i>Bacillus</i> Species Preeti Parmar, B. K. Rajkumar, and Naresh Butani	99
14	Methylobacterium	111

viii	Contents

15	Isolation and Identification of <i>Beijerinckia</i>	119
16	Isolation of Streptomyces from Soil Sample Vishnu Raja Vijayakumar and Dharumadurai Dhanasekaran	127
17	Isolation and Identification of <i>Trichoderma</i> Spp. from Different Agricultural Samples	131
18	Extraction, Isolation and Culturing of <i>Mycorrhizal</i> Spores from Rhizospheric Soil	145
19	Isolation and Identification of <i>Metarhizium</i>	151
20	Isolation and Identification of Bacteriophage for Biocontrol Mitesh Dwivedi	161
21	Isolation of Bacterivorous Protozoan, Acanthamoeba Spp., as New-Age Agro Bio-Input Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil	173
Par	TT II ISOLATION OF MINERAL SOLUBILIZING MICROBES	
22	Isolation and Screening of Zinc Solubilizing Microbes: As Essential Micronutrient Bio-Inputs for Crops Satish V. Patil, Hemant P. Borase, Jitendra D. Salunkhe, and Rahul K. Suryawanshi	181
23	Isolation and Screening of Mineral Phosphate Solubilizing Microorganisms	187
24	Isolation and Identification of Potassium-Solubilizing Microbes Prittesh Patel and Swati Patel	193
25	Isolation and Identification of Sulfur-Oxidizing Bacteria Vimalkumar Prajapati, Swati Patel, Radhika Patel, and Vaibhavkumar Mehta	197
26	Isolation of Ammonia Oxidizing Bacteria Naresh Butani, Shruti Satashia, Hemanshi Kanpariya, and Preeti Parmar	203
27	Isolation and Identification of Nitrite-Oxidizing Microbes Prittesh Patel, Ami Naik, and Abhishek Sharma	219
28	Isolation and Identification of Iron-Oxidizing Microbes Ami Naik and Pooja Patel	223
29	Isolation and Identification of Phytin Mineralizing Microbes Swati Patel and Prittesh Patel	231
30	Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil	237

31	Isolation of Selenium Biotransforming Microbes as New Age Bioinputs Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil	243
Par	T III ESTIMATION OF PHYTOHORMONES BY BENEFICIAL MICROBES	
32	Auxin Dixita Panchal, Jemisha Mistry, and Dhruti Amin	251
33	Abscisic Acid Natarajan Amaresan, A. Sankaranarayanan, and Dhruti Amin	257
34	Cytokinins Jawahar Ganapathy, Jemisha Mistry, and Dhruti Amin	263
35	Ethylene Ketankumar J. Panchal and Dhruti Amin	269
36	Gibberellin Dhruti Amin, Sanket Ray, and Abhishek Sharma	273
37	Brassinosteroids	277
38	Strigolactones: Extraction and Characterization	283
39	Estimation of Jasmonic Acid Using Non-pathogenic Microbes Jasmonic Acid Krutika S. Abhyankar and Monisha Kottayi	289
Par	TT IV SCREENING OF AGRICULTURALLY IMPORTANT ENZYMES	
40	Chitinase Purvesh B. Bharvad and Harsha J. Algotar	301
41	Glucanase Purvesh B. Bharvad and Harsha J. Algotar	309
42	Identification of Cellulase Enzyme Involved in Biocontrol Activity	317
43	Identification of Protease Enzymes Involved in Biocontrol Activity Vimalkumar Prajapati, Swati Patel, Sanket Ray, and Kamlesh C. Patel	323
44	Isolation and Screening of Naringinase Producing Microbes: As Industrial Inputs for Agro Waste Base Enzyme Industry Satish V. Patil, Jitendra D. Salunkhe, and Vishal Marathe	331
45	Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil	337

Part V		Identification of Compounds Involved in Biocontrol Activity	
46	Hydrogen Cyanide Khyati Bhatt and Dhruti Amin		345
47		ophores sa Patel	351
48	for D	tion and Screening of ACC Deaminase-Producing Microbes Prought Stress Management in Crops	361
49		olysaccharides a Chandwani and Natarajan Amaresan	369
Par	тVI	Assessment of Soil Microbial Activity by Soil Respiration	
50		matic Analyses in Soils ar Bilen and Veysel Turan	377
Par	т VII	Selection of Microbial Strains for Inoculant Production and Quality Control	
51		tion of <i>Rhizobium</i> Strain for Inoculum Production	389
52		her Culture, Broth, and Peat Test uti Amin, Sanket Ray, and Vrushali Wagh	395
Ind	ex		405

Contributors

- KRUTIKA S. ABHYANKAR School of Science, Navrachana University, Vadodara, Gujarat, India
- HARSHA J. ALGOTAR . D. L. Patel Science College, Himatnagar, Gujarat, India
- NATARAJAN AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- DHRUTI AMIN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- HIMANSHU BARIYA Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- PURVESH B. BHARVAD . D. L. Patel Science College, Himatnagar, Gujarat, India
- KHYATI BHATT Post Graduate Department of Biosciences, Sardar Patel University, Gujarat, India
- SERDAR BILEN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Atatürk University, Erzurum, Turkey
- HEMANT P. BORASE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- NARESH BUTANI Department of Microbiology, Faculty of Science, Sarvajanik University, Surat, India
- SAPNA CHANDWANI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- SHREYA DESAI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- DHANASEKARAN DHARUMADURAI Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- MITESH DWIVEDI C. G. Bhakta Institute of Biotechnology, Faculty of Science, Uka Tarsadia University, Surat, Gujarat, India
- HARSHIDA A. GAMIT C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- JAWAHAR GANAPATHY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- TEJAS GOHIL Sabarmati Ashram Gaushala, Kheda, Gujarat, India
- HEMANSHI KANPARIYA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- MONISHA KOTTAYI School of Science, Navrachana University, Vadodara, Gujarat, India
- V. MAGESHWARAN ICAR-National Bureau of Agriculturally Important Microorganisms, Mau, Uttar Pradesh, India
- MAHADEVASWAMY University of Agricultural Sciences, Raichur, Karnataka, India
- NARAYANASAMY MARAPPA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- KIRAN MARATHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- VISHAL MARATHE N.E.S. Science College, Nanded, Maharashtra, India

- VAIBHAVKUMAR MEHTA Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- HARSH MISTRY Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- JEMISHA MISTRY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- BHAVANA V. MOHITE Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India
- AMI NAIK C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- VRUTUJA NAIK National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- PRADNYA B. NIKAM School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- THAJUDDIN NOORUDDIN Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- DIXITA PANCHAL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- KETANKUMAR J. PANCHAL Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, India
- K. PANDIYAN . ICAR-Central Institute for Research on Cotton Technology, Mumbai, India
- PREETI PARMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- HARSH PATEL P. G. Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, Gujarat, India
- KAMLESH C. PATEL PG Department of Biosciences, Sardar Patel University, Anand, Gujarat, India
- NAFISA PATEL Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- POOJA PATEL . Government Medical College, Surat, Gujarat, India
- PRITTESH PATEL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- RADHIKA PATEL PG Department of Biosciences, Sardar Patel University, Gujarat, India
- SWATI PATEL Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- TARUN KUMAR PATEL Department of Biotechnology, Sant Guru Ghasidas Government P.G. College, Dhamtari, Chattisgarh, India; Department of Biotechnology, Guru Ghasidas Vishwavidyalaya (a Central University), Bilaspur, Chattisgarh, India
- CHANDRASHEKHAR D. PATIL Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- SATISH V. PATIL School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India
- VIKAS S. PATIL University Institute of Chemical Technology, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- MOHINI PIMPALSE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, India
- VIMALKUMAR PRAJAPATI Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India

- PRAVEEN RAHI National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- VIJAYAKUMAR VISHNU RAJA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- B. K. RAJKUMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- SANKET RAY Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- NARENDRA SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- JITENDRA D. SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- A. SANKARANARAYANAN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- SHRUTI SATASHIA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- ABHISHEK SHARMA C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- K. SOWMIYA University of Agricultural Sciences, Raichur, Karnataka, India
- RAHUL K. SURYAWANSHI Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- RASHMI THAKOR Department of Life Sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- VEYSEL TURAN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Bingöl University, Bingöl, Turkey
- VRUSHALI WAGH Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India



Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility

Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil

Abstract

Phytase is one of the identified enzymes which plays a vital role in phosphorous (P) cycle by mobilization of immobilized P from plant material. Although there is various phosphate solubilizing biofertilizer agents reported, they are majorly targeted for mobilizing rock phosphates in soil. The Phosphate solubilizing biofertilizers are required for mobilizing the immobilized form of P from plant source i.e., from phytate. The protocol is based on the principle that specific phytate agar medium contains calcium/sodium phytate, which is utilized as P source by phytase producing microorganism indicated by zone of clearance around the colony. The further quantification is executed by measuring the free phosphate by spectrophotometric method.

Key words Phytase, Phosphate, Phosphorous, Phytate, Macronutrient

1 Introduction

Phosphorous (P) is one of the major macronutrients for plant growth. It contributes approximately 0.2% of plant's dry biomass. It is important component in nucleic acid, ATP, phospholipids, etc. which contribute for plant growth and metabolic activity. P determines many vital activities like development of flower, seed development, germination and maturity, and consequently majorly affects the crop maturity and yield [1]. Besides these, various vital activities like N-fixation in legumes, energy metabolism, synthesis of membranes, photosynthesis, respiration, enzyme regulation, crop quality, and resistance against various biotic and abiotic stress are also subject to P nutrition. Soil has always less utilizable form of P for plant; it warrants the application of P fertilizers in soil. The reaction with metals, absorption and precipitation of inorganic P in soil are major physical phenomenon which cause added fertilizers unavailable to plants. The high accumulation of unavailable phosphate also leads to eutrophication and infertility of soil. In plants, P

Natarajan Amaresan et al. (eds.), Practical Handbook on Agricultural Microbiology,

Springer Protocols Handbooks, https://doi.org/10.1007/978-1-0716-1724-3_45,

[©] The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

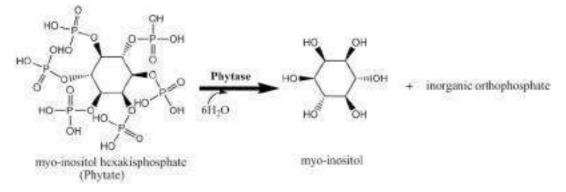


Fig. 1 Mechanism of phytase action for phytate solubilization. (Reproduced from Mohite et al. [3] with permission from Springer Nature)

is in organic form of phytate which forms salts with monovalent and divalent cations like Fe²⁺, Mn²⁺, K⁺, Mg²⁺, and Ca²⁺, making it unavailable for assimilation [2]. Phytases are group of enzymes which naturally make this P available for recycling in nature (Fig. 1). Phytases are classified according to pH activity, catalytic mechanism, source and initiation of site of dephosphorylation of substrate. As per IUPAC-IUBMB, phytase grouped in major three classes, i.e., 3-phytases (EC 3.1.3.8), 6-phytases (EC 3.1.3.26), and 5-phytases (EC 3.1.3.72), which further divided as acid and alkaline phytases on basis of optimum pH [4] (Fig. 2). As per recent reports, microbial phytases are considered as new bioinputs for soil as various fungi and bacteria utilize phytate as substrate and make available in assimilable form to crops or plants [5, 6].

1.1 Principle of the Assay The microorganism having phytase producing capacity will utilize phytate as phosphorus source and indicate by calcium/sodium phosphate dissolution. The phytate dissolution by organic acid will be differentiated by flooding cobalt chloride, ammonium molybdate, and ammonium vanadate solution, these agents reprecipitate the acid solubilized phytases, and the zone due to organic acid dissolution will be disappeared (*see* Note 1).

2 Materials

All the culture media used are from HiMedia, Mumbai including the sodium selenite. Pure grade distilled water was used for dilution.

- 1. Calcium or sodium phytate.
- 2. 0.2 M sodium acetate buffer (pH 4.0).
- 3. Trichloroacetic acid.

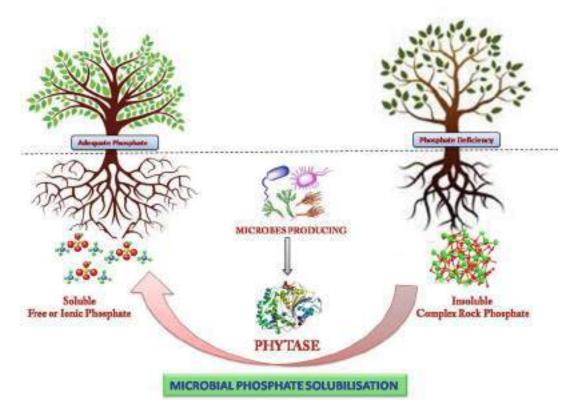


Fig. 2 Microbial phosphate solubilization. (Reproduced from Mohite et al. [3] with permission from Springer Nature)

- 4. 100 mM glycine/HCl buffer (pH 2.5).
- 5. Rhizospheric soil sample from the agricultural crops such as wheat or maize and root nodules of the bean crops.
- 6. Phytase Isolation agar (g/l): Glucose 10.0, Calcium Phytate 1.0, Ammonium nitrate 0.1, KCL 0.5, NH₄NO₃ 0.5, MnSO₄·7H₂O 0.01, and FeSO₄·7H₂O 0.01. Dissolve in slightly hot water, add 3.0 g agar agar powder, and sterilize at 21 °C for 15 min.
- 7. 10 mM sodium phytate and 100 mM glycine/HCl buffer (pH 2.5).
- 8. Confirmation reagents: **Reagent A**—2% (w/v) aqueous cobalt chloride solution and **Reagent B**—10 ml (25% (w/v) ammonium molybdate + 10 ml 0.42% (w/v) ammonium vanadate solution.
- 9. Ammonium molybdate solution (AMS): 1.5 g ammonium molybdate in 5.5% sulfuric acid.
- 10. 2.7% Ferrous sulfate w/v.

11. Reagent for phosphate estimation: 4 ml of AMS + 1 ml of ferrous sulfate solution.

3 Methods

- 1. Take 1.0 g of collected soil sample, add in 9.0 ml sterile saline, mix well by shaking and then further dilute up to ten folds, i.e., 10^{-9} .
- 2. Prepare phytase agar by using pure distilled water in clean 250 ml Erlenmeyer flasks and sterilize them at 121 °C for 20 min under 15 psi pressure.
- 3. Let the media cool at 40 °C, pour it into the sterile petri plates and incubate it at room temperature for 2–3 h, for solidifying. Also prepare control plates of each culture media but without calcium phytate.
- 4. Take loopful of soil suspension 10^{-5} to 10^{-6} dilution tube and streak on phytate agar plates.
- 5. Incubate the plates at room temperature up to approximately 48 h, and observe that the plates for growth appear with hollow zone around colonies.
- 6. Compare the phytate containing plates with the control plates, colonies showing high zone of dissolution, subculture it for further study.
- 7. The confirmation of phytase production, i.e., phytate solubilization due to phytase but not by organic acid or other metabolic product, could be confirmed by counterstaining treatment [7]. In brief, do spot inoculation of potent phytase producer on sterile phytate agar, and allow growing at 48 h as per organism optimum incubation period. After incubating flood plate with reagent A (2% cobalt chloride solution), incubate for 5 min. Remove extra solution from the plate and overlay the petri plates with Reagent B (ammonium molybdate solution and ammonium vanadate). Incubate plates for 5 min. Observe for the zone of clearance (*see* **Note 2**).
- 8. The zone of dissolution after counterstain indicates the real phytate dissolution by phytase.
- 9. Quantitative assay: In quantitative phytase assay, the active culture grow in 0.1% sodium phytate containing medium for 24 h, and centrifuge supernatant as 10,000 lb for 10 min [8]. The crude enzyme in supernatant is collected. Add 1 ml of supernatant broth in 600µl of sodium phytate solution, incubate at 37 °C for 30 min. Terminate the reaction by adding 700µl of 5% TCA solution. After that take 1 ml of this reaction mixture, add 0.5 ml AMS reagent. Thereafter phytase activity is

determined as micromole inorganic phosphate release per min at 700 nm by using standard graph of $K_2H PO_4$ (*see* Note 3).

4 Notes

- 1. The zone after counterstaining avoids false results of phytate dissolution and pseudo phytase activity. The colony shows dissolution on phytase agar but not after counter staining confirm that phytate utilization is not either by metabolic product of media utilization or by organic acid production.
- 2. Prepare counterstain **Reagent A**, i.e., (cobalt chloride solution) and **Reagent B** (ammonium molybdate solution and ammonium vanadate mixture) in distilled water.
- 3. One unit phytase activity (U/mL) was defined as the amount of enzyme required to liberate 1µmol of inorganic phosphate per minute.

References

- Khan AA, Jilani G, Akhtar MS et al (2009) Phosphorus solubilizing bacteria: occurrence, mechanisms and their role in crop production. J Agric Biol Sci 1:48–58
- 2. Raboy V, Dickinson DB (1987) The timing and rate of phytic acid accumulation in developing soybean seeds. Plant Physiol 85(3):841–844
- 3. Mohite BV, Koli SH, Borase HP et al (2019) New edge agricultural bioinputs. In: Singh P, Gupta VK, Prabha R (eds) Microbial interventions in agriculture and environment. Springer, Singapore, pp 353–380
- Jorquera M, Martinez O, Maruyama F (2008) Current and future biotechnological applications of bacterial phytases and phytaseproducing bacteria. Microbes Environ 23:182–191

- Gibson DM (1987) Production of extracellular phytase from *Aspergillus ficuum* on starch media. Biotechnol Lett 9:305–310
- Choi YM, Suh HJ, Kim JM (2001) Purification and properties of extracellular phytase from Bacillus sp. KHU-10. J Protein Chem 20:287–292
- Bae HD, Yanke LJ, Cheng K-J, Selinger LB (1999) A novel staining method for detecting phytase activity. J Microbiol Methods 39:17–22
- Pandey A, Szakacs G, Soccol CR et al (2001) Production, purification and properties of microbial phytases. Bioresour Technol 77 (3):203–214

Springer Protocols

Natarajan Amaresan Prittesh Patel Dhruti Amin *Editors*

Practical Handbook on Agricultural Microbiology



Springer Protocols Handbooks

For further volumes: http://www.springer.com/series/7657 Springer Protocols Handbooks collects a diverse range of step-by-step laboratory methods and protocols from across the life and biomedical sciences. Each protocol is provided in the Springer Protocol format: readily-reproducible in a step-by-step fashion. Each protocol opens with an introductory overview, a list of the materials and reagents needed to complete the experiment, and is followed by a detailed procedure supported by a helpful notes section offering tips and tricks of the trade as well as troubleshooting advice. With a focus on large comprehensive protocol collections and an international authorship, Springer Protocols Handbooks are a valuable addition to the laboratory.

More information about this series at http://www.springer.com/series/8623

Practical Handbook on Agricultural Microbiology

Edited by

Natarajan Amaresan, Prittesh Patel, and Dhruti Amin

Uka Tarsadia University, Surat, Gujarat, India

💥 Humana Press

Editors Natarajan Amaresan Uka Tarsadia University Surat, Gujarat, India

Dhruti Amin Uka Tarsadia University Surat, Gujarat, India Prittesh Patel Uka Tarsadia University Surat, Gujarat, India

ISSN 1949-2448 ISSN 1949-2456 (electronic) Springer Protocols Handbooks ISBN 978-1-0716-1723-6 ISBN 978-1-0716-1724-3 (eBook) https://doi.org/10.1007/978-1-0716-1724-3

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Humana imprint is published by the registered company Springer Science+Business Media, LLC part of Springer Nature.

The registered company address is: 1 New York Plaza, New York, NY 10004, U.S.A.

Preface

Agricultural Microbiology is a part of the microbiology branch dealing with beneficial or harmful microbes associated with either plants or soil. This manual focuses on beneficial microbes dealing with soil fertility, microbial degradation of organic matter, soil nutrient transformations, and biocontrol agents. Nowadays, techniques involved in the study of beneficial microbes in agricultural microbiology toward enhancing global agricultural productivity are in trend. This manual covers a wide range of basic and advanced techniques associated with research on the isolation of agriculturally important microbes, identification, biological nitrogen fixation, microbe-mediated plant nutrient use efficiency, and biological control of plant diseases and pests. Introduction to each protocol explains the role/importance of chemicals involved, uniqueness, and protocol application. A proper understanding of the protocol helps the researchers to manipulate them as per their need.

This book is composed of seven parts with 52 protocol chapters. Parts I and II represent the importance, isolation, and purification methods of agriculturally important microbes and include mineral-solubilizing microbes. Part III deals with phytohormones quantitative protocols directly or indirectly associated with microbes. Parts IV and V provide deep insights into protocols for screening agriculturally important enzymes and compounds related to biocontrol activity. Part VI represents assessment methods of soil microbial activity by soil respiration. The final Part VII deals with protocols for selecting microbial strains for inoculant production and quality control ultimately representing commercial biofertilizers production criteria. This book will help postgraduate students, research scholars, postdoctoral fellows, and teachers belonging to different disciplines of Plant Microbiology and Pathology. Moreover, this manual may also serve as a textbook for undergraduate courses like Techniques on Plant-Microbe Interaction/Biological Control of Plant Diseases/Nutrient Use Efficiency.

Surat, Gujarat, India Surat, Gujarat, India Surat, Gujarat, India Natarajan Amaresan Prittesh Patel Dhruti Amin

Contents

	face stributors	v xi
Par	AT I ISOLATION AND IDENTIFICATION OF AGRICULTURALLY IMPORTANT MICROBES	
1	Methods for Isolation and Identification of Rhizobia	3
2	Isolation of Frankia from Casuarina Root Nodule Narayanasamy Marappa, Dhanasekaran Dharumadurai, and Thajuddin Nooruddin	15
3	Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum Bhavana V. Mohite and Satish V. Patil	25
4	Isolation and Identification of Azospirillum	35
5	Isolation and Identification of <i>Gluconacetobacter diazotrophicus</i>	41
6	Isolation and Identification of Nitrogen Fixing Bacteria: Azoarcus Species	47
7	Isolation and Identification of <i>Derxia</i> Species from the Soil Sample Harshida A. Gamit and Natarajan Amaresan	57
8	Isolation and Characterization of Enterobacter, Klebsiella,and ClostridiumV. Mageshwaran and K. Pandiyan	63
9	Isolation and Characterization of Genus Desulfotomaculum Mobini Pimpalse, Harshida A. Gamit, and Natarajan Amaresan	71
10	Isolation and Identification of Associative Symbiotic N ₂ Fixing Microbes: Desulfovibrio	77
11	Cyanobacteria Dhruti Amin, Abhishek Sharma, and Sanket Ray	85
12	Pseudomonas Sanket Ray and Harsh Patel	93
13	Isolation and Identification of Entomopathogenic <i>Bacillus</i> Species Preeti Parmar, B. K. Rajkumar, and Naresh Butani	99
14	Methylobacterium	111

viii	Contents

15	Isolation and Identification of <i>Beijerinckia</i>	119
16	Isolation of Streptomyces from Soil Sample Vishnu Raja Vijayakumar and Dharumadurai Dhanasekaran	127
17	Isolation and Identification of <i>Trichoderma</i> Spp. from Different Agricultural Samples	131
18	Extraction, Isolation and Culturing of <i>Mycorrhizal</i> Spores from Rhizospheric Soil	145
19	Isolation and Identification of <i>Metarhizium</i>	151
20	Isolation and Identification of Bacteriophage for Biocontrol Mitesh Dwivedi	161
21	Isolation of Bacterivorous Protozoan, Acanthamoeba Spp., as New-Age Agro Bio-Input Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil	173
Par	TT II ISOLATION OF MINERAL SOLUBILIZING MICROBES	
22	Isolation and Screening of Zinc Solubilizing Microbes: As Essential Micronutrient Bio-Inputs for Crops Satish V. Patil, Hemant P. Borase, Jitendra D. Salunkhe, and Rahul K. Suryawanshi	181
23	Isolation and Screening of Mineral Phosphate Solubilizing Microorganisms	187
24	Isolation and Identification of Potassium-Solubilizing Microbes Prittesh Patel and Swati Patel	193
25	Isolation and Identification of Sulfur-Oxidizing Bacteria Vimalkumar Prajapati, Swati Patel, Radhika Patel, and Vaibhavkumar Mehta	197
26	Isolation of Ammonia Oxidizing Bacteria Naresh Butani, Shruti Satashia, Hemanshi Kanpariya, and Preeti Parmar	203
27	Isolation and Identification of Nitrite-Oxidizing Microbes Prittesh Patel, Ami Naik, and Abhishek Sharma	219
28	Isolation and Identification of Iron-Oxidizing Microbes Ami Naik and Pooja Patel	223
29	Isolation and Identification of Phytin Mineralizing Microbes Swati Patel and Prittesh Patel	231
30	Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil	237

31	Isolation of Selenium Biotransforming Microbes as New Age Bioinputs Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil	
Par	T III ESTIMATION OF PHYTOHORMONES BY BENEFICIAL MICROBES	
32	Auxin Dixita Panchal, Jemisha Mistry, and Dhruti Amin	251
33	Abscisic Acid Natarajan Amaresan, A. Sankaranarayanan, and Dhruti Amin	257
34	Cytokinins Jawahar Ganapathy, Jemisha Mistry, and Dhruti Amin	263
35	Ethylene	269
36	Gibberellin Dhruti Amin, Sanket Ray, and Abhishek Sharma	273
37	Brassinosteroids	277
38	Strigolactones: Extraction and Characterization	283
39	Estimation of Jasmonic Acid Using Non-pathogenic Microbes Jasmonic Acid Krutika S. Abhyankar and Monisha Kottayi	289
Par	TT IV SCREENING OF AGRICULTURALLY IMPORTANT ENZYMES	
40	Chitinase Purvesh B. Bharvad and Harsha J. Algotar	301
41	Glucanase Purvesh B. Bharvad and Harsha J. Algotar	309
42	Identification of Cellulase Enzyme Involved in Biocontrol Activity	317
43	Identification of Protease Enzymes Involved in Biocontrol Activity Vimalkumar Prajapati, Swati Patel, Sanket Ray, and Kamlesh C. Patel	323
44	Isolation and Screening of Naringinase Producing Microbes: As Industrial Inputs for Agro Waste Base Enzyme Industry Satish V. Patil, Jitendra D. Salunkhe, and Vishal Marathe	331
45	Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil	337

Par	тV	Identification of Compounds Involved in Biocontrol Activity	
46		rogen Cyanide ti Bhatt and Dhruti Amin	345
47		ophores sa Patel	351
48	for D	tion and Screening of ACC Deaminase-Producing Microbes Prought Stress Management in Crops	361
49		olysaccharides a Chandwani and Natarajan Amaresan	369
Par	T VI	Assessment of Soil Microbial Activity by Soil Respiration	
50		matic Analyses in Soils ar Bilen and Veysel Turan	377
Par	т VII	Selection of Microbial Strains for Inoculant Production and Quality Control	
51		tion of <i>Rhizobium</i> Strain for Inoculum Production	389
52		her Culture, Broth, and Peat Test uti Amin, Sanket Ray, and Vrushali Wagh	395
Ind	ex		405

Contributors

- KRUTIKA S. ABHYANKAR School of Science, Navrachana University, Vadodara, Gujarat, India
- HARSHA J. ALGOTAR . D. L. Patel Science College, Himatnagar, Gujarat, India
- NATARAJAN AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- DHRUTI AMIN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- HIMANSHU BARIYA Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- PURVESH B. BHARVAD . D. L. Patel Science College, Himatnagar, Gujarat, India
- KHYATI BHATT Post Graduate Department of Biosciences, Sardar Patel University, Gujarat, India
- SERDAR BILEN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Atatürk University, Erzurum, Turkey
- HEMANT P. BORASE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- NARESH BUTANI Department of Microbiology, Faculty of Science, Sarvajanik University, Surat, India
- SAPNA CHANDWANI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- SHREYA DESAI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- DHANASEKARAN DHARUMADURAI Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- MITESH DWIVEDI C. G. Bhakta Institute of Biotechnology, Faculty of Science, Uka Tarsadia University, Surat, Gujarat, India
- HARSHIDA A. GAMIT C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- JAWAHAR GANAPATHY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- TEJAS GOHIL Sabarmati Ashram Gaushala, Kheda, Gujarat, India
- HEMANSHI KANPARIYA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- MONISHA KOTTAYI School of Science, Navrachana University, Vadodara, Gujarat, India
- V. MAGESHWARAN ICAR-National Bureau of Agriculturally Important Microorganisms, Mau, Uttar Pradesh, India
- MAHADEVASWAMY University of Agricultural Sciences, Raichur, Karnataka, India
- NARAYANASAMY MARAPPA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- KIRAN MARATHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- VISHAL MARATHE N.E.S. Science College, Nanded, Maharashtra, India

- VAIBHAVKUMAR MEHTA Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- HARSH MISTRY Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- JEMISHA MISTRY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- BHAVANA V. MOHITE Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India
- AMI NAIK C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- VRUTUJA NAIK National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- PRADNYA B. NIKAM School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- THAJUDDIN NOORUDDIN Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- DIXITA PANCHAL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- KETANKUMAR J. PANCHAL Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, India
- K. PANDIYAN . ICAR-Central Institute for Research on Cotton Technology, Mumbai, India
- PREETI PARMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- HARSH PATEL P. G. Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, Gujarat, India
- KAMLESH C. PATEL PG Department of Biosciences, Sardar Patel University, Anand, Gujarat, India
- NAFISA PATEL Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- POOJA PATEL . Government Medical College, Surat, Gujarat, India
- PRITTESH PATEL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- RADHIKA PATEL PG Department of Biosciences, Sardar Patel University, Gujarat, India
- SWATI PATEL Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- TARUN KUMAR PATEL Department of Biotechnology, Sant Guru Ghasidas Government P.G. College, Dhamtari, Chattisgarh, India; Department of Biotechnology, Guru Ghasidas Vishwavidyalaya (a Central University), Bilaspur, Chattisgarh, India
- CHANDRASHEKHAR D. PATIL Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- SATISH V. PATIL School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India
- VIKAS S. PATIL University Institute of Chemical Technology, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- MOHINI PIMPALSE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, India
- VIMALKUMAR PRAJAPATI Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India

- PRAVEEN RAHI National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- VIJAYAKUMAR VISHNU RAJA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- B. K. RAJKUMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- SANKET RAY Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- NARENDRA SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- JITENDRA D. SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- A. SANKARANARAYANAN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- SHRUTI SATASHIA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- ABHISHEK SHARMA C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- K. SOWMIYA University of Agricultural Sciences, Raichur, Karnataka, India
- RAHUL K. SURYAWANSHI Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- RASHMI THAKOR Department of Life Sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- VEYSEL TURAN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Bingöl University, Bingöl, Turkey
- VRUSHALI WAGH Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India



Isolation of Selenium Biotransforming Microbes as New Age Bioinputs

Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil

Abstract

Selenium (Se) has a very narrow gap between its toxicity and benefits to the different forms of life which makes it as an essential micronutrient for living creatures including its importance in plant growth. It is highly soluble having toxic form in the environment such as the oxyanions; selenate and selenite needed to be transformed to the less toxic elemental selenium which has wide number of benefits to the ecosystem, if provided in appropriate required amount. Till now most of the microorganisms have been studied for biotransformation of selenite by using them as electron acceptor in the respiratory mechanism and ultimately reduced to elemental selenium in the form of nanoparticles. Despite of this, many of the microbes are unexplored and the Se-nanoparticles still less studied for their applications. This chapter explains a simple method for isolating the selenium biotransforming microorganisms from nearby soil sample into the red elemental Se in its nano form.

Key words Selenium, Selenite, Thioredoxin reductase, Selenobacter, Antioxidant

1 Introduction

Selenium (Se) has been proven as an essential micronutrient for almost all living organisms, which is also found important for plants in various aspects. Its presence in the environment ranges from inorganic oxyanions to organic selenium containing amino acids. In human beings, various enzymes such as glutathione peroxidase, thioredoxin reductase, iodothyronine deiodinase, and formate dehydrogenase have selenium as their important component and thus function as an antioxidant, preventing tumors, metabolism, other therapeutic uses and even in reproduction [1, 2]. In the environment, Se exists as highly soluble oxyanions such as Selenate (SeO₄⁻²) and Selenite (SeO₃⁻²) which are toxic for humans, hence

Satish V. Patil and Vikas S. Patil contributed equally to this work.

Natarajan Amaresan et al. (eds.), Practical Handbook on Agricultural Microbiology,

Springer Protocols Handbooks, https://doi.org/10.1007/978-1-0716-1724-3_31,

[©] The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

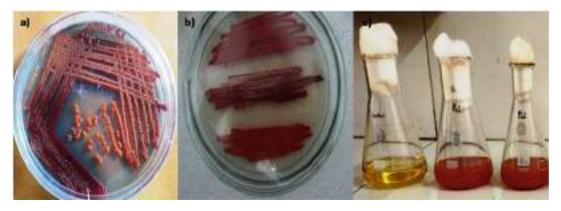


Fig. 1 (a and b) Isolated colonies of selenium transforming bacteria showing brick red color colonies; (c) Transformation of selenium in enrichment medium showing color change from yellow (control) to brick red (test isolates, PI1 and PI2) (Source: Satish V Patil's Laborotory KBC NMU, Jalgaon)

transformed into less toxic and less soluble elemental form (Se^0) by different microbial species through Se cycle. Many of the plants are seen to accumulate these Se oxyanions from the soil and form selenocysteine or may carry formation of selenomethionine, further converting it into volatile dimethylselenide (DMSe). Though Se is not essential for plant growth, it helps the crops to withstand the drought conditions, protects against oxidative stress and sometimes from the pests too [3].

As the distribution of selenium is uneven in the environment, it becomes important to provide it as less toxic and less soluble Se⁰, mostly in colloidal form for the living creatures. This can be achieved by isolating the microbes from different areas especially the agricultural rhizospheric soil or also from the endophytic microbes within the roots, as plants have the potential of accumulating the Se at any available concentrations from the soil. The agricultural rhizospheric soil and nodule samples are preferred considering the compatibility of synthesized Se-nanoparticles for further applications in agricultural practices and also most of them will be non-pathogenic [3-5]. When the sample is enriched in the growth medium supplied with selenite or selenate as a Se source, within 24-48 h the visual brick red color starts appearing along with the growth which signifies the transformation of Se by the respective organism (Fig. 1). This transformation relies on the growth phase and also the initial Se concentration in the medium, as reported earlier. Several reports declare this transformation as synthesis of the Se⁰ nanomaterial either extra- or intracellular by the microbes [6–8].

2 Materials

All the culture media used are from HiMedia (Mumbai, India) including the sodium selenite. Pure grade distilled water was used for dilution.

- 1. Rhizospheric soil sample from the agricultural crops such as wheat or maize and root nodules of the bean crops.
- 2. Sterile nutrient agar media for bacterial isolation.
- 3. Sterile Sabouraud dextrose agar for fungal isolation.
- 4. Sterile yeast extract mannitol agar (YEMA) to culture root nodule bacteria (*Rhizobia*).
- 5. Sterile saline tubes (10 tubes) for serial dilution of the soil samples.
- 6. Sodium selenite (Na₂SeO₃) (MW-172.94 g/mol).

3 Methods

- 1. Take 1.0 g of collected soil sample, add in 9.0 ml sterile saline, mix well by shaking and then further dilute up to tenfolds, i.e., 10^{-9} (*See* Note 2).
- 2. Prepare nutrient agar media, Sabouraud dextrose agar, and yeast extract mannitol agar by using pure distilled water in clean 250 ml Erlenmeyer flasks and sterilize them at 121 °C for 20 min under 15 psi pressure in the autoclave.
- Let the media cool at 40 °C, and then add sodium selenite in nutrient agar media as well as in Sabouraud dextrose agar (5 mM per volume of medium). Rotate flask clockwise and anticlockwise for uniform distribution of selenite (*See* Note 1).
- 4. Pour it into the sterile petri plates and allow solidifying at room temperature for 2–3 h. Also prepare control media plates of each culture media without sodium selenite.
- 5. Take loopful of soil suspension from 10^{-5} to 10^{-6} dilution tube and streak on selenite containing nutrient agar media and Sabouraud dextrose agar plates.
- 6. Incubate the plates at room temperature up to approximately 48 h; observe the plates for growth changed to red color.
- 7. Compare the sodium selenite containing plates with the control plates to examine the exact biotransformation of Se and formation of red colored colonies.
- 8. These isolates can further be inoculated in liquid enrichment broth medium supplemented with sodium selenite at 5 mM

concentration for selenite biotransformation and conversion to the elemental Se-nanoparticles which can be optimized and characterized further.

- 9. The synthesized nanoparticles are separated by centrifugation at 10,000 rpm for 5–8 min, and collecting the pellet and purifying with double distilled water and ethanol by washing three times to remove the untreated sodium selenite or other impurities. The nanosuspensions then pass through the dialysis membrane (6000–8000 KD cutoffs). The purified particles then lyophilized and stored at low temperatures which will be used for characterization by various spectroscopic methods.
- 10. Se-nanoparticle is characterized by UV–Vis absorption spectrophotometer by recording the spectra (200–500 nm).
- 11. The lyophilized powder is used for further particle characterization such as particle size, shape, and crystallinity by using scanning electron microscopy, transmission electron microscopy, dynamic light scattering (DLS), and X-ray diffraction.

4 Notes

- 1. Avoid adding sodium selenite into the media preparation before autoclaving as this may result into precipitation of the salt and forming red color due to heat and pressure, prior to the addition of microbial culture sample.
- 2. For the rhizospheric endophytes and the rhizobia, surface sterilize the material with 70% ethanol or 0.01% mercuric chloride washing. Then crush the sample in sterile saline, further dilute and streak the specific dilution on sterile YEMA or endophyte isolation agar medium.

Acknowledgments

We kindly acknowledge the Prof. Paola Andrea Duran, Universidad de La Frontera, Temuco, Chile for her inspiration and support.

References

- 1. Geoffrion LD, Hesabizadeh T, Medina-Cruz D et al (2020) Naked selenium nanoparticles for antibacterial and anticancer treatments. ACS Omega 5(6):2660–2669
- 2. Mehdi Y, Hornick JL, Istasse L et al (2013) Selenium in the environment, metabolism and

involvement in body functions. Molecules 18 (3):3292-3311

 Quinn CF, El Mehdawi AF, Pilon-Smits EAH (2017) Ecology of selenium in plants. In: Pilon-Smits E, Winkel L, Lin ZQ (eds) Selenium in plants. Plant ecophysiology, vol 11. Springer, Cham, pp 177–188

- Nancharaiah YV, Lens PNL (2015) Ecology and biotechnology of selenium-respiring bacteria. Microbiol Mol Biol Rev 79(1):61–80
- Patel PJ, Trivedi GR, Shah RK et al (2018) Selenorhizobacteria: as biofortification tool in sustainable agriculture. Biocatal Agric Biotechnol 14:198–203
- 6. Afzal B, Yasin D, Husain S et al (2019) Screening of cyanobacterial strains for the selenium nanoparticles synthesis and their anti-oxidant activity. Biocatal Agric Biotechnol 21:101307
- 7. Tugarova AV, Mamchenkova PV, Khanadeev VA et al (2020) Selenite reduction by the rhizobacterium Azospirillum brasilense, synthesis of extracellular selenium nanoparticles and their characterisation. New Biotechnol 58:17–24
- Wang Y, Shu X, Zhou Q et al (2018) Selenite reduction and the biogenesis of selenium nanoparticles by Alcaligenes faecalis Se03 isolated from the gut of monochamus alternatus (Coleoptera: Cerambycidae). Int J Mol Sci 19 (9):1–18

Springer Protocols

Natarajan Amaresan Prittesh Patel Dhruti Amin *Editors*

Practical Handbook on Agricultural Microbiology



Springer Protocols Handbooks

For further volumes: http://www.springer.com/series/7657 Springer Protocols Handbooks collects a diverse range of step-by-step laboratory methods and protocols from across the life and biomedical sciences. Each protocol is provided in the Springer Protocol format: readily-reproducible in a step-by-step fashion. Each protocol opens with an introductory overview, a list of the materials and reagents needed to complete the experiment, and is followed by a detailed procedure supported by a helpful notes section offering tips and tricks of the trade as well as troubleshooting advice. With a focus on large comprehensive protocol collections and an international authorship, Springer Protocols Handbooks are a valuable addition to the laboratory.

More information about this series at http://www.springer.com/series/8623

Practical Handbook on Agricultural Microbiology

Edited by

Natarajan Amaresan, Prittesh Patel, and Dhruti Amin

Uka Tarsadia University, Surat, Gujarat, India

💥 Humana Press

Editors Natarajan Amaresan Uka Tarsadia University Surat, Gujarat, India

Dhruti Amin Uka Tarsadia University Surat, Gujarat, India Prittesh Patel Uka Tarsadia University Surat, Gujarat, India

ISSN 1949-2448 ISSN 1949-2456 (electronic) Springer Protocols Handbooks ISBN 978-1-0716-1723-6 ISBN 978-1-0716-1724-3 (eBook) https://doi.org/10.1007/978-1-0716-1724-3

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Humana imprint is published by the registered company Springer Science+Business Media, LLC part of Springer Nature.

The registered company address is: 1 New York Plaza, New York, NY 10004, U.S.A.

Preface

Agricultural Microbiology is a part of the microbiology branch dealing with beneficial or harmful microbes associated with either plants or soil. This manual focuses on beneficial microbes dealing with soil fertility, microbial degradation of organic matter, soil nutrient transformations, and biocontrol agents. Nowadays, techniques involved in the study of beneficial microbes in agricultural microbiology toward enhancing global agricultural productivity are in trend. This manual covers a wide range of basic and advanced techniques associated with research on the isolation of agriculturally important microbes, identification, biological nitrogen fixation, microbe-mediated plant nutrient use efficiency, and biological control of plant diseases and pests. Introduction to each protocol explains the role/importance of chemicals involved, uniqueness, and protocol application. A proper understanding of the protocol helps the researchers to manipulate them as per their need.

This book is composed of seven parts with 52 protocol chapters. Parts I and II represent the importance, isolation, and purification methods of agriculturally important microbes and include mineral-solubilizing microbes. Part III deals with phytohormones quantitative protocols directly or indirectly associated with microbes. Parts IV and V provide deep insights into protocols for screening agriculturally important enzymes and compounds related to biocontrol activity. Part VI represents assessment methods of soil microbial activity by soil respiration. The final Part VII deals with protocols for selecting microbial strains for inoculant production and quality control ultimately representing commercial biofertilizers production criteria. This book will help postgraduate students, research scholars, postdoctoral fellows, and teachers belonging to different disciplines of Plant Microbiology and Pathology. Moreover, this manual may also serve as a textbook for undergraduate courses like Techniques on Plant-Microbe Interaction/Biological Control of Plant Diseases/Nutrient Use Efficiency.

Surat, Gujarat, India Surat, Gujarat, India Surat, Gujarat, India Natarajan Amaresan Prittesh Patel Dhruti Amin

Contents

	face stributors	v xi
Par	AT I ISOLATION AND IDENTIFICATION OF AGRICULTURALLY IMPORTANT MICROBES	
1	Methods for Isolation and Identification of Rhizobia	3
2	Isolation of Frankia from Casuarina Root Nodule Narayanasamy Marappa, Dhanasekaran Dharumadurai, and Thajuddin Nooruddin	15
3	Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum Bhavana V. Mohite and Satish V. Patil	25
4	Isolation and Identification of Azospirillum	35
5	Isolation and Identification of <i>Gluconacetobacter diazotrophicus</i>	41
6	Isolation and Identification of Nitrogen Fixing Bacteria: Azoarcus Species	47
7	Isolation and Identification of <i>Derxia</i> Species from the Soil Sample Harshida A. Gamit and Natarajan Amaresan	57
8	Isolation and Characterization of Enterobacter, Klebsiella,and ClostridiumV. Mageshwaran and K. Pandiyan	63
9	Isolation and Characterization of Genus Desulfotomaculum Mobini Pimpalse, Harshida A. Gamit, and Natarajan Amaresan	71
10	Isolation and Identification of Associative Symbiotic N ₂ Fixing Microbes: Desulfovibrio	77
11	Cyanobacteria Dhruti Amin, Abhishek Sharma, and Sanket Ray	85
12	Pseudomonas Sanket Ray and Harsh Patel	93
13	Isolation and Identification of Entomopathogenic <i>Bacillus</i> Species Preeti Parmar, B. K. Rajkumar, and Naresh Butani	99
14	Methylobacterium	111

viii	Contents

15	Isolation and Identification of <i>Beijerinckia</i>	119
16	Isolation of Streptomyces from Soil Sample Vishnu Raja Vijayakumar and Dharumadurai Dhanasekaran	127
17	Isolation and Identification of <i>Trichoderma</i> Spp. from Different Agricultural Samples	131
18	Extraction, Isolation and Culturing of <i>Mycorrhizal</i> Spores from Rhizospheric Soil	145
19	Isolation and Identification of <i>Metarhizium</i>	151
20	Isolation and Identification of Bacteriophage for Biocontrol Mitesh Dwivedi	161
21	Isolation of Bacterivorous Protozoan, Acanthamoeba Spp., as New-Age Agro Bio-Input Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil	173
Par	TT II ISOLATION OF MINERAL SOLUBILIZING MICROBES	
22	Isolation and Screening of Zinc Solubilizing Microbes: As Essential Micronutrient Bio-Inputs for Crops Satish V. Patil, Hemant P. Borase, Jitendra D. Salunkhe, and Rahul K. Suryawanshi	181
23	Isolation and Screening of Mineral Phosphate Solubilizing Microorganisms	187
24	Isolation and Identification of Potassium-Solubilizing Microbes Prittesh Patel and Swati Patel	193
25	Isolation and Identification of Sulfur-Oxidizing Bacteria Vimalkumar Prajapati, Swati Patel, Radhika Patel, and Vaibhavkumar Mehta	197
26	Isolation of Ammonia Oxidizing Bacteria Naresh Butani, Shruti Satashia, Hemanshi Kanpariya, and Preeti Parmar	203
27	Isolation and Identification of Nitrite-Oxidizing Microbes Prittesh Patel, Ami Naik, and Abhishek Sharma	219
28	Isolation and Identification of Iron-Oxidizing Microbes Ami Naik and Pooja Patel	223
29	Isolation and Identification of Phytin Mineralizing Microbes Swati Patel and Prittesh Patel	231
30	Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil	237

31	Isolation of Selenium Biotransforming Microbes as New Age Bioinputs Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil	
Par	T III ESTIMATION OF PHYTOHORMONES BY BENEFICIAL MICROBES	
32	Auxin Dixita Panchal, Jemisha Mistry, and Dhruti Amin	251
33	Abscisic Acid Natarajan Amaresan, A. Sankaranarayanan, and Dhruti Amin	257
34	Cytokinins Jawahar Ganapathy, Jemisha Mistry, and Dhruti Amin	263
35	Ethylene Ketankumar J. Panchal and Dhruti Amin	269
36	Gibberellin Dhruti Amin, Sanket Ray, and Abhishek Sharma	273
37	Brassinosteroids	277
38	Strigolactones: Extraction and Characterization	283
39	Estimation of Jasmonic Acid Using Non-pathogenic Microbes Jasmonic Acid Krutika S. Abhyankar and Monisha Kottayi	289
Par	TT IV SCREENING OF AGRICULTURALLY IMPORTANT ENZYMES	
40	Chitinase Purvesh B. Bharvad and Harsha J. Algotar	301
41	Glucanase Purvesh B. Bharvad and Harsha J. Algotar	309
42	Identification of Cellulase Enzyme Involved in Biocontrol Activity	317
43	Identification of Protease Enzymes Involved in Biocontrol Activity Vimalkumar Prajapati, Swati Patel, Sanket Ray, and Kamlesh C. Patel	323
44	Isolation and Screening of Naringinase Producing Microbes: As Industrial Inputs for Agro Waste Base Enzyme Industry Satish V. Patil, Jitendra D. Salunkhe, and Vishal Marathe	331
45	Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil	337

Par	тV	Identification of Compounds Involved in Biocontrol Activity	
46		rogen Cyanide ti Bhatt and Dhruti Amin	345
47		ophores sa Patel	351
48	for D	tion and Screening of ACC Deaminase-Producing Microbes Prought Stress Management in Crops	361
49		olysaccharides a Chandwani and Natarajan Amaresan	369
Par	T VI	Assessment of Soil Microbial Activity by Soil Respiration	
50		matic Analyses in Soils ar Bilen and Veysel Turan	377
Par	т VII	Selection of Microbial Strains for Inoculant Production and Quality Control	
51		tion of <i>Rhizobium</i> Strain for Inoculum Production	389
52		her Culture, Broth, and Peat Test uti Amin, Sanket Ray, and Vrushali Wagh	395
Ind	ex		405

Contributors

- KRUTIKA S. ABHYANKAR School of Science, Navrachana University, Vadodara, Gujarat, India
- HARSHA J. ALGOTAR . D. L. Patel Science College, Himatnagar, Gujarat, India
- NATARAJAN AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- DHRUTI AMIN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- HIMANSHU BARIYA Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- PURVESH B. BHARVAD . D. L. Patel Science College, Himatnagar, Gujarat, India
- KHYATI BHATT Post Graduate Department of Biosciences, Sardar Patel University, Gujarat, India
- SERDAR BILEN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Atatürk University, Erzurum, Turkey
- HEMANT P. BORASE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- NARESH BUTANI Department of Microbiology, Faculty of Science, Sarvajanik University, Surat, India
- SAPNA CHANDWANI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- SHREYA DESAI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- DHANASEKARAN DHARUMADURAI Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- MITESH DWIVEDI C. G. Bhakta Institute of Biotechnology, Faculty of Science, Uka Tarsadia University, Surat, Gujarat, India
- HARSHIDA A. GAMIT C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- JAWAHAR GANAPATHY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- TEJAS GOHIL Sabarmati Ashram Gaushala, Kheda, Gujarat, India
- HEMANSHI KANPARIYA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- MONISHA KOTTAYI School of Science, Navrachana University, Vadodara, Gujarat, India
- V. MAGESHWARAN ICAR-National Bureau of Agriculturally Important Microorganisms, Mau, Uttar Pradesh, India
- MAHADEVASWAMY University of Agricultural Sciences, Raichur, Karnataka, India
- NARAYANASAMY MARAPPA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- KIRAN MARATHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- VISHAL MARATHE N.E.S. Science College, Nanded, Maharashtra, India

- VAIBHAVKUMAR MEHTA Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- HARSH MISTRY Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- JEMISHA MISTRY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- BHAVANA V. MOHITE Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India
- AMI NAIK C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- VRUTUJA NAIK National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- PRADNYA B. NIKAM School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- THAJUDDIN NOORUDDIN Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- DIXITA PANCHAL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- KETANKUMAR J. PANCHAL Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, India
- K. PANDIYAN . ICAR-Central Institute for Research on Cotton Technology, Mumbai, India
- PREETI PARMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- HARSH PATEL P. G. Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, Gujarat, India
- KAMLESH C. PATEL PG Department of Biosciences, Sardar Patel University, Anand, Gujarat, India
- NAFISA PATEL Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- POOJA PATEL . Government Medical College, Surat, Gujarat, India
- PRITTESH PATEL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- RADHIKA PATEL PG Department of Biosciences, Sardar Patel University, Gujarat, India
- SWATI PATEL Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- TARUN KUMAR PATEL Department of Biotechnology, Sant Guru Ghasidas Government P.G. College, Dhamtari, Chattisgarh, India; Department of Biotechnology, Guru Ghasidas Vishwavidyalaya (a Central University), Bilaspur, Chattisgarh, India
- CHANDRASHEKHAR D. PATIL Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- SATISH V. PATIL School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India
- VIKAS S. PATIL University Institute of Chemical Technology, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- MOHINI PIMPALSE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, India
- VIMALKUMAR PRAJAPATI Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India

- PRAVEEN RAHI National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- VIJAYAKUMAR VISHNU RAJA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- B. K. RAJKUMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- SANKET RAY Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- NARENDRA SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- JITENDRA D. SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- A. SANKARANARAYANAN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- SHRUTI SATASHIA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- ABHISHEK SHARMA C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- K. SOWMIYA University of Agricultural Sciences, Raichur, Karnataka, India
- RAHUL K. SURYAWANSHI Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- RASHMI THAKOR Department of Life Sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- VEYSEL TURAN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Bingöl University, Bingöl, Turkey
- VRUSHALI WAGH Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India



Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops

Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil

Abstract

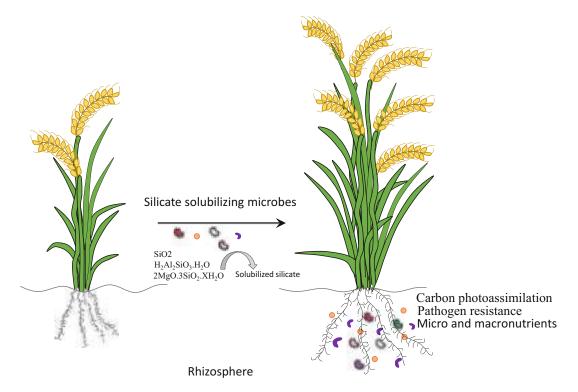
Pertaining to the importance of silica as fertilizer for various crops, silica solubilizers become new component of biofertilizers for sustainable crop and fertility management. Importance of silica metal has been identified for tolerance of biotic and abiotic factors, growth enhancement, etc. Soil is rich source of silica however; it is majorly available in un-utilizable form of silicates. Therefore, silicate solubilizing microbes are essential part of micronutrient management program. The present protocol describes method of isolation and screening of silicate solubilizers from various sources. The application of pH indicator dyes in medium will also help to find out microbe-mediated mechanism of silicate solubilization. Medium with differential sources of immobilized or polymeric silicates will allow isolation of potent and versatile silicate solubilizers useful as modern bioinputs for crops.

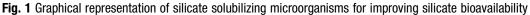
Key words Silicate, Rice, Drought, Organic acid, Ammonia

1 Introduction

The silicon (Si) is one of the abundant component of soil. Si was previously considered as nonessential element for crop development, but recent findings proved significance of Si to induce tolerance of biotic and abiotic factors and growth enhancement by carbon photoassimilation in crops like rice, maize, wheat, etc. [1–3] (Fig. 1) and play vital role in development of pathogen resistance in silicon accumulating crops like rice. Various diseases of rice crop, rice blast, powdery mildew, brown spot, stem rot and sheath brown rot on rice were found to be successfully managed by new bioinputs such as silicon fertilizer application [5–7]. Additionally, Si is also known to facilitate the availability of other micro- and macronutrients such as phosphorus, potassium, zinc, and copper [4] (Fig. 1).

Natarajan Amaresan et al. (eds.), *Practical Handbook on Agricultural Microbiology*, Springer Protocols Handbooks, https://doi.org/10.1007/978-1-0716-1724-3_30, © The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022





Silicon is abundantly available in soil but in immobilized or mineral form which is unavailable for direct assimilation by plant. The need of plant is fulfilled by extraneous addition of silicon fertilizers, but at majority times silicon fulfillment is done by adding industrial slag, which causes other heavy metal contamination in farm soil. Besides these, silicon fertilizers in market such as bentonite, mica feldspars, and diatomaceous earth are present but they contain very little bioavailable Si and so require application of higher dose. Currently, potassium silicate is applied as popular silicon fertilizer but it is too expensive, and may cause phytotoxicity, eutrophication, etc. The application of Si solubilizing biofertilizers proved most ecofriendly, cheap, and significant to fulfill the crop Si need.

Silicon solubilization is first reported by Aleksandrov et al. [8] during their work on microbial phosphoric acid liberation by silicate bacteria. Various microbes were reported for silicon depolymerizing/solubilizing potential, i.e., Rhizobium, Burkholderia, *Pseudomonas* sp. [9]. Some studies proved that fungi actively solubilize the silicate mineral from rock mineral, e.g., *Fusarium oxysporum* [10, 11].

The naturally occurring silicon in the form of soil minerals is majorly converted by the process of withering into the monosilicic acid which is assimilated by plant roots. The withering of silicon



Fig. 2 (a) Isolation of silicate solubilizers on silicate agar, (b) silicate solubilization by bacteria, (c) silicate solubilization by fungi, (d) confirmation silicate solubilization mode of action by pH indicator silicate (Phenol red). (Isolates From Dr. SV Patil, Laboratory School of Life Sciences, KBC, NMU, Jalgaon)

minerals take place by various ways, i.e., by organic and inorganic acids, actions of divalent cations, by nucleophilic attacks of alkali compounds, etc. Majority of these products are produced by microbial action.

Recent studies revealed that the foliar applications of silicate solutions stimulate growth of wheat cereals, soybean, rapeseed, sugar beet, potato, meadows, berries and vegetables, as well as orchard and ornamental plants in diverse abiotic stress like drought by stimulating chlorophyll production, maintaining water content and cellular membrane integrity [12].

1.1 Principle Plant rhizosphere is a rich source of silicate solubilizers. Screening of Assay of silicate solubilizers uses a classical approach of cultivating test microorganisms isolated from plant rhizosphere on complex silicate mixture containing minimal medium and isolating microbes capable of making zone of clearance (Fig. 2). pH indicator dye can be used to detect acid or alkali based microbial mechanisms of silicate solubilization. The solubilized zinc and its presence in plant can further be estimated through colorimetric, coupled plasma mass spectrometry or absorption/emission spectrometry.

2 Materials

- 1. Rhizospheric soil sample from the agricultural crops such as rice, wheat or maize and root nodules of the bean crops (*see* **Note 2**).
- 2. Sterile saline tubes (10 tubes) for serial dilution of the soil samples.
- 3. Following media will be needed for isolations of different silicate solubilizers
 - (a) Sterile Silicate Agar for Bacteria: (g/L) Magnesium sulfate 0.1, Calcium carbonate 0.1, magnesium trisilicate $[Mg_2O_8Si_3]$ 1.0 or Potassium alumino silicate 1.5,

Glucose 1.0, Ferric chloride 0.005, Calcium phosphate 1.0, ammonium sulfate 0.2, Agar 30 g, Final pH (at 25 °C) 7.2 \pm 0.2 + 5 ml of 10 mg% Phenol red or 20 mg% bromothymol blue per liter (*see* **Note 1**).

- (b) Sterile Silicate Agar for Fungi: (g/L) Glucose 0.5, Yeast extract 0.5, Ammonium sulfate 0.4, magnesium trisilicate $[Mg_2O_8Si_3]$, Potassium alumino silicate 1.5, Calcium phosphate 1.0, Agar 30.0, Final pH (at 25 °C) 6.5 \pm 0.2 + 5 ml of 10 mg% Phenol red or 20 mg% bromothymol blue per liter (*see* Note 1).
- (c) Sterile Silicate Agar for Actinomycetes: (g/L) Magnesium sulfate 0.1, calcium carbonate 0.1, potassium alumino silicate 1.5, Starch 2 g ferric chloride, asparagine 0.01, Casein 0.02 g, calcium phosphate 1.0 g, ammonium sulfate 0.2, Agar 30 g, final pH (at 25 °C) $7.2 \pm 0.2 + 5$ ml of 10 mg% Phenol red or 20 mg% bromothymol blue per liter (*see* Note 1). (d) Aleksandrov medium used for potassium solubilizers is also applicable for isolation of silicate solubilizers.

3 Methods

- 1. Take 1 g of collected soil sample, add in 9 ml of sterile saline, mix well by shaking and then serially dilute up to tenfold, i.e., 10^{-9} .
- 2. Prepare silicate agar for bacteria, fungi, and actinomycetes by using pure distilled water in clean 250 ml Erlenmeyer flasks and sterilize them at 121 °C for 20 min under 15 psi pressure.
- 3. Let the media to cool at 40 $^{\circ}$ C, and mix well the settled silicate and carbonate.
- 4. Pour it into the sterile petri plates and allow solidifying at room temperature for 2–3 h. Also prepare control media plates of each culture media without silicate.
- 5. Take loopful of soil suspension from 10^{-5} to 10^{-6} dilution tube and streak on selenite containing silicate agar plates.
- 6. Incubate the plates at room temperature approximately up to 48 h; observe the plates for growth and zone of clearance around colony (Fig. 2).
- 7. Compare the silicate containing plates with the control plates to examine the solubilization of Si and thereby formation of zone around the colonies.
- 8. These isolates can be further purified in liquid enrichment medium supplemented with magnesium silicate at low concentration of silicate. Isolate specific media required for the

conversion of silicate into depolymerized form of soluble silicon/nanoparticles can be further optimized and the silicon particles can be characterized for solubilization properties (Fig. 2).

- For confirmation of solubilizing mechanism, use pH indicator in the medium. Add 3–5 ml of 10 mg% phenol red or 10 mg% bromophenol blue and mix well (*see* Note 3).
- 10. The color change will indicate whether solubilization might be due to acid or alkali.
- 11. Estimation of Solubilized silica: Centrifuge the broth medium of silica solubilizers, take 5 ml of supernatant, mix with 15 ml 2.5% boric acid and 5 ml 54% ammonium molybdate, incubate for 5 min and add 1 ml of 0.5% ascorbic acid solution, mix the solution by vortex and measure absorbance at 650 nm. The concentration of silicate could be determined by the standard graph of SiO₂ by spectrophotometric method.
- 12. Bioassay and analysis of Si content in test plant: Plant ten rice *oryza sativa/Triticum aestivum*, sugar cane, maize, wheat seedling in the 5×3 cm pots, filled with sterilized black cotton soil and one pot can be inoculated with silicate solubilizers inoculum. The uninoculated pots should be considered as control which should be allowed to grow for 3 weeks, then uproot the plants, wash with distilled water and dry it at room temperature and measure weight of control and test plant. Dry the plant at 60 °C for 3 h and use for silicate analysis in plant (*see* Notes 4 and 5).
- 13. Analysis of Si content of the plant: Ground the 0.5–1.0 g of dried biomass of control and test plant. As per method described by Kaya et al [13] take 0.5 g of the lyophilized crushed powder and soak in 0.5 M HCl for 1 h, then wash by distilled water and dry in oven. Add the dried powder in the mixture of nitric acid, sulfuric acid, and perchloric acid (10:1:4 v/v/v) keep it for 1 h, then this digested sample can be analyzed by coupled plasma mass spectrometry (*see* Notes 4 and 5).

4 Notes

- 1. For isolation of silicate solubilizers, various polymeric silica sources can be used in medium such as quartz, bentonite, magnesium trisilicate), aluminum silicate, calcium fuller's earth, kaolin potassium alumino silicate), potassium, aluminum and calcium silicates, phyllosilicates, etc.
- 2. For the rhizospheric endophytes and the rhizobia, surface sterilize the material with 70% ethanol or 0.01% mercuric chloride. Then crush the sample in sterile saline, further dilute and

streak the specific dilution on sterile silicate solubilization agar medium.

- 3. Add pH indicators like bromocresol green, phenol red, bromophenol blue, etc. in medium to know mechanism of solubilization of silicates. Prepare pH indicator solution in 70% ethanol to avoid contaminations. pH indicators can be added after sterilization.
- 4. Many times these depolymerized or solubilized Si particles are in nanoform as hence as nano silicates, which have various significant potential for various industrial and agriculture applications. The nanoforms are characterized by various spectroscopic methods. Si-nanoparticle is characterized by UV–Vis absorption spectrophotometer by recording the spectra (200–500 nm).
- 5. The lyophilized powder is used for further particle characterization such as particle size, shape, and crystallinity by using scanning electron microscopy, transmission electron microscopy, dynamic light scattering (DLS), and X-ray diffraction.

References

- Rudnick RL, Gao S (2003) Composition of the continental crust. In: Holland HD, Turekian KK (eds) Treatise on geochemistry, vol 3. Elsevier Science, New York, pp 1–64
- Gascho GJ (2001) Silicon sources for agriculture. In: Datnoff LE, Snyder GH, Korndorfer GH (eds) Silicon in agriculture, vol 8. Elsevier, Amsterdam, pp 197–207
- Savant NK, Snyder GH, Datnoff LE (1997) Silicon management and sustainable rice production. Adv Agron 58:151–199
- Arkadiusz A (2018) Effect of silicon fertilization on crop yield quantity and quality—a literature review in Europe. Plan Theory 7(3):54
- 5. Onodera I (1917) Chemical studies on rice blast (Dactylaria parasitance Cavara). J Agr Sci 180:606–617
- 6. Miyake Y, Takahashi E (1983) Effect of silicon on the growth of solution-cultured cucumber plant. Soil Sci Plant Nutr 29(1):71–83
- Datnoff LE, Brecht MO, Kucharek TA, Nagata RT (2002) The role of silicon in turf grass disease management. Abstract of second silicon in agriculture conference pp 105–110

- Aleksandrov VG, Blagodyr RN, Live IP (1967) Liberation of phosphoric acid from apatite by silicate bacteria. Microchem J 29:111–114
- 9. Chen J, Uroz S, Calvaruso C et al (2009) Mineral weathering by bacteria: ecology, actors and mechanisms. Trends Microbiol 17:378–387
- Henderson ME, Duff RB (1963) The release of metallic and silicate ions from minerals, rocks, and soils by fungal activity. J Soil Sci 14:237–245
- Bansal V, Rautaray D, Ahmad A et al (2004) Biosynthesis of zirconia nanoparticles using the fungus *Fusarium oxysporum*. J Mater Chem 14 (22):3303–3305
- 12. Maghsoudi K, Emam Y, Ashraf M (2015) Foliar application of silicon at different growth stages alters growth and yield of selected wheat cultivars. J Plant Nutr 39:1194–1203
- Kaya C, Tuna AL, Sonmez O, Ince F, Higgs D (2009) Mitigation effects of silicon on maize plants grown at high zinc. J. Plant Nutr., 32: 1788–1798. https://doi.org/10.1080/ 01904160903152624

Springer Protocols

Natarajan Amaresan Prittesh Patel Dhruti Amin *Editors*

Practical Handbook on Agricultural Microbiology



Springer Protocols Handbooks

For further volumes: http://www.springer.com/series/7657 Springer Protocols Handbooks collects a diverse range of step-by-step laboratory methods and protocols from across the life and biomedical sciences. Each protocol is provided in the Springer Protocol format: readily-reproducible in a step-by-step fashion. Each protocol opens with an introductory overview, a list of the materials and reagents needed to complete the experiment, and is followed by a detailed procedure supported by a helpful notes section offering tips and tricks of the trade as well as troubleshooting advice. With a focus on large comprehensive protocol collections and an international authorship, Springer Protocols Handbooks are a valuable addition to the laboratory.

More information about this series at http://www.springer.com/series/8623

Practical Handbook on Agricultural Microbiology

Edited by

Natarajan Amaresan, Prittesh Patel, and Dhruti Amin

Uka Tarsadia University, Surat, Gujarat, India

💥 Humana Press

Editors Natarajan Amaresan Uka Tarsadia University Surat, Gujarat, India

Dhruti Amin Uka Tarsadia University Surat, Gujarat, India Prittesh Patel Uka Tarsadia University Surat, Gujarat, India

ISSN 1949-2448 ISSN 1949-2456 (electronic) Springer Protocols Handbooks ISBN 978-1-0716-1723-6 ISBN 978-1-0716-1724-3 (eBook) https://doi.org/10.1007/978-1-0716-1724-3

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Humana imprint is published by the registered company Springer Science+Business Media, LLC part of Springer Nature.

The registered company address is: 1 New York Plaza, New York, NY 10004, U.S.A.

Preface

Agricultural Microbiology is a part of the microbiology branch dealing with beneficial or harmful microbes associated with either plants or soil. This manual focuses on beneficial microbes dealing with soil fertility, microbial degradation of organic matter, soil nutrient transformations, and biocontrol agents. Nowadays, techniques involved in the study of beneficial microbes in agricultural microbiology toward enhancing global agricultural productivity are in trend. This manual covers a wide range of basic and advanced techniques associated with research on the isolation of agriculturally important microbes, identification, biological nitrogen fixation, microbe-mediated plant nutrient use efficiency, and biological control of plant diseases and pests. Introduction to each protocol explains the role/importance of chemicals involved, uniqueness, and protocol application. A proper understanding of the protocol helps the researchers to manipulate them as per their need.

This book is composed of seven parts with 52 protocol chapters. Parts I and II represent the importance, isolation, and purification methods of agriculturally important microbes and include mineral-solubilizing microbes. Part III deals with phytohormones quantitative protocols directly or indirectly associated with microbes. Parts IV and V provide deep insights into protocols for screening agriculturally important enzymes and compounds related to biocontrol activity. Part VI represents assessment methods of soil microbial activity by soil respiration. The final Part VII deals with protocols for selecting microbial strains for inoculant production and quality control ultimately representing commercial biofertilizers production criteria. This book will help postgraduate students, research scholars, postdoctoral fellows, and teachers belonging to different disciplines of Plant Microbiology and Pathology. Moreover, this manual may also serve as a textbook for undergraduate courses like Techniques on Plant-Microbe Interaction/Biological Control of Plant Diseases/Nutrient Use Efficiency.

Surat, Gujarat, India Surat, Gujarat, India Surat, Gujarat, India Natarajan Amaresan Prittesh Patel Dhruti Amin

Contents

	face stributors	v xi
Par	AT I ISOLATION AND IDENTIFICATION OF AGRICULTURALLY IMPORTANT MICROBES	
1	Methods for Isolation and Identification of Rhizobia	3
2	Isolation of Frankia from Casuarina Root Nodule Narayanasamy Marappa, Dhanasekaran Dharumadurai, and Thajuddin Nooruddin	15
3	Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum Bhavana V. Mohite and Satish V. Patil	25
4	Isolation and Identification of Azospirillum	35
5	Isolation and Identification of <i>Gluconacetobacter diazotrophicus</i>	41
6	Isolation and Identification of Nitrogen Fixing Bacteria: Azoarcus Species	47
7	Isolation and Identification of <i>Derxia</i> Species from the Soil Sample Harshida A. Gamit and Natarajan Amaresan	57
8	Isolation and Characterization of Enterobacter, Klebsiella,and ClostridiumV. Mageshwaran and K. Pandiyan	63
9	Isolation and Characterization of Genus Desulfotomaculum Mobini Pimpalse, Harshida A. Gamit, and Natarajan Amaresan	71
10	Isolation and Identification of Associative Symbiotic N ₂ Fixing Microbes: Desulfovibrio	77
11	Cyanobacteria Dhruti Amin, Abhishek Sharma, and Sanket Ray	85
12	Pseudomonas Sanket Ray and Harsh Patel	93
13	Isolation and Identification of Entomopathogenic <i>Bacillus</i> Species Preeti Parmar, B. K. Rajkumar, and Naresh Butani	99
14	Methylobacterium	111

viii	Contents

15	Isolation and Identification of <i>Beijerinckia</i>	119
16	Isolation of Streptomyces from Soil Sample Vishnu Raja Vijayakumar and Dharumadurai Dhanasekaran	127
17	Isolation and Identification of <i>Trichoderma</i> Spp. from Different Agricultural Samples	131
18	Extraction, Isolation and Culturing of <i>Mycorrhizal</i> Spores from Rhizospheric Soil	145
19	Isolation and Identification of <i>Metarhizium</i>	151
20	Isolation and Identification of Bacteriophage for Biocontrol Mitesh Dwivedi	161
21	Isolation of Bacterivorous Protozoan, Acanthamoeba Spp., as New-Age Agro Bio-Input Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil	173
Par	TT II ISOLATION OF MINERAL SOLUBILIZING MICROBES	
22	Isolation and Screening of Zinc Solubilizing Microbes: As Essential Micronutrient Bio-Inputs for Crops Satish V. Patil, Hemant P. Borase, Jitendra D. Salunkhe, and Rahul K. Suryawanshi	181
23	Isolation and Screening of Mineral Phosphate Solubilizing Microorganisms	187
24	Isolation and Identification of Potassium-Solubilizing Microbes Prittesh Patel and Swati Patel	193
25	Isolation and Identification of Sulfur-Oxidizing Bacteria Vimalkumar Prajapati, Swati Patel, Radhika Patel, and Vaibhavkumar Mehta	197
26	Isolation of Ammonia Oxidizing Bacteria Naresh Butani, Shruti Satashia, Hemanshi Kanpariya, and Preeti Parmar	203
27	Isolation and Identification of Nitrite-Oxidizing Microbes Prittesh Patel, Ami Naik, and Abhishek Sharma	219
28	Isolation and Identification of Iron-Oxidizing Microbes Ami Naik and Pooja Patel	223
29	Isolation and Identification of Phytin Mineralizing Microbes Swati Patel and Prittesh Patel	231
30	Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil	237

31	I Isolation of Selenium Biotransforming Microbes as New Age Bioinputs Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil	
Par	T III ESTIMATION OF PHYTOHORMONES BY BENEFICIAL MICROBES	
32	Auxin Dixita Panchal, Jemisha Mistry, and Dhruti Amin	251
33	Abscisic Acid Natarajan Amaresan, A. Sankaranarayanan, and Dhruti Amin	257
34	Cytokinins Jawahar Ganapathy, Jemisha Mistry, and Dhruti Amin	263
35	Ethylene	269
36	Gibberellin Dhruti Amin, Sanket Ray, and Abhishek Sharma	273
37	Brassinosteroids	277
38	Strigolactones: Extraction and Characterization	283
39	Estimation of Jasmonic Acid Using Non-pathogenic Microbes Jasmonic Acid Krutika S. Abhyankar and Monisha Kottayi	289
Par	TT IV SCREENING OF AGRICULTURALLY IMPORTANT ENZYMES	
40	Chitinase Purvesh B. Bharvad and Harsha J. Algotar	301
41	Glucanase Purvesh B. Bharvad and Harsha J. Algotar	309
42	Identification of Cellulase Enzyme Involved in Biocontrol Activity	317
43	Identification of Protease Enzymes Involved in Biocontrol Activity Vimalkumar Prajapati, Swati Patel, Sanket Ray, and Kamlesh C. Patel	323
44	Isolation and Screening of Naringinase Producing Microbes: As Industrial Inputs for Agro Waste Base Enzyme Industry Satish V. Patil, Jitendra D. Salunkhe, and Vishal Marathe	331
45	Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil	337

Par	тV	Identification of Compounds Involved in Biocontrol Activity	
46		rogen Cyanide ti Bhatt and Dhruti Amin	345
47		ophores sa Patel	351
48	for D	tion and Screening of ACC Deaminase-Producing Microbes Prought Stress Management in Crops	361
49		olysaccharides a Chandwani and Natarajan Amaresan	369
Par	T VI	Assessment of Soil Microbial Activity by Soil Respiration	
50		matic Analyses in Soils ar Bilen and Veysel Turan	377
Par	т VII	Selection of Microbial Strains for Inoculant Production and Quality Control	
51		tion of <i>Rhizobium</i> Strain for Inoculum Production	389
52		her Culture, Broth, and Peat Test uti Amin, Sanket Ray, and Vrushali Wagh	395
Ind	ex		405

Contributors

- KRUTIKA S. ABHYANKAR School of Science, Navrachana University, Vadodara, Gujarat, India
- HARSHA J. ALGOTAR . D. L. Patel Science College, Himatnagar, Gujarat, India
- NATARAJAN AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- DHRUTI AMIN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- HIMANSHU BARIYA Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- PURVESH B. BHARVAD . D. L. Patel Science College, Himatnagar, Gujarat, India
- KHYATI BHATT Post Graduate Department of Biosciences, Sardar Patel University, Gujarat, India
- SERDAR BILEN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Atatürk University, Erzurum, Turkey
- HEMANT P. BORASE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- NARESH BUTANI Department of Microbiology, Faculty of Science, Sarvajanik University, Surat, India
- SAPNA CHANDWANI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- SHREYA DESAI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- DHANASEKARAN DHARUMADURAI Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- MITESH DWIVEDI C. G. Bhakta Institute of Biotechnology, Faculty of Science, Uka Tarsadia University, Surat, Gujarat, India
- HARSHIDA A. GAMIT C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- JAWAHAR GANAPATHY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- TEJAS GOHIL Sabarmati Ashram Gaushala, Kheda, Gujarat, India
- HEMANSHI KANPARIYA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- MONISHA KOTTAYI School of Science, Navrachana University, Vadodara, Gujarat, India
- V. MAGESHWARAN ICAR-National Bureau of Agriculturally Important Microorganisms, Mau, Uttar Pradesh, India
- MAHADEVASWAMY University of Agricultural Sciences, Raichur, Karnataka, India
- NARAYANASAMY MARAPPA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- KIRAN MARATHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- VISHAL MARATHE N.E.S. Science College, Nanded, Maharashtra, India

- VAIBHAVKUMAR MEHTA Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- HARSH MISTRY Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- JEMISHA MISTRY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- BHAVANA V. MOHITE Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India
- AMI NAIK C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- VRUTUJA NAIK National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- PRADNYA B. NIKAM School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- THAJUDDIN NOORUDDIN Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- DIXITA PANCHAL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- KETANKUMAR J. PANCHAL Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, India
- K. PANDIYAN . ICAR-Central Institute for Research on Cotton Technology, Mumbai, India
- PREETI PARMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- HARSH PATEL P. G. Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, Gujarat, India
- KAMLESH C. PATEL PG Department of Biosciences, Sardar Patel University, Anand, Gujarat, India
- NAFISA PATEL Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- POOJA PATEL . Government Medical College, Surat, Gujarat, India
- PRITTESH PATEL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- RADHIKA PATEL PG Department of Biosciences, Sardar Patel University, Gujarat, India
- SWATI PATEL Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- TARUN KUMAR PATEL Department of Biotechnology, Sant Guru Ghasidas Government P.G. College, Dhamtari, Chattisgarh, India; Department of Biotechnology, Guru Ghasidas Vishwavidyalaya (a Central University), Bilaspur, Chattisgarh, India
- CHANDRASHEKHAR D. PATIL Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- SATISH V. PATIL School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India
- VIKAS S. PATIL University Institute of Chemical Technology, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- MOHINI PIMPALSE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, India
- VIMALKUMAR PRAJAPATI Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India

- PRAVEEN RAHI National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- VIJAYAKUMAR VISHNU RAJA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- B. K. RAJKUMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- SANKET RAY Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- NARENDRA SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- JITENDRA D. SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- A. SANKARANARAYANAN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- SHRUTI SATASHIA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- ABHISHEK SHARMA C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- K. SOWMIYA University of Agricultural Sciences, Raichur, Karnataka, India
- RAHUL K. SURYAWANSHI Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- RASHMI THAKOR Department of Life Sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- VEYSEL TURAN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Bingöl University, Bingöl, Turkey
- VRUSHALI WAGH Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India



Isolation of Bacterivorous Protozoan, *Acanthamoeba* Spp., as New-Age Agro Bio-Input

Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil

Abstract

Protozoan is widely distributed in the environment. *Acanthamoeba* genus are free-living protozoan of nonpathogenic and pathogenic nature. Its intrinsic ability to feed on soil bacteria contributes greatly to the soil nutrient turnover and thereby plant growth. This protocol provides a firsthand guide to isolate *Acanthamoeba* from soil samples and their preliminary identification through staining procedure. The isolates could be further tested for plant beneficial roles in laboratory settings.

Key words Protozoan, Acanthamoeba, Agriculture, Cyst, Soil fertility

1 Introduction

Protozoa are single-celled eukaryotic microorganisms. They are heterotrophic and mostly predate on the soil bacteria. They are widely present in water and soil habitat. Through their feeding behavior, they play an important role in recycling of soil organic nutrients. Members of *Acanthamoeba* genus are free-living Protozoa and most predominant bacterivores species in the environment. The classification scheme of *acanthamoeba* genus is as follows:

Kingdom: Protista. Subkingdom: Protozoa. Phylum: Sarcomastigophora. Subphylum: Sarcodina. Superclass: Rhizopoda. Class: Lobosea. Subclass: Gymnamoebia.

Chandrashekhar D. Patil and Satish V. Patil contributed equally to this work.

Natarajan Amaresan et al. (eds.), Practical Handbook on Agricultural Microbiology,

Springer Protocols Handbooks, https://doi.org/10.1007/978-1-0716-1724-3_21,

[©] The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

Order: *Amoebida*. Family: *Acanthamoebidae*. Genus: *Acanthamoeba*.

Free-living Amoebae of Acanthamoeba genus include nonpathogenic and pathogenic strains that are currently classified in 18 different genotypes, T1–T18 [1]. The life cycle of Acanthamoeba consists of two stages, metabolically active, trophozoite, and metabolically inactive cyst. Both trophozoite and cysts are characterized by a nucleus surrounded by a dense central nucleolus [2]. The interchange between two life stages depends on the external stimuli such as nutrient depletion or availability. A gram of soil generally contains 10^3 – 10^7 amoeba with varying size < 10μ m. The differences between cyst size and shape were the early markers for the differentiation between Acanthamoeba spp.

However, distinction between closely similar species was a major challenge due to the irregularity and variation in shapes. Latest molecular tools are highly recommended to profile the soil isolates and characterize their genotypes [3]. Due to the most abundant nature of *Acanthamoeba* in the environment, it is a suitable model organism for soil protozoan function studies [4].

Top organic soil layer (10–15 cm depth) harbors the high number of protozoa and microbial communities. *Acanthamoeba* voraciously feeds on soil bacteria at a rate from 0.2 to 1465 bacterial h^{-1} [5, 6]. Therefore, presence of protozoan, microorganisms, and soil nutrition quality are closely associated with conditions for the successful nutrient turnover via formation of "microbial loop in soil" [7]. Plant root exudates contain the nutrient that triggers the bacterial growth through assimilation of carbon and nitrogen in the rhizosphere. The abundance of bacteria in rhizosphere favors the progression of bacterivorous protozoan and ultimately regulation of bacterial population in soil. Protozoa preying on bacteria thus make nitrogen available for plant growth [4].

Considering the potential of bacterivorous protozoan in organic farming and as a next generation of agriculture bio-inputs [8], this chapter outlines the practical steps to isolate the *acantha-moeba* spp. as a model organism and their preliminary identification.

2 Material and Methods

2.1 Requirements

- 1. Soil sample.
- Minimal nutrient medium (peptone: 1.25 g/l, yeast extract: 1.25 g/l, dextrose: 3.0 g/l, agar: 20.0 g/l).
- 3. Petri plates.
- 4. E. coli culture.

- 5. Autoclave.
- 6. Heating chamber.
- 7. Microscope.
- 8. Staining reagents.
- **2.2** Culture Medium **Preparation** Acanthamoeba spp. can be easily cultivated on non-nutrient agar or agar media containing minimal concentrations of nutrients. Sterilize the medium by autoclaving at 15 lb. pressure for 20 min, allow the medium to cool at 45 °C, then pour the medium into petri plates, and allow them to solidify for 1 h at room temperature or 30 °C for 2 h. After solidification, aseptically overlay 1 ml of 24 h old culture of *E. coli* heated at 60 °C for 10 min, then incubate the plates at room temperature for 1 h.
- **2.3 Collection and** In sterile plastic bags, collect 1 g of rhizospheric soil from the irrigated soil/farms and crop plants as per need. Close the bag and label the sample appropriately (e.g., location, crop, date, etc.). The sample should be store at 4 °C until further experimentation (*see* Note 1).
- **2.4 Primary** The cultivated bacterivorous protozoan can be identified by observing loopful of culture with loopful of water under $40 \times$, note down the structure (Fig. 1). The *Acanthamoeba* cyst and trophozoite stage can be identified by using simple staining methods like methylene blue, cotton blue, and Giemsa staining.
- 2.5 Detail Procedure
 2.5.1 Part I
 1. Dilute the collected rhizospheric soil with sterile saline by routine serial dilution techniques up to 10⁹ dilution and mix well.
 - 2. Aseptically remove 0.5 ml from diluted sample up to 10^{6} and plate it on sterile culture medium, and spread the sample by rotating plates clockwise and anticlockwise to spread sample uniformly.
 - 3. Incubate plates at 30 °C for 24–72 h.
 - 4. At every 24 h, take loopful of culture and dilute with drop of sterile water.
 - 5. Put this drop carefully at the center of cavity slide with wax at four corners, and invert the slide to make hanging drop preparation and observe under microscope at $40 \times$.
 - 6. Note down the shape and structure of cultivated protozoan (*see* **Note 2**).
 - Further observation for identification can also be done by using methylene blue, cotton blue, iodine, and Giemsa staining (*see* Notes 3–7).

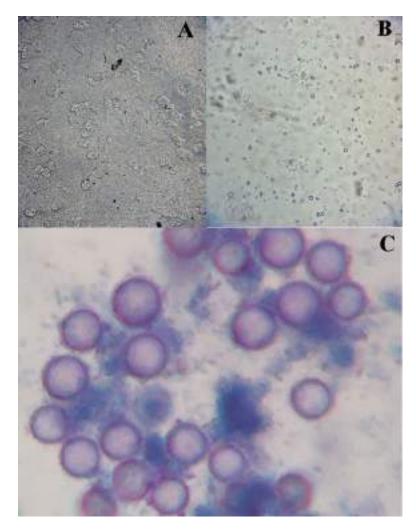


Fig. 1 Microscopic observation of different *Acanthamoeba* life forms. (a) *Acanthamoeba* cyst (10×). (b) Free-living trophozoites of *Acanthamoeba*. (c) Stained *Acanthamoeba* cyst (methylene blue staining, $40\times$)

- 2.5.2 Part II: Staining 1. Take a loopful culture of protozoan and spread on a glass slide and gently heat on the flame.
 - 2. Flood the fixed smears of protozoan cyst with freshly filtered carbol fuchsin solution.
 - 3. Then wash the smear in tap water and flood the smear with malachite green as counterstain for 10 s.
 - 4. Then rinse the smear again in tap water and air-dry.

2.5.3 Part III	The pure culture of <i>Acanthamoeba</i> can be done by following steps:
Axenic Culture of Acanthamoeba	1. Harvest the cysts from the peptone-yeast extract-dextrose (PYD) plate cultures by adding 5 ml sterile saline in 2-week-old <i>Acanthamoeba</i> seeded plates (<i>see</i> Note 8).
	2. Collect the suspension and then centrifuge at 7000 rpm (centrifugal g force 16052). For 10 min, then collect the pellet.
	3. Incubate the pellet with 3% HCl overnight for elimination of the bacteria.
	4. Then wash the cysts 2–3 with physiological saline (7000 rpm for 10 min) to remove remaining HCl.
	5. Then transfer it to liquid cultures sterile PYD medium.
	6. The culture should be maintained axenic by transferring every second week to fresh sterile PYD medium.

3 Notes

- 1. Since *Acanthamoeba* is both pathogenic and nonpathogenic in nature, appropriate safety measure should be taken to handle the samples. Sample collection from natural sites with minimum human exposure should be preferred for the isolation of nonpathogenic *Acanthamoeba* spp.
- 2. Cysts can be counted using a hemocytometer.
- 3. Basic fuchsin stain in aqueous solution with phenol and ethanol will stain *Acanthamoeba* cysts in red/magenta color against a greenish-blue background.
- 4. Eosin, iodine, and methylene blue stains are useful for wet mount staining. Eosin stain can be prepared by adding 1 g eosin in 100 ml distilled water, and 0.5 ml acetic acid.
- 5. Prepare methylene blue stain by dissolving 1 g of methylene blue powder in 100 ml physiological saline.
- 6. For wet mount staining, spread two drops $(25-30 \ \mu l)$ of *Acanthamoeba* suspension on a glass slide; add a drop of the stain, cover with glass coverslip, and observe under microscope at $40 \times$ power.
- 7. The all above staining methods allow to observe cysts of *Acanthamoeba* known to form double-walled cysts, the outer wall ectocyst which is differentiated from the variably stained surrounding background. The inner wall appears as stellated, polygonal, round, or oval forms.
- Sodium dodecyl sulfate (SDS, 0.5% final concentration) can be used to confirm the transformation of trophozoites into cysts. Trophozoites are SDS-sensitive and any remaining are lysed immediately upon addition of SDS, while cysts remain intact (Dudley, 2005).

Acknowledgments

We acknowledge Prof. Michael Bonkowski, University of Cologne, Germany for pioneering the work in ecosystem engineers in shaping rhizosphere microbial diversity and functioning and their feedback on plant productivity. We are also thankful to our research collaborator Prof. Marilise B Rott, Universidade Federal do Rio Grande do Sul, Brazil for inspiring us on protozoan study.

References

- Khan NA (2006) Acanthamoeba: biology and increasing importance in human health. FEMS Microbiol Rev 30(4):564–595
- Visvesvara GS (1991) Classification of Acanthamoeba. Clin Infect Dis 13(Supplement_5): S369–S372. https://doi.org/10.1093/clind/ 13.supplement_5.s369
- Reyes-Batlle M, Todd CD, Martín-Navarro CM, López-Arencibia A, Cabello-Vilchez AM, González AC, Córdoba-Lanús E, Lindo JF, Valladares B, Piñero JE, Lorenzo-Morales J (2014) Isolation and characterization of *Acanthamoeba* strains from soil samples in Gran Canaria, Canary Islands, Spain. Parasitol Res 113(4):1383–1388
- Bonkowski M, Brandt F (2002) Do soil protozoa enhance plant growth by hormonal effects? Soil Biol Biochem 34(11):1709–1715

- Heaton K, Drinkall J, Minett A, Hunt AP, Parry JD (2001) Amoeboid grazing on surfaceassociated prey. In: Gilbert P, Allison D, Brading M, Verran J, Walker J (eds) Biofilm community interaction – chance or necessity? Bioline, Cardiff University, Cardiff, pp 293–301
- Huws SA, McBain AJ, Gilbert P (2005) Protozoan grazing and its impact upon population dynamics in biofilm communities. J Appl Microbiol 98:238–244
- 7. Clarholm M (1985) Interactions of bacteria, protozoa and plants leading to mineralization of soil nitrogen. Soil Biol Biochem 17 (2):181–187
- Mohite BV et al (2019) New age agricultural bioinputs. In: Singh D, Gupta V, Prabha R (eds) Microbial interventions in agriculture and environment. Springer, Singapore. https://doi. org/10.1007/978-981-13-8391-5_14

Springer Protocols

Natarajan Amaresan Prittesh Patel Dhruti Amin *Editors*

Practical Handbook on Agricultural Microbiology



Springer Protocols Handbooks

For further volumes: http://www.springer.com/series/7657 Springer Protocols Handbooks collects a diverse range of step-by-step laboratory methods and protocols from across the life and biomedical sciences. Each protocol is provided in the Springer Protocol format: readily-reproducible in a step-by-step fashion. Each protocol opens with an introductory overview, a list of the materials and reagents needed to complete the experiment, and is followed by a detailed procedure supported by a helpful notes section offering tips and tricks of the trade as well as troubleshooting advice. With a focus on large comprehensive protocol collections and an international authorship, Springer Protocols Handbooks are a valuable addition to the laboratory.

More information about this series at http://www.springer.com/series/8623

Practical Handbook on Agricultural Microbiology

Edited by

Natarajan Amaresan, Prittesh Patel, and Dhruti Amin

Uka Tarsadia University, Surat, Gujarat, India

💥 Humana Press

Editors Natarajan Amaresan Uka Tarsadia University Surat, Gujarat, India

Dhruti Amin Uka Tarsadia University Surat, Gujarat, India Prittesh Patel Uka Tarsadia University Surat, Gujarat, India

ISSN 1949-2448 ISSN 1949-2456 (electronic) Springer Protocols Handbooks ISBN 978-1-0716-1723-6 ISBN 978-1-0716-1724-3 (eBook) https://doi.org/10.1007/978-1-0716-1724-3

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Humana imprint is published by the registered company Springer Science+Business Media, LLC part of Springer Nature.

The registered company address is: 1 New York Plaza, New York, NY 10004, U.S.A.

Preface

Agricultural Microbiology is a part of the microbiology branch dealing with beneficial or harmful microbes associated with either plants or soil. This manual focuses on beneficial microbes dealing with soil fertility, microbial degradation of organic matter, soil nutrient transformations, and biocontrol agents. Nowadays, techniques involved in the study of beneficial microbes in agricultural microbiology toward enhancing global agricultural productivity are in trend. This manual covers a wide range of basic and advanced techniques associated with research on the isolation of agriculturally important microbes, identification, biological nitrogen fixation, microbe-mediated plant nutrient use efficiency, and biological control of plant diseases and pests. Introduction to each protocol explains the role/importance of chemicals involved, uniqueness, and protocol application. A proper understanding of the protocol helps the researchers to manipulate them as per their need.

This book is composed of seven parts with 52 protocol chapters. Parts I and II represent the importance, isolation, and purification methods of agriculturally important microbes and include mineral-solubilizing microbes. Part III deals with phytohormones quantitative protocols directly or indirectly associated with microbes. Parts IV and V provide deep insights into protocols for screening agriculturally important enzymes and compounds related to biocontrol activity. Part VI represents assessment methods of soil microbial activity by soil respiration. The final Part VII deals with protocols for selecting microbial strains for inoculant production and quality control ultimately representing commercial biofertilizers production criteria. This book will help postgraduate students, research scholars, postdoctoral fellows, and teachers belonging to different disciplines of Plant Microbiology and Pathology. Moreover, this manual may also serve as a textbook for undergraduate courses like Techniques on Plant-Microbe Interaction/Biological Control of Plant Diseases/Nutrient Use Efficiency.

Surat, Gujarat, India Surat, Gujarat, India Surat, Gujarat, India Natarajan Amaresan Prittesh Patel Dhruti Amin

Contents

	face stributors	v xi
Par	AT I ISOLATION AND IDENTIFICATION OF AGRICULTURALLY IMPORTANT MICROBES	
1	Methods for Isolation and Identification of Rhizobia	3
2	Isolation of Frankia from Casuarina Root Nodule Narayanasamy Marappa, Dhanasekaran Dharumadurai, and Thajuddin Nooruddin	15
3	Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum Bhavana V. Mohite and Satish V. Patil	25
4	Isolation and Identification of Azospirillum	35
5	Isolation and Identification of <i>Gluconacetobacter diazotrophicus</i>	41
6	Isolation and Identification of Nitrogen Fixing Bacteria: Azoarcus Species	47
7	Isolation and Identification of <i>Derxia</i> Species from the Soil Sample Harshida A. Gamit and Natarajan Amaresan	57
8	Isolation and Characterization of Enterobacter, Klebsiella,and ClostridiumV. Mageshwaran and K. Pandiyan	63
9	Isolation and Characterization of Genus Desulfotomaculum Mobini Pimpalse, Harshida A. Gamit, and Natarajan Amaresan	71
10	Isolation and Identification of Associative Symbiotic N ₂ Fixing Microbes: Desulfovibrio	77
11	Cyanobacteria Dhruti Amin, Abhishek Sharma, and Sanket Ray	85
12	Pseudomonas Sanket Ray and Harsh Patel	93
13	Isolation and Identification of Entomopathogenic <i>Bacillus</i> Species Preeti Parmar, B. K. Rajkumar, and Naresh Butani	99
14	Methylobacterium	111

viii	Contents

15	Isolation and Identification of <i>Beijerinckia</i>	119
16	Isolation of Streptomyces from Soil Sample Vishnu Raja Vijayakumar and Dharumadurai Dhanasekaran	127
17	Isolation and Identification of <i>Trichoderma</i> Spp. from Different Agricultural Samples	131
18	Extraction, Isolation and Culturing of <i>Mycorrhizal</i> Spores from Rhizospheric Soil	145
19	Isolation and Identification of <i>Metarhizium</i>	151
20	Isolation and Identification of Bacteriophage for Biocontrol Mitesh Dwivedi	161
21	Isolation of Bacterivorous Protozoan, Acanthamoeba Spp., as New-Age Agro Bio-Input Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil	173
Par	TT II ISOLATION OF MINERAL SOLUBILIZING MICROBES	
22	Isolation and Screening of Zinc Solubilizing Microbes: As Essential Micronutrient Bio-Inputs for Crops Satish V. Patil, Hemant P. Borase, Jitendra D. Salunkhe, and Rahul K. Suryawanshi	181
23	Isolation and Screening of Mineral Phosphate Solubilizing Microorganisms	187
24	Isolation and Identification of Potassium-Solubilizing Microbes Prittesh Patel and Swati Patel	193
25	Isolation and Identification of Sulfur-Oxidizing Bacteria Vimalkumar Prajapati, Swati Patel, Radhika Patel, and Vaibhavkumar Mehta	197
26	Isolation of Ammonia Oxidizing Bacteria Naresh Butani, Shruti Satashia, Hemanshi Kanpariya, and Preeti Parmar	203
27	Isolation and Identification of Nitrite-Oxidizing Microbes Prittesh Patel, Ami Naik, and Abhishek Sharma	219
28	Isolation and Identification of Iron-Oxidizing Microbes Ami Naik and Pooja Patel	223
29	Isolation and Identification of Phytin Mineralizing Microbes Swati Patel and Prittesh Patel	231
30	Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil	237

31	I Isolation of Selenium Biotransforming Microbes as New Age Bioinputs Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil	
Par	T III ESTIMATION OF PHYTOHORMONES BY BENEFICIAL MICROBES	
32	Auxin Dixita Panchal, Jemisha Mistry, and Dhruti Amin	251
33	Abscisic Acid Natarajan Amaresan, A. Sankaranarayanan, and Dhruti Amin	257
34	Cytokinins Jawahar Ganapathy, Jemisha Mistry, and Dhruti Amin	263
35	Ethylene Ketankumar J. Panchal and Dhruti Amin	269
36	Gibberellin Dhruti Amin, Sanket Ray, and Abhishek Sharma	273
37	Brassinosteroids	277
38	Strigolactones: Extraction and Characterization	283
39	Estimation of Jasmonic Acid Using Non-pathogenic Microbes Jasmonic Acid Krutika S. Abhyankar and Monisha Kottayi	289
Par	TT IV SCREENING OF AGRICULTURALLY IMPORTANT ENZYMES	
40	Chitinase Purvesh B. Bharvad and Harsha J. Algotar	301
41	Glucanase Purvesh B. Bharvad and Harsha J. Algotar	309
42	Identification of Cellulase Enzyme Involved in Biocontrol Activity	317
43	Identification of Protease Enzymes Involved in Biocontrol Activity Vimalkumar Prajapati, Swati Patel, Sanket Ray, and Kamlesh C. Patel	323
44	Isolation and Screening of Naringinase Producing Microbes: As Industrial Inputs for Agro Waste Base Enzyme Industry Satish V. Patil, Jitendra D. Salunkhe, and Vishal Marathe	331
45	Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil	337

Par	тV	Identification of Compounds Involved in Biocontrol Activity	
46		rogen Cyanide ti Bhatt and Dhruti Amin	345
47		ophores sa Patel	351
48	for D	tion and Screening of ACC Deaminase-Producing Microbes Prought Stress Management in Crops	361
49		olysaccharides a Chandwani and Natarajan Amaresan	369
Par	T VI	Assessment of Soil Microbial Activity by Soil Respiration	
50		matic Analyses in Soils ar Bilen and Veysel Turan	377
Par	т VII	Selection of Microbial Strains for Inoculant Production and Quality Control	
51		tion of <i>Rhizobium</i> Strain for Inoculum Production	389
52		her Culture, Broth, and Peat Test uti Amin, Sanket Ray, and Vrushali Wagh	395
Ind	ex		405

Contributors

- KRUTIKA S. ABHYANKAR School of Science, Navrachana University, Vadodara, Gujarat, India
- HARSHA J. ALGOTAR . D. L. Patel Science College, Himatnagar, Gujarat, India
- NATARAJAN AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- DHRUTI AMIN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- HIMANSHU BARIYA Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- PURVESH B. BHARVAD . D. L. Patel Science College, Himatnagar, Gujarat, India
- KHYATI BHATT Post Graduate Department of Biosciences, Sardar Patel University, Gujarat, India
- SERDAR BILEN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Atatürk University, Erzurum, Turkey
- HEMANT P. BORASE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- NARESH BUTANI Department of Microbiology, Faculty of Science, Sarvajanik University, Surat, India
- SAPNA CHANDWANI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- SHREYA DESAI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- DHANASEKARAN DHARUMADURAI Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- MITESH DWIVEDI C. G. Bhakta Institute of Biotechnology, Faculty of Science, Uka Tarsadia University, Surat, Gujarat, India
- HARSHIDA A. GAMIT C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- JAWAHAR GANAPATHY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- TEJAS GOHIL Sabarmati Ashram Gaushala, Kheda, Gujarat, India
- HEMANSHI KANPARIYA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- MONISHA KOTTAYI School of Science, Navrachana University, Vadodara, Gujarat, India
- V. MAGESHWARAN ICAR-National Bureau of Agriculturally Important Microorganisms, Mau, Uttar Pradesh, India
- MAHADEVASWAMY University of Agricultural Sciences, Raichur, Karnataka, India
- NARAYANASAMY MARAPPA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- KIRAN MARATHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- VISHAL MARATHE N.E.S. Science College, Nanded, Maharashtra, India

- VAIBHAVKUMAR MEHTA Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- HARSH MISTRY Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- JEMISHA MISTRY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- BHAVANA V. MOHITE Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India
- AMI NAIK C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- VRUTUJA NAIK National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- PRADNYA B. NIKAM School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- THAJUDDIN NOORUDDIN Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- DIXITA PANCHAL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- KETANKUMAR J. PANCHAL Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, India
- K. PANDIYAN . ICAR-Central Institute for Research on Cotton Technology, Mumbai, India
- PREETI PARMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- HARSH PATEL P. G. Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, Gujarat, India
- KAMLESH C. PATEL PG Department of Biosciences, Sardar Patel University, Anand, Gujarat, India
- NAFISA PATEL Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- POOJA PATEL . Government Medical College, Surat, Gujarat, India
- PRITTESH PATEL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- RADHIKA PATEL PG Department of Biosciences, Sardar Patel University, Gujarat, India
- SWATI PATEL Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- TARUN KUMAR PATEL Department of Biotechnology, Sant Guru Ghasidas Government P.G. College, Dhamtari, Chattisgarh, India; Department of Biotechnology, Guru Ghasidas Vishwavidyalaya (a Central University), Bilaspur, Chattisgarh, India
- CHANDRASHEKHAR D. PATIL Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- SATISH V. PATIL School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India
- VIKAS S. PATIL University Institute of Chemical Technology, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- MOHINI PIMPALSE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, India
- VIMALKUMAR PRAJAPATI Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India

- PRAVEEN RAHI National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- VIJAYAKUMAR VISHNU RAJA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- B. K. RAJKUMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- SANKET RAY Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- NARENDRA SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- JITENDRA D. SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- A. SANKARANARAYANAN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- SHRUTI SATASHIA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- ABHISHEK SHARMA C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- K. SOWMIYA University of Agricultural Sciences, Raichur, Karnataka, India
- RAHUL K. SURYAWANSHI Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- RASHMI THAKOR Department of Life Sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- VEYSEL TURAN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Bingöl University, Bingöl, Turkey
- VRUSHALI WAGH Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India



Extraction, Isolation and Culturing of *Mycorrhizal* Spores from Rhizospheric Soil

Satish V. Patil, Bhavana V. Mohite, and Chandrashekhar D. Patil

Abstract

Agriculture is experiencing the time of innovation with soil and its biological interactions. *Mycorrhiza* is a fungus associated with plant roots. *Mycorrhiza* is an ancient living ubiquitous fungus in soil. A decade ago, JL Harley pronounced a sentence "Plants don't have roots, they have mycorrhiza" in order to attract the attention of scientists towards plant-associated fungi in mutualistic association. *Mycorrhizae* increase the absorption of various nutrients, particularly phosphorus along with K, Si, Se, Zn, and Fe, and thus improve the crop productivity. The present chapter is focused on extraction, isolation, and culturing of *Mycorrhizal* fungi.

Key words Micronutrient, Mycorrhiza, Spores, Solubilization

1 Introduction

It is well known that the fungus plays various vital roles for mineral metabolism in soil and supplement to various plant. Bacteria and fungi are major contributors for P supplier to plant, besides these they promote nutrient availability, P, K, Si, Se, Zn, and Fe mobilizations through production of organic acids, metal chelators, protein, amino acids, and enzyme production (Fig. 1). Mycorrhizae are the nonpathogenic fungi which are associated with plant rhizosphere by mutualistic associations. They cause mild parasitism by invading roots for specific nutrients and provide various nutrients to plants. It is assumed that 90% of plants depend on Mycorrhizal supply of mineral nutrients especially phosphorus and iron (Fig. 2c). During some seasonal changes, they also provide nutrients like carbohydrate, sugar, and nitrogenous compounds. There are major two types as per the association, i.e., ecto Mycorrhiza and endo Mycorrhizae. Ecto Mycorrhiza means the fungus which is associated with external root surfaces and endo Mycorrhiza is associated with internal root cells of plant growing internally in plant

Natarajan Amaresan et al. (eds.), Practical Handbook on Agricultural Microbiology,

Springer Protocols Handbooks, https://doi.org/10.1007/978-1-0716-1724-3_18,

[©] The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

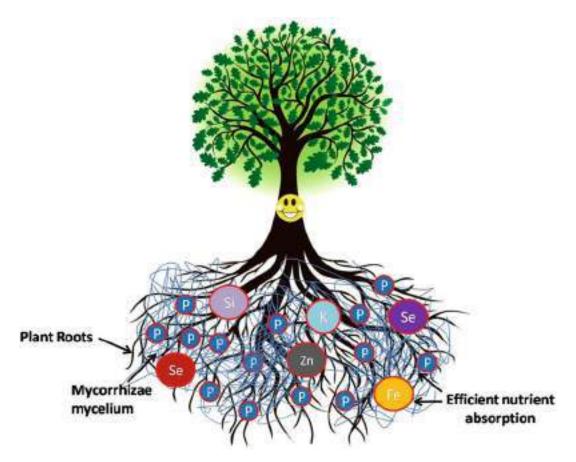


Fig. 1 Mycorrhizal plant roots mutualistic interaction

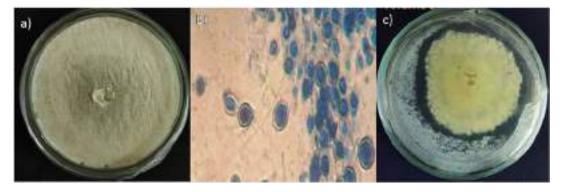


Fig. 2 (a) Growth of representative *Mycorrhizal* culture on agar plate (*Piriformospora indica*) (b) Microscopic observation of pear-shape *Mycorrhizal* spores (c) Zinc solubilization by *Piriformospora indica* (Dr. Satish V Patil Laboratory)

organs. Many times they are plant root specific, e.g., truffles *Terfe*zia boudieri with oaks, Larix with larch plants, some Rhizoctonia with orchids. There is vast diversity of *Mycorrhizae* as per crop, plant, and area but generally the *Mycorrhizae* are reported by detecting or extracting *Mycorrhizal* spores from soil and root samples. Although, there are some *Mycorrhizae* which do not produce always spore, such cultures may be identified by growing with plant tissue culture technique and other advance methods. Even if there are various techniques such as sucrose centrifugations [1] and adhesion floatation technique (AF) [2], capillarity adhesion method for spore extraction from soil and root are reported but wet sieving and decanting is the most practicable method for *Mycorrhizal* spore detection and isolation [3].

2 Principle of Assay

Extraction assay is based on principle that the *Mycorrhiza* are hydrophobic and light weight, and they float on water. These spores are separated by micro mesh sieves (250, 150, 53, and 45μ m) and collected from the residue present on the sieves in petri plate and used for microscopic observation and cultivation (*see* Note 4).

3 Materials

- Rhizospheric soil/roots, rootlets, etc.
- Sterile containers for soil sample collection.
- Sterile tubes for serial dilution of soil samples.
- Staining dye 0.05% trypan blue in lactophenol.
- Fragaria vesca L., Festuca ovina L., and Plectranthus plantlets for cultivation of VAM.

4 Methods

4.1 Collection of Soil Samples	1. Collect the 100 g of Rhizospheric or root associated soil sam- ples/roots of specific plants at different locations.
and Extraction	2. Add 100 g soil sample in 1000 ml sterile water in glass con- tainer, mix vigorously for 2 min, and keep at 10 °C in refriger- ator or in BOD incubator for 8–12 h.
	3. After incubation, the supernatant slowly passes through the series of sieves 250, 150, 53, and 45/38μm (<i>see</i> Note 4).
	4. Collect the residue present on each sieve separately in petri plates by washing sieves with sterile distilled water.
	5. Observe the collected sieve wash under microscope.
	6. For the plant root, dry the root material at 60 °C oven for 10–12 h, and grind the roots in grinder and soak the root

powder in sterile distilled water for 5 h, mix vigorously, and remove floating cellulosic material physically.

- 7. Allow the remaining water to pass through the sieves and follow above **steps 4** and **5**.
- 8. Collect the residue present on each sieve separately by washing with sterile water in petri plates.
- 9. Observe the spore morphology under microscope and identify the type of *Mycorrhizal* spore comparing with standard available cultures.
- 10. Using the dissecting microscope and micropipette, separate the *Mycorrhizal* spores and inoculate in sterile soil with plantlets of *Arabidopsis thaliana*, white clover *Trifolium repens*, Coleus (*Plectranthus scutellarioides*), in small pots, and allow to grow for 30 days (*see* **Note 3**).
- 11. Observe the root for *Mycorrhizal* colonization on root cells by root staining methods.
- 12. The spores collected/identified are inoculated with suitable coculturing plant and after 30 days observed *Mycorrhizal* association/spore, etc., and calculate its efficiency by following methods (*see* Note 5).
- 13. The culturable *Mycorrhiza*, i.e., *Piriformospora indica*, truffles *Terfezia boudieri* may be possible to culture on artificial medium in laboratory (*see* **Note 2**).
- 1. Cut the roots in very small pieces, wash thoroughly under tap water and boil the pieces at 95 °C in 10% KOH for different time 10, 15, 20, and 25–30 min.
- 2. Then cool the material. Separate it, wash again with tap water then again treat with 1 N HCL for 5 min.
- 3. Stain the root pieces with 0.05% trypan blue lactophenol reagent. Mount the material on glass slide with fixing reagent, cover the material with coverslip, seal it by applying wax on corner, and observe under $40 \times$ (Fig. 2b) (*see* Note 1).
- 4. Measure the segment with spores and calculate the percentage of *Mycorrhizal* association by the following formula [4].
 % *Mycorrhizal* association = Number of *Mycorrhiza* associated segments/Total Number of segments analyzed × 100.

5 Notes

- 1. The fixative solution for staining the root is acetic acid; glycerol (1:1 V/V).
- 2. There are few *Mycorrhizae* spp. that are culturable on artificial medium, e.g., *Truffles. p indicus* (Fig. 2a). Other *Mycorrhizae*

4.2 Staining and Determination of Percent of Mycorrhizal Association fungi are only maintained with seedlings of various plants, e.g., Trifolium repens, Coleus (Plectranthus scutellarioides), Arabidopsis thaliana.

- 3. The plant specificity was reported for various *Mycorrhizae* sp., hence it should maintain on specific plantlets as per *Mycorrhizae sp*.
- 4. Glomus species spores generally retain in the 38/45μm sieve. It also catches the majority of spores including large *Gigaspora gigantea* and visible as bright greenish dots under microscope.
- For better identification of spores, use spore plate photograph from diversity of arbuscular *Mycorrhizal* fungi associated with some medicinal plants in Western Ghats of Karnataka region, India or Distribution of arbuscular *Mycorrhizal* fungi (AMF) in Terceira and São Miguel Islands (Azores)Biodiversity Data Journal 8: e49759 doi: https://doi.org/10.3897/BDJ.8. e49759

Acknowledgments

We sincerely acknowledge our mentor and Former Director of School of Life Sciences KBC North Maharashtra University, Jalgaon, MH, India late *Prof. Sudhir B Chincholkar* for his inspiration.

References

- Daniel BA, Skipper HD (1982) Methods of recovery and quantitative estimation of propagules from soil. In: Schenck NC (ed) Methods and principles of mycorrhizal research. The American Phytopathological Society, St. Paul, MN, pp 29–35
- Sutton JC, Barron GL (1972) Population dynamics of Endogone spores in soil. Can J Bot 50(9):1909–1914
- Gerdemann JW, Nicolson TH (1963) Spores of mycorrhizal Endogone speciesextracted from soil by wet sieving and decanting. Trans Br Mycol Soc 46(2):235–244
- 4. Phillips JM, Hayman DS (1970) Improved procedures for clearing roots and staining parasitic and vesicular-arbuscular mycorrhizal fungi for rapid assessment of infection. Trans Br Mycol Soc 55(1):158–161

Springer Protocols

Natarajan Amaresan Prittesh Patel Dhruti Amin *Editors*

Practical Handbook on Agricultural Microbiology



Springer Protocols Handbooks

For further volumes: http://www.springer.com/series/7657 Springer Protocols Handbooks collects a diverse range of step-by-step laboratory methods and protocols from across the life and biomedical sciences. Each protocol is provided in the Springer Protocol format: readily-reproducible in a step-by-step fashion. Each protocol opens with an introductory overview, a list of the materials and reagents needed to complete the experiment, and is followed by a detailed procedure supported by a helpful notes section offering tips and tricks of the trade as well as troubleshooting advice. With a focus on large comprehensive protocol collections and an international authorship, Springer Protocols Handbooks are a valuable addition to the laboratory.

More information about this series at http://www.springer.com/series/8623

Practical Handbook on Agricultural Microbiology

Edited by

Natarajan Amaresan, Prittesh Patel, and Dhruti Amin

Uka Tarsadia University, Surat, Gujarat, India

💥 Humana Press

Editors Natarajan Amaresan Uka Tarsadia University Surat, Gujarat, India

Dhruti Amin Uka Tarsadia University Surat, Gujarat, India Prittesh Patel Uka Tarsadia University Surat, Gujarat, India

ISSN 1949-2448 ISSN 1949-2456 (electronic) Springer Protocols Handbooks ISBN 978-1-0716-1723-6 ISBN 978-1-0716-1724-3 (eBook) https://doi.org/10.1007/978-1-0716-1724-3

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Science+Business Media, LLC, part of Springer Nature 2022

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Humana imprint is published by the registered company Springer Science+Business Media, LLC part of Springer Nature.

The registered company address is: 1 New York Plaza, New York, NY 10004, U.S.A.

Preface

Agricultural Microbiology is a part of the microbiology branch dealing with beneficial or harmful microbes associated with either plants or soil. This manual focuses on beneficial microbes dealing with soil fertility, microbial degradation of organic matter, soil nutrient transformations, and biocontrol agents. Nowadays, techniques involved in the study of beneficial microbes in agricultural microbiology toward enhancing global agricultural productivity are in trend. This manual covers a wide range of basic and advanced techniques associated with research on the isolation of agriculturally important microbes, identification, biological nitrogen fixation, microbe-mediated plant nutrient use efficiency, and biological control of plant diseases and pests. Introduction to each protocol explains the role/importance of chemicals involved, uniqueness, and protocol application. A proper understanding of the protocol helps the researchers to manipulate them as per their need.

This book is composed of seven parts with 52 protocol chapters. Parts I and II represent the importance, isolation, and purification methods of agriculturally important microbes and include mineral-solubilizing microbes. Part III deals with phytohormones quantitative protocols directly or indirectly associated with microbes. Parts IV and V provide deep insights into protocols for screening agriculturally important enzymes and compounds related to biocontrol activity. Part VI represents assessment methods of soil microbial activity by soil respiration. The final Part VII deals with protocols for selecting microbial strains for inoculant production and quality control ultimately representing commercial biofertilizers production criteria. This book will help postgraduate students, research scholars, postdoctoral fellows, and teachers belonging to different disciplines of Plant Microbiology and Pathology. Moreover, this manual may also serve as a textbook for undergraduate courses like Techniques on Plant-Microbe Interaction/Biological Control of Plant Diseases/Nutrient Use Efficiency.

Surat, Gujarat, India Surat, Gujarat, India Surat, Gujarat, India Natarajan Amaresan Prittesh Patel Dhruti Amin

Contents

	face stributors	v xi
Par	AT I ISOLATION AND IDENTIFICATION OF AGRICULTURALLY IMPORTANT MICROBES	
1	Methods for Isolation and Identification of Rhizobia	3
2	Isolation of Frankia from Casuarina Root Nodule Narayanasamy Marappa, Dhanasekaran Dharumadurai, and Thajuddin Nooruddin	15
3	Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum Bhavana V. Mohite and Satish V. Patil	25
4	Isolation and Identification of Azospirillum	35
5	Isolation and Identification of <i>Gluconacetobacter diazotrophicus</i>	41
6	Isolation and Identification of Nitrogen Fixing Bacteria: Azoarcus Species	47
7	Isolation and Identification of <i>Derxia</i> Species from the Soil Sample Harshida A. Gamit and Natarajan Amaresan	57
8	Isolation and Characterization of Enterobacter, Klebsiella,and ClostridiumV. Mageshwaran and K. Pandiyan	63
9	Isolation and Characterization of Genus Desulfotomaculum Mobini Pimpalse, Harshida A. Gamit, and Natarajan Amaresan	71
10	Isolation and Identification of Associative Symbiotic N ₂ Fixing Microbes: Desulfovibrio	77
11	Cyanobacteria Dhruti Amin, Abhishek Sharma, and Sanket Ray	85
12	Pseudomonas Sanket Ray and Harsh Patel	93
13	Isolation and Identification of Entomopathogenic <i>Bacillus</i> Species Preeti Parmar, B. K. Rajkumar, and Naresh Butani	99
14	Methylobacterium	111

viii	Contents

15	Isolation and Identification of <i>Beijerinckia</i>	119
16	Isolation of Streptomyces from Soil Sample Vishnu Raja Vijayakumar and Dharumadurai Dhanasekaran	127
17	Isolation and Identification of <i>Trichoderma</i> Spp. from Different Agricultural Samples	131
18	Extraction, Isolation and Culturing of <i>Mycorrhizal</i> Spores from Rhizospheric Soil	145
19	Isolation and Identification of <i>Metarhizium</i>	151
20	Isolation and Identification of Bacteriophage for Biocontrol Mitesh Dwivedi	161
21	Isolation of Bacterivorous Protozoan, Acanthamoeba Spp., as New-Age Agro Bio-Input Chandrashekhar D. Patil, Bhavana V. Mohite, and Satish V. Patil	173
Par	TT II ISOLATION OF MINERAL SOLUBILIZING MICROBES	
22	Isolation and Screening of Zinc Solubilizing Microbes: As Essential Micronutrient Bio-Inputs for Crops Satish V. Patil, Hemant P. Borase, Jitendra D. Salunkhe, and Rahul K. Suryawanshi	181
23	Isolation and Screening of Mineral Phosphate Solubilizing Microorganisms	187
24	Isolation and Identification of Potassium-Solubilizing Microbes Prittesh Patel and Swati Patel	193
25	Isolation and Identification of Sulfur-Oxidizing Bacteria Vimalkumar Prajapati, Swati Patel, Radhika Patel, and Vaibhavkumar Mehta	197
26	Isolation of Ammonia Oxidizing Bacteria Naresh Butani, Shruti Satashia, Hemanshi Kanpariya, and Preeti Parmar	203
27	Isolation and Identification of Nitrite-Oxidizing Microbes Prittesh Patel, Ami Naik, and Abhishek Sharma	219
28	Isolation and Identification of Iron-Oxidizing Microbes Ami Naik and Pooja Patel	223
29	Isolation and Identification of Phytin Mineralizing Microbes Swati Patel and Prittesh Patel	231
30	Isolation and Screening of Silicate Solubilizing Microbes: Modern Bioinputs for Crops Chandrashekhar D. Patil, Bhavana V. Mohite, Rahul K. Suryawanshi, and Satish V. Patil	237

31	I Isolation of Selenium Biotransforming Microbes as New Age Bioinputs Pradnya B. Nikam, Narendra Salunkhe, Vishal Marathe, Bhavana V. Mohite, Satish V. Patil, and Vikas S. Patil	
Par	T III ESTIMATION OF PHYTOHORMONES BY BENEFICIAL MICROBES	
32	Auxin Dixita Panchal, Jemisha Mistry, and Dhruti Amin	251
33	Abscisic Acid Natarajan Amaresan, A. Sankaranarayanan, and Dhruti Amin	257
34	Cytokinins Jawahar Ganapathy, Jemisha Mistry, and Dhruti Amin	263
35	Ethylene Ketankumar J. Panchal and Dhruti Amin	269
36	Gibberellin Dhruti Amin, Sanket Ray, and Abhishek Sharma	273
37	Brassinosteroids	277
38	Strigolactones: Extraction and Characterization	283
39	Estimation of Jasmonic Acid Using Non-pathogenic Microbes Jasmonic Acid Krutika S. Abhyankar and Monisha Kottayi	289
Par	TT IV SCREENING OF AGRICULTURALLY IMPORTANT ENZYMES	
40	Chitinase Purvesh B. Bharvad and Harsha J. Algotar	301
41	Glucanase Purvesh B. Bharvad and Harsha J. Algotar	309
42	Identification of Cellulase Enzyme Involved in Biocontrol Activity	317
43	Identification of Protease Enzymes Involved in Biocontrol Activity Vimalkumar Prajapati, Swati Patel, Sanket Ray, and Kamlesh C. Patel	323
44	Isolation and Screening of Naringinase Producing Microbes: As Industrial Inputs for Agro Waste Base Enzyme Industry Satish V. Patil, Jitendra D. Salunkhe, and Vishal Marathe	331
45	Isolation and Screening of Phytase Producing Microorganisms: An Essential Bioinput for Soil Fertility Bhavana V. Mohite, Kiran Marathe, Narendra Salunkhe, and Satish V. Patil	337

Par	тV	Identification of Compounds Involved in Biocontrol Activity	
46		rogen Cyanide ti Bhatt and Dhruti Amin	345
47		ophores sa Patel	351
48	for D	tion and Screening of ACC Deaminase-Producing Microbes Prought Stress Management in Crops	361
49		olysaccharides a Chandwani and Natarajan Amaresan	369
Par	T VI	Assessment of Soil Microbial Activity by Soil Respiration	
50		matic Analyses in Soils ar Bilen and Veysel Turan	377
Par	т VII	Selection of Microbial Strains for Inoculant Production and Quality Control	
51		tion of <i>Rhizobium</i> Strain for Inoculum Production	389
52		her Culture, Broth, and Peat Test uti Amin, Sanket Ray, and Vrushali Wagh	395
Ind	ex		405

Contributors

- KRUTIKA S. ABHYANKAR School of Science, Navrachana University, Vadodara, Gujarat, India
- HARSHA J. ALGOTAR . D. L. Patel Science College, Himatnagar, Gujarat, India
- NATARAJAN AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- DHRUTI AMIN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- HIMANSHU BARIYA Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- PURVESH B. BHARVAD . D. L. Patel Science College, Himatnagar, Gujarat, India
- KHYATI BHATT Post Graduate Department of Biosciences, Sardar Patel University, Gujarat, India
- SERDAR BILEN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Atatürk University, Erzurum, Turkey
- HEMANT P. BORASE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- NARESH BUTANI Department of Microbiology, Faculty of Science, Sarvajanik University, Surat, India
- SAPNA CHANDWANI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- SHREYA DESAI C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, India
- DHANASEKARAN DHARUMADURAI Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- MITESH DWIVEDI C. G. Bhakta Institute of Biotechnology, Faculty of Science, Uka Tarsadia University, Surat, Gujarat, India
- HARSHIDA A. GAMIT C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- JAWAHAR GANAPATHY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- TEJAS GOHIL Sabarmati Ashram Gaushala, Kheda, Gujarat, India
- HEMANSHI KANPARIYA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- MONISHA KOTTAYI School of Science, Navrachana University, Vadodara, Gujarat, India
- V. MAGESHWARAN ICAR-National Bureau of Agriculturally Important Microorganisms, Mau, Uttar Pradesh, India
- MAHADEVASWAMY University of Agricultural Sciences, Raichur, Karnataka, India
- NARAYANASAMY MARAPPA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- KIRAN MARATHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- VISHAL MARATHE N.E.S. Science College, Nanded, Maharashtra, India

- VAIBHAVKUMAR MEHTA Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- HARSH MISTRY Department of Life sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- JEMISHA MISTRY C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- BHAVANA V. MOHITE Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India
- AMI NAIK C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- VRUTUJA NAIK National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- PRADNYA B. NIKAM School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- THAJUDDIN NOORUDDIN Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- DIXITA PANCHAL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- KETANKUMAR J. PANCHAL Department of Animal Biotechnology, College of Veterinary Science and Animal Husbandry, Anand Agricultural University, Anand, India
- K. PANDIYAN . ICAR-Central Institute for Research on Cotton Technology, Mumbai, India
- PREETI PARMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- HARSH PATEL P. G. Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, Gujarat, India
- KAMLESH C. PATEL PG Department of Biosciences, Sardar Patel University, Anand, Gujarat, India
- NAFISA PATEL Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- POOJA PATEL . Government Medical College, Surat, Gujarat, India
- PRITTESH PATEL C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Gujarat, India
- RADHIKA PATEL PG Department of Biosciences, Sardar Patel University, Gujarat, India
- SWATI PATEL Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India
- TARUN KUMAR PATEL Department of Biotechnology, Sant Guru Ghasidas Government P.G. College, Dhamtari, Chattisgarh, India; Department of Biotechnology, Guru Ghasidas Vishwavidyalaya (a Central University), Bilaspur, Chattisgarh, India
- CHANDRASHEKHAR D. PATIL Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- SATISH V. PATIL School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India
- VIKAS S. PATIL University Institute of Chemical Technology, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- MOHINI PIMPALSE C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, India
- VIMALKUMAR PRAJAPATI Aspee Shakilam Biotechnology Institute, Navsari Agricultural University, Surat, Gujarat, India

- PRAVEEN RAHI National Centre for Microbial Resource, National Centre for Cell Science, Pune, India
- VIJAYAKUMAR VISHNU RAJA Department of Microbiology, School of Life Sciences, Bharathidasan University, Tiruchirappalli, Tamil Nadu, India
- B. K. RAJKUMAR Main Cotton Research Station (MCRS), Navsari Agricultural University (NAU), Surat, Gujarat, India
- SANKET RAY Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India
- NARENDRA SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- JITENDRA D. SALUNKHE School of Life Sciences, KBC North Maharashtra University, Jalgaon, Maharashtra, India
- A. SANKARANARAYANAN C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- SHRUTI SATASHIA Department of Microbiology, Atmanand Saraswati Science College, Surat, Gujarat, India
- ABHISHEK SHARMA C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India
- K. SOWMIYA University of Agricultural Sciences, Raichur, Karnataka, India
- RAHUL K. SURYAWANSHI Department of Ophthalmology and Visual Sciences, University of Illinois at Chicago, Chicago, IL, USA
- RASHMI THAKOR Department of Life Sciences, Hemchandracharya North Gujarat University, Patan, Gujarat, India
- VEYSEL TURAN Department of Soil Science and Plant Nutrition, Faculty of Agriculture, Bingöl University, Bingöl, Turkey
- VRUSHALI WAGH Department of Microbiology, Naran Lala College of Professional and Applied Sciences, Navsari, Gujarat, India



Isolation and Identification of Nonsymbiotic Azotobacter and Symbiotic Azotobacter Paspali–Paspalum notatum

Bhavana V. Mohite and Satish V. Patil

Abstract

Azotobacter is a renowned nonsymbiotic nitrogen fixer. Since the discovery of Azotobacter in 1901, it has magnetized microbiologists' attention for its interesting potential in agriculture for nitrogen fixation as well as synthesis of biologically active substances. It has distinctly enhanced effect on crop production in agriculture by which it deciphers the growing demand of food for ever-increasing population. The exploration of free-living Azotobacter sp. along with unique symbiotic A. paspali will be an attempt towards augmentation of soil fertility with enhanced crop yield. This chapter will brief the general strategy for isolation and identification of Azotobacter with elementary approach.

Key words Cyst, Pigment, Exopolysaccharide, Nitrogen-free medium, Biofertilizer

1 Introduction

Azotobacter belongs to Azotobacteraceae family, proteobacteria subclass including nonsymbiotic-free nitrogen fixers and frequently has habitat in soil and water together with sediments [1]. Azotobacter chroococcum is the foremost reported species of Azotobacter from Holland soil by Beijerinck [2]. Subsequently, various new variety of Azotobacter sp. has been reported from soil and rhizosphere.

The Azotobacter genus has seven reported species namely A. chroococcum, A. beijerinckii, A. vinelandii, A. paspali, A. armeniacus, A. nigricans, and A. salinestris [3]. The utmost quantity of DNA in Azotobacter in comparison with other bacteria may be due to larger cells of Azotobacter [1]. The mol% GC content of Azotobacter is 52 to 67.5. The amount of DNA and quantity of chromosomes is augmented together with ageing.

NifH gene is expansively sequenced gene used for identification of nitrogen fixing *Azotobacter* [4]. The *Azotobacter* sp. has the metabolic potential of fixation of atmospheric nitrogen into ammonia. The three discrete nitrogenase enzymes, molybdenum

Springer Protocols Handbooks, https://doi.org/10.1007/978-1-0716-1724-3_3,

Natarajan Amaresan et al. (eds.), Practical Handbook on Agricultural Microbiology,

(Mo) nitrogenase, vanadium (V) nitrogenase, and iron-only (Fe) nitrogenases, are the remarkable characteristic to study *Azoto-bacter* as fascinating nitrogen fixer with its noteworthy agriculture potential.

Azotobacter shows a mixotrophic, autotrophy, or heterotrophic mode of nutrition. The combined nitrogen-free medium with appropriate carbon resource is the ideal prerequisite for the Azotobacter growth. The optimum temperature of Azotobacter growth is 28–37 °C, but may differ as per the species. The acidic to alkaline pH range beginning from pH 4.8 up to pH 8.5 is applicable for Azotobacter growth.

Azotobacter plays a remarkable role among the free-living nitrogen fixing microorganisms as widely distributed in the natural habitat; soil, water, and sediments. The *Azotobacter* has proved as excellent bioinput for crops by nitrogen fixation and affecting the plant growth and yield, producing different plant growth promoting substances as well as stimulating the microflora of the rhizosphere [1].

Most of the Azotobacter sp. is from soil, slightly acidic to alkaline soil, but few are from water. The requirement for high phosphorous leads to prevalence mostly in fertile soil. One remarkable sp., A. paspali, was isolated from roots system of Bahia grass (Paspalum notatum cv Batatais), a tropical grass, due to accessibility of organic substances and appropriate pH by the plant in the rhizosphere. Azotobacter paspali grows in the rhizosphere of P. notatum, and nitrogen fixed by it may be transferred to Bahia grass and hence improve pasture growth. This restriction to the plant rhizosphere may be due restriction of utilization of wide variety of organic substances. A. paspali has the unique antagonistic property against the Gram-positive bacteria which is a beneficial property for life in the rhizosphere. Increases in the nitrogen content of the roots and in the total nitrogen content of the sand plant system were associated with successful Azotobacter colonization [5]. The A. paspali shows symbiotic highly specific diazotrophic association with P. notatum and hence it is interesting to culture and study it.

Azotobacter is straight rods having rounded ends becoming ellipsoidal or coccoid-shaped based on culture age and medium. The dimension is 2 or more $\times 4\mu m$ (diameter \times length). A. paspali is generally longer ($5 \times 10\mu m$ length) and filamentous (up to $60\mu m$ long). The cells are single but can present in pairs or irregular clumps like in A. paspali. The morphological type changed according to culture condition. This property along with inability to use several carbon sources, make it a unique species. The increase in carbon: nitrogen (C: N) ratio and culture age [6] causes aggregation of A. paspali cells in late logarithmic or stationary phase. A. vinelandii and A. paspali are the only Azotobacter sp. carrying nif, vnf, and anf genes producing either of the three nitrogenases enzymes based on Mo or V supply in the environment.

Azotobacter is nonmotile or motile with peritrichous flagella. The pigment production and cyst formation are the peculiar characteristics of Azotobacter apart from nitrogen fixation potential. The Azotobacter sp. undergoes encystment during late stationary stage with production of water soluble or insoluble type of pigments.

1.1 Isolation of Azotobacter sp. is practiced using specific designed medium based on its nutrition category as chemoheterotrophs and potential of fixation of dinitrogen. The enrichment technique is principally based on the potential of nitrogen fixation by aerobic/microaerobic way and utilization of organic substrates as energy source. The enrichment can be carried out by addition of specific stimulatory or inhibitory selective substrate into N₂-free medium such as erythritol or D-arabitol, L-rhamnose, ethylene glycol as carbon source for *A. vinelandii*, *O*-hydroxybenzoate, D-glucuronate or D-galacturonate, and L-tartrate, pH 6 or less for *A. vinelandii*, *A. beijerinckii*, caprylate for *A. armeniacus*, and 35–37 °C temperature for *A. paspali*.

The soil paste plate and silica gel method were conventionally described for isolation of *Azotobacter* sp., in which soil/silica gel is fortified with suitable carbon source and other nutrient elements, seeded with sieved soil and allowed to incubate [3].

The isolation technique comprised of utilization of various nitrogen-free agar medium such as Winogradsky's [7] nitrogen-free media, LG medium [8], Norris medium [9], Ashby's medium [10], and Burk medium [11]. These reported media are fairly similar, merely get differ with a few carbon sources and proportion of micro and macronutrient and minerals (Table 2) (*see* Note 1).

Apart from general N₂-free medium, some medium can be designed to make it selective with addition of particular constituent for particular isolation of *Azotobacter* species. For example, for isolation of *A. paspali*: the sucrose medium can be made selective with addition of 0.5% bromothymol blue and using rhizospheric sample from *Paspalum notatum*, for *A. beijerinckii* using α -hydroxyl benzoate, Tartarate and D-galaturonate with maintaining pH 4.9–5.5, for *A. vinelandii* isolation addition of erythritol, butanol, rhamnose, ethylene glycol, 0.1% phenol, and 10% sodium benzoate, for *A. chroococcum* pH need to be maintained at pH 7.0–7.5, for *A. nigricans* addition of citrate, *n*-valerate and for *A. salinetris* Burk medium is fortified with 1.0–2.0% sodium salt [12].

1.2 Identification of the Azotobacter The organism isolation after the enrichment culturing has increased the possibility of isolation of free-nitrogen fixer. The primary confirmation of *Azotobacter* genus is carried with principal morphological tests, i.e., cyst formation and pigment production. *Azotobacter* could be differentiated from residual nitrogen fixers based on simple characteristic property of cyst-forming potential.

1.2.1 Cyst Formation Azotobacter vegetative cells show rods to ovoid shape morphology, and consequently they may also present in larger clumps. In stress condition, the vegetative cells change to round, dormant cell structure referred to as cyst by the process of encystment. The formation of cyst is the leading criteria for taxonomic classification of *Azotobacter* [13]. *Azotomonas* and *Derecio Azomonas* are nitrogen fixers, but do not have the ability to produce cyst. The formation of cyst could be induced by particular carbon sources like ethanol, butanol, β -hydroxybutyrate, and isopropanol as a carbon source [7] (see Note 2).

1.2.2 Pigment Production The diffusible and fluorescent pigment production is characteristic property of *Azotobacter* sp. and can be studied in daylight or under ultraviolet light, respectively. The basal agar media of Thompson and Skerman [14] enriched with sodium gluconate could be used for diffusible pigment while Stainer and Scholte medium is used for the nondiffusible pigment. The colonies of *Azotobacter* appear first white, flat, and mucoid, later on become quite glossy, convex, although the type of medium and carbon sources varies the colony morphology [14]. The further incubation allows pigment production (Fig. 1). The identification of *Azotobacter* sp. could be carried out based on pigmentation type as unique type of pigment productured by specific *Azotobacter* sp. (Table 1).

1.2.3 IdentificationThe cyst formation property on N_2 -free medium like Burk's, Ash-
of Azotobacterof Azotobacterby's, and Norris proved that the isolate is belonging to Azotobacter
genus. The species-level identification is relying on range of pheno-
typic and biochemical investigation. The use of typical carbon
source, type and color of pigment, response to particular antibiotics
are basis for species-level identification of Azotobacter [12]
(see Note 3).

1.3 AzotobacterThe new species of Azotobacter named Azotobacter paspali [15] was
isolated using the silica gel plates, containing Winogradsky's salts
and calcium citrate as a carbon source from the rhizosphere soil of
Paspalum notatum. This name was later changed to Azorhizophilus
paspali [16]. The unique characteristic for identification of the
A. paspali sp. is younger filamentous long rods cells (5–10µm
length and 1.3–1.7µm in width). It produces red violet water-
soluble pigment or yellowish-green fluorescent colonies. Döberei-
ner [15] examined growth of A. paspali on N2-free modified and
Lipman medium [17] having sucrose as solitary carbon source and
bromothymol blue indicator.

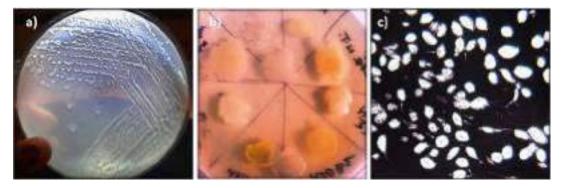


Fig. 1 (a) *Azotobacter* sp. on nitrogen-free medium, (b) Different *Azotobacter* sp. pigment production, and (c) *Azotobacter* with surrounding biopolymer by negative staining

Table 1

Azotobacter sp. with its specific type of p

Azotobacter species	Type of pigment
A. vinelandii	Yellow-green, fluorescent, water-soluble pigment
A. beijerinckii	Yellowish or cinnamon pigment
A. paspali	Yellow-green fluorescent or red-violet water-soluble pigment
A. chroococcum	Brown or blackish-brown
A. nigricans	Yellow nondiffusible pigment
A. armeniacus	Diffusible brown-black or red-violet
A. salinetris	Black-brown

A. paspali produces yellow color on a blue background on a sucrose minerals medium containing bromothymol blue indicator, representing the characteristic property of organic acid production (Table 2). After 48 h of incubation, yellow-centered colonies of *A. paspali* are appeared due to the medium acidification and hence resulting bromothymol blue assimilation. *A. paspali* is motile with peritrichous flagella while some strains have curli flagella. *A. paspali* can produce H₂S from thiosulphate and has potential to grow at 14° C.

A. paspali has the unique property of definite rhizospheric association with the wild grass; hence, it is considered to represent the symbiotic nitrogen association. Azotobacter paspali affects the growth and development of plant by appreciable increase in weight of roots and shoot [18]. A. paspali has specificity for a Paspalum notatum, a wild grass, along with some additional Paspalum sp., i.e., P. plicatulu and P. virgatum [19].

Table 2Differential characteristics of A. paspali

Characteristic	A. paspali
Motility by peritrichous flagella	+
Cell morphology Cells in pairs, irregular clumps Long filamentous cell Undulate edged unevenly convex colony with rough surface	+ + +
Water-soluble pigments Yellow-green fluorescent pigment	+
Brown-black to red-violet	+
Nitrogen fixation occurs at pH 5.0–5.5 6.0 6.5–9.5 10	_ d + +
Growth at temperature of 9 °C 14 °C 18 °C 32 °C 37 °C	 + + + + + +
Enzyme production Peroxidase Urease Oxidase	 + +
Production of H ₂ S from Thiosulphate Cysteine	+ d
Utilization of sole carbon source Fructose, glucose, acetate, pyruvate, fumarate, malate, succinate, α-oxoglutarate, lactate, DL gluconate, acetylmethylcarbinol Sucrose Propan-1-ol Maltose, trehalose, melibiose, raffinose, mannitol	+ + - +
Utilization of sole nitrogen source Ammonia Nitrate Glutamate	+ + -
Nitrate reduced to nitrite	-
Nitrogen fixation genes Nif Vnf Anf	+ + +
Mol % G + C (T_m) 2 Genome size	63.2–65.6 4.3–4.6 Mb

Key: +: Positive, -: Negative, d: variable

2 Materials

- 1. Soil sample from a fertile soil/macerated roots or leaves or other samples.
- 2. Sterile saline tubes (10 tubes) for serial dilution of the soil samples.
- Sterile N₂-free medium liquid enrichment medium (Burk's medium, Sergei Winogradsky's N₂-free medium, LG medium, Ashby's medium, Norris medium, Brown's medium, and Döbereiner sucrose mineral medium).
- Sterile N₂-free medium agar plates (Burk's medium, Sergei Winogradsky's N₂-free medium, LG medium, Ashby's medium, Norris medium, Brown's medium, and Döbereiner sucrose mineral medium).
- 5. Stain for cyst: violamine/acridine orange/mixture of neutral red, light green, SF yellowish.
- 6. 0.5 ml of 10% (v/v) glycerol or paraffin oil or in 7% dimethyl sulfoxide in 0.1% phosphate buffer (pH 7.0) for maintenance of culture.

3 Methods

- 1. Take 1.0 g of collected soil/macerated roots or leaves or other samples, add in 9.0 ml sterile saline, mix well by shaking, and then further dilute up to tenfold, i.e., 10^{-9} .
- Prepare N₂-free medium (Burk's N₂-free medium, Sergei Winogradsky's N₂-free medium, LG medium, Ashby's medium, Norris medium, and Brown's medium) in pure distilled water in clean 250 ml Erlenmeyer flasks and sterilize it (121 °C for 20 min at 15 psi).
- 3. The aliquots of diluted suspension (use last 3–4 dilutions, 10^{-6} to 10^{-9}) are incubated in liquid N₂-free agar medium (24–48 h at 30 °C). After the appearance of macroscopic growth, the culture is observed microscopically and cell morphology is observed.
- 4. The positive cultures are streaked from that liquid medium to the surface of specific N_2 -free medium agar plates with selective substance.
- 5. After incubation of 24–48 h, colonies will appear; further allow it to incubate for 3–5 days to observe the diffusible and fluorescent pigment production and about 2 weeks for cyst formation.

- 6. Cultures grown for 24–48 h on liquid and solid medium are studied for general morphology by microscopic observations like Gram staining, motility, and cultural characteristics.
- After incubation of 3–5 days, the plates are observed for pigment production (diffusible pigment in daylight and fluorescent pigment under ultraviolet light (364 nm wavelength)).
- The cyst may be stained with violamine/acridine orange/mix up of light green SF yellowish neutral red and observed under phase-contrast microscope.
- 9. The further identification of *Azotobacter* at species level includes biochemical tests using specific compounds/conditions in selective medium as mentioned earlier in text and in Table 2 (for *A. paspali*).
- 10. The *Azotobacter* isolates can be maintained by subculturing at bimonthly interval at Burk's or Winogradsky's medium with sucrose. The agar grown culture can be maintained by suspending in 0.5 ml of 10% (v/v) glycerol or paraffin oil or in 7% dimethyl sulfoxide in 0.1% phosphate buffer (pH 7.0).

4 Notes

- 1. In N₂-free Winogradsky's medium, the new cells of varying species are appeared enormously analogous and hence the old age culture should be compared for species-level identification.
- 2. The exopolysaccharide synthesis is the prominent characteristics of the cyst-forming *Azotobacter*.
- 3. All *Azotobacter* sp. are very susceptible to streptomycin and kanamycin/neomycin.

Acknowledgments

We acknowledge Prof. Ninfa Rosas-García, Centro de Biotecnología Genómica, Instituto Politécnico Nacional, Mexico for inspiring agrobiotech work and moral support. Author BVM is thankful to Principal, Bajaj College of Science, Wardha for the support and encouragement.

References

- Aquilanti L, Favilli F, Clementi F (2004) Comparison of different strategies for isolation and preliminary identification of *Azotobacter* from soil samples. Soil Biol Biochem 36 (9):1475–1483
- Beijerinck MW (1901) On oligonitrophilous bacteria. Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen 3:586–595
- 3. Becking JH (2006) The family *Azotobacteraceae*. In: Dworkin M, Falkow S, Rosenberg E,

Schleifer K-H, Stackebrandt E (eds) The prokaryotes. Springer, New York, pp 759–783

- 4. Zehr JP, Mellon M, Braun S et al (1995) Diversity of heterotrophic nitrogen fixation genes in a marine cyanobacterial mat. Appl Environ Microbiol 61(7):2527–2532
- Kass DC, Drosdoff M, Alexander M (1971) Nitrogen fixation by Azotobacter paspali in association with Bahia grass (*Paspalum notatum*). Am J Soil Sci Proc 35:286–289
- Abbass Z, Okon Y (1993) Physiological properties of *Azotobacter paspali* in culture and the rhizosphere. Soil Biol Biochem 25 (8):1061–1073
- Winogradsky S (1938) Etudes sur la microbiologie du sol et des eaux. Ann Inst Pasteur 60:351–400
- Lipman JG (1904) Soil bacteriological studies. Further contributions to the physiology and morphology of the members of the Azotobacter group. Report of the New Jersey State Agricultural Experiment Station 25:237–289
- 9. Norris JR (1959) The isolation and identification of Azotobacter. Lab Pract 8:239–243
- 10. Ashby SF (1907) Some observations on the assimilation of atmospheric nitrogen by a free living soil organism.-*Azotobacter chroococcum* of Beijerinck. J Agric Sci 2(1):35–51
- Wilson PW, Knight SC, (1952) Experiments in bacterial physiology. Burguess, Minneapolis, USA, 49
- 12. Patil SV, Mohite BV, Patil CD, Koli SH, Borase HP, Patil VS (2020) Azotobacter. In:

Amaresan N, Senthil KM, Annapurna K, Krishna K, Sankaranarayanan A (eds) Beneficial microbes in agro-ecology: Bacteria & Fungi. Academic Press, Cambridge, pp 397–426

- Jensen V, Petersen EJ (1954) Studies on the occurrence of *Azotobacter* in Danish forest soils. In: Royal Veterinary and Agricultural College Yearbook. Kandrup & Wunsch, Copenhagen, pp 95–110
- Thompson JP, Skerman VBD (1979) Azotobacteraceae: the taxonomy and ecology of the aerobic nitrogen-fixing bacteria. Academic Press Inc. (London) Ltd., London
- 15. Döbereiner J (1966) Azotobacter paspali sp. nov., uma bactéria fixadora de nitrogênio na rizosfera de Paspalum. Pesq Agropec Bras 1:357–365
- 16. Thompson JP, Skerman VBD (1981) Azorhizophilus paspali, comb. nov. invalidation of the publication of new names and new combinations previously effectively published outside the IJSB n.6. Int J Syst Bacteriol 31:215–218
- Lipman JG (1903) Experiments on the transformation and fixation of nitrogen by bacteria. Rep NJ St Agric Exp Stn:217–285
- Abbass Z, Okon Y (1993b) Plant growth promotion by Azotobacter paspali in the rhizosphere. Soil Biol Biochem 25(8):1075–1083
- Döbereiner J (1970) Fürther research on Azotobacter paspali and its variety specific occurrence in the rhizosphere of Paspalum notatum Flügge. Zentralb Bakteriol Parasint Infektion Hyg 124:224–230

As per Semester - wise New Syllabus of Rashtrasant Tukadoji Maharaj Nagpur University, Nagpur

mastry

A TEXT BOOK OF

-

B.Sc. SEMESTER - V

Paper I - Organic Chemistry Paper II - Physical Chemistry

AUTHORS

Dr. C. S. Bhaskar Dr. A. R. Yaul Dr. M. G. Dhonde Dr. M. R. Raghuwanshi Dr. N. C. Kongre Dr. P. B. Thakare Dr. U. P. Meshram Dr. V. Tekade

EDITORS

Dr. N. J. Siddiqui Dr. D. M. Borikar

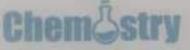
FREE misible

University Guestion Paper
 Periodic Table

Copyright @ DnyanPath Publication, Amravati (INDIA)

No part of this publication may be reproduced or distributed in any form or by any means, electronic, mechanical, photocopy, recording, or otherwise or stored in a database or retrieval system without the prior written permission of publishers. This edition can be exported from India only by the Publishers.

A TEXT BOOK OF



B.Sc. SEMESTER - V

Published by DayanPath Publication, Amravati (INDIA) The edition published in 15 August, 2021

ISBN: 978-93-87278-87-5





Reg. Office	: FFS-A, Block C, First Floor, Venus Plaza, Shegaon Naxa, V.M.V. Music		
	Amravati - 444 603 (Maharashtra)		
Our Network	: Maharashtra, Delhi, Gujrat, Chattisgarh, Telangana, Bihar.		
Visit us	www.dnyanpath.org		
Contact us	: dnyanpathpub@gmail.com		
Phone	: 08600353712, 09503237806		

Printed at - Shri Gurudeo Printers, Amravati. Mahatma Fule Sankul, Shegaon Naka, V.M.V. Road, Amravati - 444603 (Maharashtra)

Marketed by - Sachin Educational Traders (A Complete Education Solution) 10A, Shivrasik Nagar, Kathora Naka, Near Ring Road, Amravati - 444 604 Phone - 09405321101

Price : ₹ 200/-



This is to Certify that

DR. BHAVANA VISHWAS MOHITE

has participated in 2nd Virtual International Conference on Naturopathy, Nanotechnology, Nutraceuticals, and Immunotherapy in Cancer Research - 2021 (ICN3IC-21) Organized by Association of Cancer Education and Research (ACER), School of Life Sciences, B.S. Abdur Rahman Crescent Institute of Science & Technology, Chennai, India, in association with Purdue University, USA and sponsored by Tamil Nadu State Council for Science & Technology (TNSCST) and Microbiologist Society, India (MSI) held during 11th & 12th June, 2021.

Dr. P. Ashok Kumar Dr. Soumen Bera Dr. Neesar Ahmed Coordinator

Coordinator

Coordinator

Dr. S. Hemalatha Organizing Secretary

ISBN: 978-81-950236-5-3

2nd Virtual Annual International Conference on Naturopathy, Nanotechnology, Nutraceuticals and Immunotherapy in Cancer Research

11-12th June 2021

Organized by School of Life Sciences

Association of Cancer Education and Research (ACER) B.S. Abdur Rahman Crescent Institute of Science & Technology, Chennai, India

> In association with Purdue University, USA

Sponsored by Tamil Nadu State Council for Science & Technology (TNSCST) & Microbiologist Society, India (MSI)

> Editors Dr. S. Hemalatha, Dr. Neesar Ahmed Dr. Soumen Bera Dr. Ashok Kumar Pandurangan







Proceedings of ICN3IC-21

June 11 – 12, 2021

Editors

Dr. S. Hemalatha,

Prof & Dean, School of Life Sciences, B.S. Crescent Institute of Science & Technology, Vandalur, Chennai – 600048

Dr. Neesar Ahmed,

Assistant Professor (Sr.Gr.),

School of Life Sciences, B.S. Crescent Institute of Science & Technology, Vandalur, Chennai – 600048

Dr. Soumen Bera,

Assistant Professor (Sr.Gr.), School of Life Sciences, B.S. Crescent Institute of Science & Technology, Vandalur, Chennai – 600048

Dr. Ashok Kumar Pandurangan,

Associate Professor, School of Life Sciences, B.S. Crescent Institute of Science & Technology, Vandalur, Chennai – 600048

2nd Virtual International Conference on Naturopathy, Nanotechnology, Neutraceuticals and Immunotherapy Cancer Research- 2021

CCA192

MORINGA LEAVES PREPARATIONS AS MIRACULOUS WEAPON TO FIGHT AGAINST MALNUTRITION IN TRIBAL DURING COVID19 BACKGROUND

Satish V Patil^a, Vikas S Patil^b, Vishal Marathe^d and Bhavana V Mohite^c

^aSchool of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon 425001

^bUniversity Dept of Chemical technology, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon 425001

^cDepartment of Microbiology, Bajaj College of Science, Wardha 442001, Maharashtra, India ^dDeptartment of Botany, NES, science college Nanded. Maharshtra

Corresponding Author's email: satish.patil7@gmail.com

In our country around 194 million people go hungry every day. The COVID situation created this scenario more tragic. Most of the populations lack of feasibility and affordability for sufficient and nutritious food to maintain their health. 50% children and women from tribal community are anaemic. The COVID mediated unemployment created a problem of food security among poor, although it will fulfil but still the nutrient insecurity problem remains as it is. In pandamic situation immunity booster is remain the trending topic. Most of promoted advertisement of immunity boosting are merely of commercial interest. So there is urgent need to provide people knowledge of easily available natural green nutritious supplement available in their local area. This paper aims the same using a common plant "Moringa" popular as *drumstick* or *shevga* as a fruit vegetable all over the India. The miraculous nutritional potential of drumstick plant was already proved by worldwide research, but unknown for nutritional potential of its leaves. It is only known as fruit vegetable, but the Moringa leaves contain almost all essential nutrient growth factors. Hence with help of NASI, Allahabad, we validated locally occurring drumstick (Moringaoleifera) leaves nutritional potential and try to aware especially the tribal population about its use in day today food to eradicate common malnutrition problem. We selected some common food and developed common recipes including Moringa leaves. This will make easy, habitual use of Moringa leaves in day today food of tribal but also open new opportunities for entrepreneurs in tribal communities.

CCA195

SCREENING OF XANTHOMODIN PRODUCING XANTHOMONAS SP. AND ITS APPLICATIONS IN SUNSCREEN AS A NOVEL SUNLIGHT PROTECTING AGENT

Narendra S Salunkhe^a, Bhavna Mohite^b, Satish V Patil^a

^aDepartment of Microbiology, Bajaj College of Science, Wardha 442001, Maharashtra, India ^bSchool of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon 425001

Corresponding Author's email: Satish.patil7@gmail.com

Xanthomonas sp. are responsible for various *diseases* plant diseases like citrus can kerand oily spot disease in *citrus plant*. The pigment may be one of major character of xanthomonas and it may be act as major virulence factor for establishment of infection or pathogen in plant. It was also reported that the pigment protect these pathogens from physical and biological factors like UV light, temperature. By using routing microbiological techniques various bacteria isolated from the infected plant leaves and fruit part on the nutrient agar medium. Yellow pigmented bacterial colonies were consistently isolated from citrus plant leaves exhibiting bacterial streak symptoms. The causative organism was characterized as a aerobic, motile, gram-negative, rod-shaped organisms. The primary study on preferred carbon sources for high yellow pigment production was done on solid agar medium. It was primarily found that the nitrogen source play vital role pigment production while carbon source has negligible role in yellow pigment production. The pigment extracted was primarily characterized and tested for its UV protecting ability. The studies prove that Xanthomodin will be a future significant green source of UV protectant for commercial sunscreen

Keywords: Citrus, bacterial canker, Xanthomonas sp. EPS, Pigment

CCA196

SCREENING OF NARINGIN BIOTRANSFORMING MICROBES AND ITS APPLICATION

Jitendra D Salunke^a, Bhavana Mohite^b, Satish V Patil

^a Department of Microbiology, Bajaj College of Science, Wardha 442001, Maharashtra, India ^bSchool of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon 425001

Corresponding Author's email: satish.patil7@gmail.com

Nringin is Flavonoid glycoside especially present in garape, citrus like common fruits. The bitter taste of citrus fruit juice due to two flavonoid compounds specifically Naringin and limnonin. Naringin belongs to the flavonoid family compose of two sugar i.e. α-L-rhamnose and β -D-glucose with naringenine (4,5,7-trihydroxy-flavone). Naringinase s the enzyme responsible for biotransforamation of flyonoid Naringin and produce biologically active and medicinally important product such as narangenn and prunin . Naringinase enzyme also medicinal properties like anti-inflammatory, enhancement of the signaling pathway, anticancer like breast and bladder cancer to induce apoptosis in tumor cells and inhibited proliferation, it's help reduce liver disease. The enzymes are utilized in various new applications in food processing, Pharmaceutical, Agriculture, Lather, Paper industry, etc. The food and beverages industry facing bitter taste problems it's required more intense research debittering . Also most significant challenging factor in citrus fruit juice industry. The bitter taste leads to factor limiting for commercial undertaking proceed citrus fruit juice, Naringinase is the one of solution for this problem.In this investigation we are presenting screening of Naringinase producer microbes from citrus farm and citrus industry waste by using Patil et al 2019 iodine overlay method for Naringinase producer isolates. Effect of Carbon nitrogen sources on naringinase production and antimicrobial and antioxidant activity of crude product of naringin biotransformation

CCA197

SCREENING AND EXPLORATION RHIZOSPHRIC SELENOBACTERS FOR NANOSELENIUM SYNTHESIS AND ITS VARIOUS APPLICATIONS

Pradnya Nikam^a, Bhavana Mohite^b, Satish V Patil*

^aSchool of Life Sciences, KavayitriBahinabaiChaudhari North Maharashtra University, Jalgaon 425001

^bDepartment of Microbiology, Bajaj College of Science, Wardha 442001, Maharashtra, India

Corresponding Author's email: satish.patil7@gmail.com

Nanotechnology is the fast emerging and one of the most promising technologies of the modern era. The various types of metallic or non-metallic nanomaterials are replacing many of the conventional methods due to their exceptional and extraordinary properties. Selenium important micronutrients, has biologically important applications in therapeutics, agriculture, as health supplements and also as antimicrobial element. Se has been diversified in its various chemical and ionic forms within the environment. The available inorganic forms of Selenium in the soil areselenate (Se^{6+}), selenite (Se^{4+}) and selenide (Se^{2-}). The least available elemental Selenium (Se⁰) is very less toxic than its oxyanions; Selenite and selenite. The major organic forms of selenium areselenomethionine and selenocystine which are the aminoacids found in many of the proteins. The transformation of these toxic oxyanions to biologically safe, atomic form becomes important to make it more applicable with least or no toxic effects. This study particularly focuses onisolation and screening Rhizobium sp., for selenium transformations. Although various microorganisms have been reported for nanosynthesis, *Rhizobium* is safe non-pathogenic bacterium, reported for various beneficial activities. The organism was isolated from root nodules of different locations and screened for its selenium transformation potential. The isolates did show a red color colloidal suspension formed after 48 - 72 hrs of incubation. The organisms were tested for their maximum selenium tolerance. Their potential antimicrobial and antioxidant properties were also checked. The biotransformation was confirmed by ICP-AES analysis and also characterized by UV-Visible spectrophotometer. This *Rhizobium* assisted nano synthesis method can be commercialized by optimizing the physical factors in future, as it is completely an ecofriendly approach.

Keywords: Selenium oxyanions, bio-nano-factories, Se⁰Nanoparticles.

CCA199

SCREENING AND EXPLORATION OF MICROBE BASED BIO ATTRACTANT FOR MOSQUITO: ONE STEP TOWARDS NOVEL MOSQUITO CONTROL METHOD

Bhavana V Mohite^a, Amol Theng^b and Satish V Patil^c

^aDepartment of Microbiology, Bajaj College of Science, Wardha 442001, Maharashtra, India ^bDepartment of Zoology, Bajaj College of Science, Wardha 442001, Maharashtra, India ^cSchool of Life Sciences, Kaviyatri Bahinabai Chaudhari North Maharashtra University, Jalgaon 425001 Maharashtra, India

Corresponding Author's email: bhavnamohite1@gmail.com

The Chemical signals are very important key of successful life cycle of mosquitoes. The use of various chemical cues and signal were detected in various stages of mosquito life cycle. Chemical, visual and physical cues impact on the mosquito's biting and oviposition behavior. In view of the above significance of chemical cues or signals, we screened some microbes from the different preferred sites of mosquito's oviposition and biting. The basic idea behind the work is: some aquatic bacteria, actinomycetes and fungi may be a source of these chemical signals. Routine microbiological techniques were employed for isolation of microorganisms like actinomycetes, yeast and bacteria from the aquatic sites were Aedes agepty and other mosquito species eggs and larvae were detected. Besides these on the basis of survey, susceptible individuals for mosquito attracting and biting were analyzed for their skin microflora. The isolated microorganisms were tested for their mosquito attracting potential by simple pot assay in which the food colors were incorporated in sugar/honey solutions along with pure cultures of microbes and their metabolites. The purpose of food color is to determine attracting and feeding mosquito population towards the microorganism and their bioattracting metabolites. The numbers of mosquitoes attracted and feed were analyzed. On this basis of this experiment design, it was revealed that the oviposition is majorly driven by aquatic or marshy place residing Gram negative bacteria, while skin attraction was mainly due to organic and amino acid secreting bacteria like lactobacilli, micrococcus etc.

Innovative Research Trends in Science and Humanities

DNA Barcode: - New Approach for Conservation of Biodiversity

Ulka Malode-Bidwai Department of Microbiology Bajaj College of Science (Autonomous), Wardha

Kunal Kale Department of Biotechnology Bajaj college of Science (Autonomous), Wardha

Pratiksha Pohekar Department of Biotechnology Bajaj college of Science (Autonomous),

Wardha

Abstract

Overexploitation of resources, climate change, global warming, introduction of exotic species and genetically modified organisms, pollution, natural calamities and loss of sensitivity towards natural resources and the most vital anthropogenic activities are all responsible for loss of biodiversity. The diversity of plant species is at considerable risk from human activities that include habitat destruction, the introduction of plants and herbivores outside their native ranges, and anthropogenic climate change. The rates of plant extinctions are hundreds to thousands of times greater than rates of diversification. For the benefit of people and the planet, conservation of biodiversity is essential. It must be achieved through newer and swift technology to have universal acceptance. DNA Bar Coding is found to be novel and worth technique that would involve sequencing the DNA of unknown species and searching for a single sequence that could be easily isolated and used to swiftly differentiate species.

Keywords: DNA Barcoding, biodiversity, extinction of plant species

India is hot spot for biological diversity, Indian Western and Eastern Ghats, Himalaya Nilgiris mountains, Satpuras, Arivalli, Vindhya hills are rich in plant and animal, diversity still in good condition as compared to European countries as we have more than 47,000 species of plants in India. Biodiversity is the precious natural resource of nation and provide the stability of biosphere and supplies food, fodder, fiber, medicine etc. Although the origin of term biodiversity is unclear, what is clear is that biodiversity became the subject of considerable interest in both the popular and scientific literature during the last decade. It is a nonspecific term that is generally agreed to indicate the variability of all living organisms, and at all taxonomic levels, from the species to the ecosystem (Kate and Laird, 1999 and Abell, 2002).

The most useful definition of biodiversity is that given by the International Union for Conservation and Natural Resources: biodiversity is that given by the International Conservation and Natural Resources: biodiversity encompasses all life forms, ecosystems and ecological processes and acknowledges the hierarchy at genetic, taxon and ecosystem levels Biological diversity is usually considered at three different levels: genetic diversity, species diversity and ecosystem diversity. In molecular ecological terms, it can be defined as the number and distribution of different sequence types present in the DNA extracted from the community in the habitat (Dargan and Sarma 2001 and Kumar and Bhatt 2007).

Plant species, both those we know and those we don't, offer a tremendous resource of lities that could greatly add to the possibilities that could greatly add to the security of our food. But due to the human activity



Innovative Research Trends in Science and Humanities

Dr. Ninad S. Dharkar Dr. Kishor P. Suradkar Dr. Suruchi R. Kadu Dr. Dasharath M. Chavhan Dr. P. B. Ingle



Title: Innovative Research Trends in Science and Humanities

Volume: First-2021

Copyright © Editors

Dr. Ninad S. Dharkar, Assistant Professor, Department of Botany at S.P.M. Science College and Gilani Arts Commerce College Ghatanji Dist. Yavatmal (MS) India.

Dr. Kishor P. Suradkar, Assistant Professor and Head, Department of Botany at Indira Mahavidyalaya Kalamb Dist.Yavatmal (MS) India.

Dr. Suruchi R. Kadu, Assistant Professor, Department Botany at Brijlal Biyani Science College Amravati (MS) India.

Dr. Dasharath M. Chavhan, Assistant Professor, Department of Chemistry at Indira Mahavidyalaya Kalamb Dist. Yavatmal (MS) India.

Dr. P. B. Ingle, Assistant Professor, Department of Psychology at Indira Mahavidyalaya Kalamb Dist. Yavatmal (MS) India.

No part of this book may be reproduced or transmitted in any form by any means, electronic or mechanical, including photocopy, recording or any information storage and retrieval system, without permission in writing from the copyright owners.

DISCLAIMER

The author is solely responsible for the contents published in this book. The publishers or editors do not take any responsibility for the same in any manner. Errors, if any, are purely unintentional and readers are requested to communicate such errors to the editors or publishers to avoid discrepancies in future.

ISBN: 978-1-954461-44-4

Publishing Typeset & Distribution by: INSC PUBLISHING HOUSE (IPH) Pushpagiri Complex, Beside SBI Housing Board, K.M. Road Chikkamagaluru Karnataka Tel.: +91-7619574868 E-mail: iph@insc.in

IMPRINT: INSC Publishing House (IPH)

Indu Pal Kaur *Editor-in-Chief* Sandip V. Pawar Praveen Rishi *Editors*

Probiotic Research in Therapeutics

Volume 2: Modulation of Gut Flora: Management of Inflammation and Infection Related Gut Etiology



Indu Pal Kaur Editor-in-Chief

Sandip V. Pawar • Praveen Rishi Editors

Probiotic Research in Therapeutics

Volume 2: Modulation of Gut Flora: Management of Inflammation and Infection Related Gut Etiology



Contents

1	Gut Bacterial Dysbiosis and Its Clinical Implications	1
2	Probiotic Based Interventions for Improving Intestinal Health Kezia Devarapalli, Praveena Ganji, Chandrakala Gunturu, Prakasham Reddy Shetty, and Linga Banoth	29
3	Probiotics in the Prevention of Infant Infection	57
4	Animal Models for Probiotic Interventions Under Gut Inflammatory Conditions	85
5	Probiotics as Anti-Inflammatory Agents in Inflammatory BowelDisease and Irritable Bowel SyndromeVasudha Sharma and Pritika Sharma	123
6	Antibiotic-Associated Diarrhea and Update on ProbioticsRecommendationsDavid Elisha Henry and V. Venkateswara Rao	141
7	Potential Correlation Between Homeostasis Control and Tumor Microenvironment Regulation of Probiotic as a Therapeutic Agent to Manage Gastrointestinal Cancer	167
8	An Update on the Probiotic Usage in Bacterial Vaginosis Aishwarya Hattiholi, Shivani Tendulkar, and Suneel Dodamani	191
9	Indigenous Probiotic Lactobacillus Strains to Combat Gastric Pathogen Helicobacter pylori: Microbial Interference Therapy Nabendu Debnath and Ashok Kumar Yadav	215
10	Designing Probiotics and Its Clinical Applications	231

11	Probiotic Interventions for Oral Health	253
12	Probiotics Targeting Enteric Infections	271
13	Probiotics for Allergic Airway Infection and Inflammations Satish V. Patil, Bhavana V. Mohite, and Vikas S. Patil	295
14	Probiotics as Edible Vaccines	315
15	Probiotics for Atopic Dermatitis	335



13

Probiotics for Allergic Airway Infection and Inflammations

Satish V. Patil, Bhavana V. Mohite, and Vikas S. Patil

Abstract

Probiotics have expansively reported affecting the composition of the gut microbiota, and it opens promising areas of research for the discovery of probiotics in the prevention or treatment of infectious and inflammatory diseases. Probiotics exert multiple health effects such as immunomodulatory agents and activators of host defense pathways, influencing disease severity, and incidence. The normalization of the properties of unbalanced indigenous microflora by healthy gut microflora constitutes the rationale in probiotics therapy.

The probiotics microbiome is essential for the development of host immune responses, particularly within the context of allergy. The probiotics performance manifests itself in the normalization of the increased intestinal permeability, improvement of the intestine immunological barrier functions, and alleviation of the intestinal inflammatory response.

The effect of probiotics is based on the ability to differentially regulate the production of anti- and pro-inflammatory cytokines as well as the balance between types of T cell responses. Probiotics appear to be a feasible way to decrease the incidence of respiratory tract infections. Probiotics affect the lung

S. V. Patil

School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India e-mail: svpatil@nmu.ac.in

B. V. Mohite (🖂)

V. S. Patil

Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India e-mail: bvmohite@jbsw.shikshamandal.org

University Institute of Chemical Technology, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India e-mail: vs.patil@nmu.ac.in

immune response after the allergic airway inflammation due to an increase of T regulatory-dependent mechanisms. The proper development of bacterial colonization observed to downregulate the hypersensitivity reactions with alterations of the cytokine profile. There is a paucity of data regarding the study of the mechanism of probiotic. There is a need for a mechanism investigation of probiotic action to explore the putative benefit of respiratory disease.

Therefore, the current article focuses on the present scenario of the effect of probiotics on the immune system in allergic airway infections and inflammations.

Keywords

Probiotics · Gut microbiota · Allergy · Airway inflammation · Gut microflora

13.1 Introduction

13.1.1 Probiotics

The World Health Organization (WHO) defined probiotic as "living microorganisms in adequate amount confer the health benefits" (Food and Agriculture Organization of the United Nations, World Health Organization 2002). The phrase "probiotic" is a Greek term and means "for life." Originally it was termed as "substances secreted by one microorganism that stimulate the growth of another" (Lilly and Stillwell 1965). The redefinition by Parker (1974) coined the probiotics as "organisms and substances, which contribute to intestinal microbial balance."

The adapted narration by Fuller (1989) stated as "a live microbial feed supplement, which beneficially affects the host animal by improving its microbial balance." Marteau et al. (2001) provided the most accepted definition as "microbial cell preparations or components of microbial cells that have a beneficial effect on the health and well-being."

The Food Safety Department, World Health Organization (2005) defined probiotics as "live microorganisms, which, when administered in adequate amounts, confer a health benefit on the host." The international scientific community has admitted to this and has become the working definition of probiotics.

The most commonly used probiotics are *lactic acid bacteria* (LAB), particularly *Lactobacillus* and *Bifidobacterium* species, followed by the genera *Enterococcus*, *Streptococcus*, *Propionibacterium*, *Pediococcus*, *Escherichia coli*, and *Bacillus* (Szajewska et al. 2016). Few some yeast species are having potential as probiotics, e.g. *Saccharomyces cerevisiae* and *Saccharomyces boulardii* were utilized for the treatment of gastrointestinal diseases very often (Guarner et al. 2012; Sanders et al. 2013; Schreck Bird et al. 2017; Kerry et al. 2018). However, not all the bacteria can be probiotic, as they need to be strain-specific.

The probiotic produce is in the type of tablets, capsules, powders (which worked as a dietetic complement), and as a food component (e.g., kefirs, kombucha, tempeh, miso, yogurts, or a drug). The dairy products and functional foods are helpful for the restoration of healthy microbiota of the body and almost all adults, as well as children, consumed it (Reid 2015). Hence dairy probiotic has been commercialized all over the world in different forms. However, allergy and lactose intolerance are the main arrests of dairy probiotics. The milk proteins, casein and whey proteins may act as allergens (Kumar et al. 2015).

Among the food factors, the use of food dyes is also a major reason for food allergy. Various natural and synthetic dyes such as carmine, tartrazine, and so on are added to the food to enhance the aesthetic value but may cause adverse reactions of food coloring allergy (Laura et al. 2019).

Probiotics are the indigenous nonpathogenic bacteria that colonize the mammalian intestinal tract. 10% out of 10^3-10^4 bacteria/ml dwelling in the body are legitimate living bacteria (Sender et al. 2016). The probiotic bacteria colonize initially maternal vaginal and fecal bacteria flora with reductive potential to make an anaerobic condition to favor the development of *Lactobacilli* and *Bifidobacteria*.

13.1.2 Benefits of Probiotics

The gastrointestinal tract is one of the most microbiologically dynamic environments that assume a vital role in the working of the mucosal immune system (MIS). The consumed probiotic stimulates the immune response as well as signaling by intact bacteria or its cell wall structure (Galdeano et al. 2019).

The gut is the site where huge numbers of bacteria from the microbiota and from the intestine which get through food intake coexist with each other. The immune cells are associated with the lamina propria of the villi. This intestinal microbiota does not interrelate straightforwardly with the epithelial cells; however, the maturation and functionality of the immune cells are stimulated by this microbiota through their metabolites (Hooper et al. 2012).

The beneficial effects of probiotics have been widely used in improving the host well-being and for the treatment of diverse infectious and non-infectious pathologies in animal models. Specifically it is included: protection from infection (Park et al. 2017; Acurcio et al. 2017; Mallina et al. 2018), irritable bowel symptoms relief (Hungin et al. 2013), reduction in the gut inflammatory response (Fábrega et al. 2017), cancer prevention (Aragón et al. 2015; So et al. 2017), growth inhibition of *Helicobacter pylori* (Fujimura et al. 2012), and allergies prevention (Velez et al. 2015).

Even though probiotics have shown encouraging results in several health conditions in humans, such as diabetes, multi-drug resistant pathogens, irritable bowel syndrome (He et al. 2017; Abdelhamid et al. 2018; Majeed et al. 2018), extensive research is still essential to include probiotics into human health, nutrition, and regulation of diverse abnormalities.

13.1.3 How Probiotic Function for Immune System?

The primary clause for probiotic microbes is survival in the harsh conditions of the gastrointestinal (GI) tract and stomach of humans. There are various ways by which probiotic microbes modulate the immune system. Figure 13.1 presents a brief of the role of probiotics for the immune system to maintain the human health majorly include: i) Modulation of innate and adaptive immunity, ii) Growth inhibition of pathogenic bacteria, iii) Regulation of anti-inflammatory or pro-inflammatory cytokines, iv) Regulation of the gastrointestinal /mucosal immune system (Baldassarre et al. 2016).

The important properties of probiotics which help to maintain the body to exert the effects are capacity to stick to the epithelial cells, activation of innate and cytokine-mediated immune response by internalization of a fragment of probiotic bacteria inside the immune response stimulating, intestinal epithelial cells (IECs) (Galdeano and Perdigon 2004), strengthening of the intestinal barrier by increasing the number of Goblet cells which reinforce the mucus layer (De Moreno de LeBlanc et al. 2008).

Table 13.1 summarizes the diverse means to promote human health. In recent years, extensive research has been conducted on the role of probiotics in transforming the adaptive and innate immunity as a way to check or treat a wide variety of health conditions (Baldassarre et al. 2016).

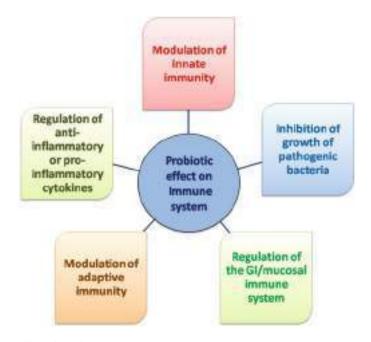


Fig. 13.1 Effect of probiotic on immune system

Sr.			
No.	Mechanism	Active component	Reference
1.	Inhibiting the growth of pathogenic bacteria through the synthesis of inhibitory compounds such as organic acid, bacteriocins, antimicrobial peptides [29].	Acetic acid, lactic acid lactacin B, plantaricin lysozyme, secretory phospholipase A2, defensins, cathelicidins	Bermudez-Brito et al. (2012); Russell and Diez- Gonzalez (1997); Nielsen et al. (2010); Sankaran- Walters et al. (2017)
2.	Reinforce intestinal barrier integrity in tight junction signaling by amplified gene impression	Actin, zonula occludens-1 (ZO-1), actinin, occludin	Resta-Lenert and Barrett (2003)
3.	Protection of epithelial barrier and increased the tight junction protein expression with activation of signaling pathway	p38 mitogen activated protein kinases (p38 MAPK) and extracellular signal regulated kinase (ERK)	Dai et al. (2012)
4.	Increase in Paneth cells, produce anti-inflammatory metabolites,	Regulatory T cells (Treg) / type 1 regulatory T (Tr1) cells	Liu et al. (2016)
5.	Activation of adaptive immune system	CD4+ regulatory T (Treg) cells, dendritic cells	De Moreno de LeBlanc et al. (2005)
6.	Induction of different cytokines.	Interferon gamma (IFN-γ), tumor necrosis factor-α TNF-α	Jiang et al. (2013)
7.	Increases the phagocytic and microbicidal activity of macrophages	Specific antibody production	Núnez et al. (2013)
8.	Decrease of IgE	Immunoglobulin (Ig) G, interleukin 10 (IL-10) and IFN-γ	Fu et al. (2017); Jerzynska et al. (2016)
9.	Improving lipid profiles, reduce blood glucose and insulin levels	High-density lipoprotein (HDL)-cholesterol	Shah and Swami (2017)
10.	Anti-cancer effect by combination of multiple mechanisms	Anti-genotoxic and anti- gene mutation function, enzyme inhibition	Russo et al. (2014)

Table 13.1 Summary of probiotic mechanisms to promote the human health

Now the probiotics have been commonly considered at therapeutical and clinical research level considering the relationship between the gut microbiome and immune disorders (Kothari et al. 2019), but the clear guidelines for the clinical application have yet to be established. This is particularly significant as the efficiency of probiotic supplementation may be reliant on the strain, dosing, condition, and duration of therapy (Toscano et al. 2017).

13.2 Role of Probiotics in Allergic Airway Infection

The normal healthy microflora constitutes the basis of probiotic therapy. Probiotics commonly mentioned as "good bacteria" or like a replacement for inhabitant stomach bacteria. Although the WHO recognizes probiotics as live microbes, when consumed in adequate quantity as an ingredient of food, it provides a health benefit to the host (Food Safety Department, World Health Organization 2005). At present, any item containing probiotics is viewed as a dietetic complement and is controlled by the principles and guidelines of the Dietary Supplement Health and Education Act of 1994. As indicated by it, the producer can give just common health declare for the manufactured food however it cannot express that any of the element in the product can fix, treat, or avoid illness (Alvarez-Olmos and Oberhelman 2001).

The dysbiosis, an inequity of the microflora constitution has adversely affected the health status. Three subcategory of dysbiosis have been recognized as below: (1) beneficial microbial agents loss, (2) spreading out of potentially harmful microorganisms, and (3) overall microbial diversity loss (Petersen and Round 2014).

Microbial dysbiosis has been concerned for different chronic inflammatory diseases, together with asthma (Sutherland and Martin 2007; Smits et al. 2016), chronic rhinosinusitis (CRS) (Hoggard et al. 2017; Aurora et al. 2013), Crohn's disease (Marin et al. 1983), and ulcerative colitis (Schmitz et al. 1999). The allergic infants reported an augmented number of *Clostridia* and a lower number of *Bifidobacteria* (Goktepe et al. 2005).

Amazingly all these persistent infections found to have altered membrane permeability and distorted functioning of epithelial barrier (Soyka et al. 2012; Steelant et al. 2016).

Probiotics have been publicized for a range of situation such as allergies, respiratory infections, including acute diarrhea, inflammatory bowel disease, and irritable bowel syndrome. This is been a choice to re-establish a healthy immune system (Dorval, 2015). Diverse probiotic strains and the mixing of microorganisms have a wide and differing range of clinical and immunologic potential and can manipulate gut microbiota in human beneficial ways (Table 13.2). The improved presence of probiotic bacteria in the intestinal microbiota has been found to correspond with defense from atopy (Moura et al. 2019). The predominance of hypersensitive ailment allergic diseases such as asthma, atopic dermatitis, and allergic rhinitis has expanded harshly over the past 2–3 decades in numerous nation, and sensitivities/allergies are presently most widely recognized chronic disease among youngsters all through the world (Tang et al. 2015).

The utilization of probiotic live forms could offer advantage to the patient's immunity, prompting improved management of the ailment, along with advanced lung functioning and reduced symptoms. Moreover, another mechanism of working of the probiotics comprised the enhancement in the epithelium membrane obstruction, hindrance of the adhesion of pathogens, binding to the intestinal mucosa, prohibition from pathogenic microorganisms by rivalry, and antimicrobial substance production (Bermudez-Brito et al. 2012).

Sr. No.	Strain	Mechanism	Outcome	Reference
	Strain		Outcome	
1.	L. plantarum, L. lactis, L. casei, Lactobacillus rhamnosus GG	Lesser IL-4 and IL-5 discharge	Reduced Th2 responses	Pochard et al. (2002)
2.	Lactobacillus rhamnosus GG and L. bulgaricus	Induction of IL1b, IL-6, IL-8, and TNF-a	Reduced Th2 responses	Niers et al. (2005)
3.	Lactic acid bacteria	Augmented IFN-g, TNF-a with IL-10	Reduced Th2 responses	Miettinen et al. (1998)
4.	Lactobacillus rhamnosus GG and B. lactis Bb12	Inducing transforming growth factor-β (TGF- β) secreting Tregs	Suppressed allergic symptoms	Feleszko et al. (2007)
5.	L. acidophilus W55	Stimulate functional FoxP3p(C) post- translational modification and Treg from CD25 cells	Supporting the species- specific effects of probiotics	de Roock et al. (2010)
6.	<i>Microbiota</i> <i>including</i> Bifidobacteria, lactobacilli,	Induction of mucosal IgA amount in addition to allergic B and T cell immunity	Modulation of allergy	Prescott and Björkstén (2007), Marschan et al. (2008), Galdeano et al. (2011)
7.	Lactobacillus reuteri	Reduced airway eosinophils, aryl hydrocarbon receptor (AHR) and TNF-a, IL-5 and IL-13 levels	Attenuate allergic airway disease	Forsythe et al. (2007)
8.	Commensal bacteria	Activation of DC andTh1 response	Stimulation of Th1 cytokines and, suppress Th2 response	Winkler et al. (2007)
9.	Commensal bacteria	Stimulation of mucosal IgA level	Allergen specific B and T cell response	Toh et al. (2012)

Table 13.2 Representative studies demonstrating Probiotic effect in allergy

Allergic ailment represents a convincing challenge for community well-being concern due to their expanding predominance in evolved and evolving nations. Universally roughly 1 thousand million people are facing allergic symptoms and could be reached to 4 thousand million in the following 3–4 decades (Spacova et al. 2018).

Allergy is defined as a hypersensitive reaction to a particular antigen called an allergen by an immunological reaction (Ring, 2014). The commonly found allergies are against pollen grains, animal dander, mites of dust, or specific foodstuffs. Allergies are caused due to an increase in the amount of IgE (Akdis and Agache,

2014). The repeated exposure to allergen elicits activation of mast cell and basophile cells and release of allergic mediators like histamine and leukotriene resulting in five cardinal signs of allergy that vary from mild symptoms like sneezing but may become serious like difficulty in breathing and hypersensitivity.

The number of studies carried out to study the probiotic as therapy for airway allergy such as a Stockert et al. (2007) in a pilot study investigated the influence of probiotics for asthma suffering kids and discovered improved lung functioning (peak of expiratory flow [PEF]) but no effect on the quality of lives and use of asthma treatment. Furthermore, Chen et al. (2010) observed progress in signs, lung functioning, and immunological criterion in probiotic taking kids. Liu et al. (2016) described the effect of probiotics to improve the curative impact of allergen-definite immune treatment in asthma sufferers. The in vivo trial in rats having airways allergic inflammation when inoculated with *Lactobacillus reuteri*, improvement of inflammation and airway over sensitiveness in the probiotic receiving group of animals was observed (Forsythe et al. 2007; Karimi et al. 2009).

Moura et al. (2019) confirmed the role of probiotics as a complementary therapy for asthmatic children and teenagers. Furthermore, study is suggested to confirm the effectiveness of probiotics in asthma medication, particularly indiscriminate restricted experimental groundwork and ultimate cluster investigation, to assemble supplementary evidence and information on the promising expected advantage of probiotics for asthma sufferers.

There is a growing indication to put forward that each probiotic strain does not have a single exclusive mechanics of activity regardless of common taxonomical rank (Sanders et al. 2018).

The substantial cluster of proof is demonstrating that probiotics amend the type 1 helper T cell (Th1)/ type 2 helper T cell (Th2) (Th1/Th2) parity to forestall the improvement of inflammation infections such as allergy. The gut microbiota is having a vital role in re-establishing Th1/Th2 immunity.

The altered Th2 phenotype prompts an elevated number of IgE and hence activation of a mast cell, which will result in sensitivity to hypersensitivity disorders. The Th2- dominant phenotype of newborn displays higher receptiveness to hypersensitivity diseases. Amazingly, commensal colonization is contributed to this attribute, showing the important function of gut microflora. Commensals likewise assume a job in managing immune cell allocation. Therefore, susceptibility was accounted in adults following intense antibiotic course (Walker and Iyengar, 2015).

Another point of view of the perceptions is demonstrated in the "hygiene hypothesis." This recommends less microbial contact through early stages due to the improved community cleanliness. It is one of the essential reasons for uplifted receptiveness to allergic hypersensitivity. Likewise, these studies set up the role of microflora to affect the allergy immune response (Sharma and Im, 2018).

13.3 The Rationale behind the Mechanism of Probiotics for Allergy

This new strategy is originated from diversified information revealing the pleiotropic impacts of probiotics that incorporate immunomodulation, re-establishment of intestinal imbalance of microbiota just as keeping up epithelium hindrance solidarity (Toh et al. 2012).

Inflammation is an elementary defense mechanism of the immune system against unknown immunogen; however, allergy is a host defensive immunity on recurring presentation to a particular unknown particle as an antigen, yet possibly harmful to the horde. Inflammation is a type of innate immune response against the foreign virulent particles associated with tissue rejuvenation. Probiotics presumably work as immunomodulators and actuator of human defense mechanism, that propose to impact disease seriousness and its rate. Probiotics therapy is established on the idea of typical fine microflora. The probiotic therapy is based on normalization of the properties of unbalanced indigenous microflora by specific strains of the healthy gut microflora. The advancement of mucosal and fundamental resilience depends on immunosuppressant action coordinated by T cells that assuage both Th1 and Th2 responses, mechanisms may incorporate regulation of the useful properties of the microbiota, epithelial cells, DC, and safe cell types.

The superior adhesion properties of probiotic facilitate the maintenance of the mucosal barrier and avoid the absorption of foreign particles and expansion of IgA mediated immune response. The proper development of bacterial colonization observed to downregulate the hypersensitivity reactions with alterations of the cytokine profile.

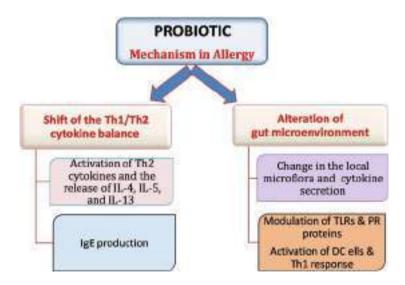


Fig. 13.2 Mechanism of probiotic in allergic reaction

Figure 13.2 describes the foremost activities of probiotic to undertake the airway allergic condition. The probiotic presents in the standardization of the extended intestinal permeableness and distorted gut microbial bionomics, development of the intestinal immunological fence job, and improvement of the response of gut inflammation.

The microbiome is fundamental for the advancement and learning of host immunity, mainly in the framework of allergic diseases. The use of probiotic influences the lung immunity followed by allergic airway infection due to augmentation of T regulatory-dependent mechanisms, however; whether this will impact the lung microbiota ruins to be determined. In reality, there is a need of elucidation of the mechanism of working of probiotic with assumed advantage for respiratory infections but there is paucity of data for airway microbiome composition.

13.3.1 Host Factors

The pathophysiology of susceptible illness, i. e. allergic disease results from an intricate series of actions including various ways of the natural immune response of innate and adaptive type. The allergic immune response involves stimulation of mast and basophil cells by IgE and succeeding allergen exposure resulted in allergic inflammation.

Host-associated factors can impact the working of the operation of the immune response in allergic hypersensitivity conditions and host and microorganisms communication (Laukens et al. 2016). Some vital characters are age, sex, host genetic structure, and microbiological status and can deviate in both human and animal investigation system (Laukens et al. 2016; Martín et al. 2017).

The pathogenic biofilm formation is the major host factor that leads to chronic infections. Biofilm formation is an accounted for about 65% and 80% of all microbial and chronic infections, respectively. Probiotic has the benefit as less cytotoxic than another quorum sensing (QS) suppressing agents and do not create strong pressure for resistance development like antibiotics. Hence probiotic could be an ideal alternative as an anti-virulent agent (Barzegari et al. 2020).

Probiotic prevents QS, biofilm formation, co-aggregation, and the survival of biofilm pathogens by interfering with biofilm formation and its quality. This is accomplished by decreasing the pH, competing for the adhesion sites with pathogens, and production of various antimicrobial agents like bacteriocin, hydrogen peroxide, and organic acids (Vuotto et al. 2014).

13.4 Allergy Prevention Studies with Probiotics

Current studies on meta-analysis of probiotics indicated a direct helpful impact on preliminary eczema impediment (Cuello-Garcia et al. 2015; Zuccotti et al. 2015), particularly to subsequent nativity to maternal and child to whom probiotics are administered. The probiotic will reduce the frequency of allergic sensitization with

perinatal intercession, which is not at all the condition for pre- or postpartum cure only (Zhang et al. 2016). Nevertheless, the support of probiotic for the avoidance of allergic airway disease is rare. There is no noteworthy outcome on the breathless incident or asthma improvement (Azad et al. 2013).

Lactobacillus probiotics strain is found to modulate the pro-inflammatory cytokines such as TNF- α , IL- 6, IL 1–10, and IL 1 β by activating the macrophage (Rocha-Ramírez et al. 2017).

Probiotic consumption could decrease the occurrence of respiratory tract infections. Aerosol delivery of probiotic diminishes tumor seeding in the lung and improves chemotherapy against exploratory metastases. Probiotic seems to defeat commensal microbes incited tolerance encouraging the maturation of resident antigen presenting cells.

The prevention or repairing of "leaky" epithelial barriers could serve for the pro-inflammatory response. The epithelium barrier is the primary defensive physical obstacle of the individual for the entry of detrimental particles like any pathogen, irritants, and allergic compounds (Koch and Nusrat, 2012).

Eventually, probiotics can influence the inflammatory response by contrasting the basis of pro-inflammatory motivation related with low-quality endotoxemia. Besides, probiotics and some of their emitted metabolic products can straightforwardly influence key pro-inflammatory pathways by acting as ligands for innate immune system receptors. Intercellular junctions, for instance, tight junctions (TJs), adherence junctions (AJs), and desmosomes contribute to the construction and continuation of the physical barrier.

The probiotics have an advantageous impact on epithelium barrier malfunction which is widely considered for the digestive tract. The example may include *Lactobacillus plantarum* MB452 which elevate the articulation of TJ-related genes by in vitro testing in well abdominal epithelial cells (IECs) (Anderson et al. 2010).

Related encouraging impacts were confirmed in case of probiotic strains such as *Lactobacillus rhamnosus* GG (Orlando et al., 2014), *L. plantarum* MB452 (Resta-Lenert and Barrett 2003), *Streptococcus thermophiles* ATCC19258, and the gramnegative probiotic strain *Escherichia coli* Nissle (Ukena et al., 2007; Zyrek et al. 2007) on abdominal epithelium barrier intactness and TJ expression. Moreover, certain Lactobacillus strains show the potential to elevate epithelium barrier integrity through the stabilization of AJs expression (Hummel et al. 2012).

In particular, to mention, the tested lactobacilli strain enhances the E-cadherin and b-catenin and diminishes the ample protein kinase C expression in T84 human abdominal epithelium cell line. Protein kinase C is the enzyme responsible for the disassembling of adherens junctions (AJs) (Hummel et al. 2012).

Several barrier-rebuilding characteristics of probiotics have also been verified in diverse in vivo models (Laval et al. 2015). There are at present scarce reports in the airways, relating the dictatorial characteristics of probiotics on the epithelium lining. The oral medication with *L. rhamnosus* CRL1505 could circumvent the polycytidylic acid [poly (I:C)]-induced improved permeable nature of the bronchoalveolar-capillarity barrier for in vivo experimentation, as find out by albumin levels in the lungs (Zelaya et al. 2014). This progress was associated to diminish

the activation and synthesis of pro-inflammatory cells and cytokines in the lungs (Zelaya et al. 2014). Alike results were reported by nasally managed *Lactococcus lactis* NZ9000, which could neutralize *S. pneumonia* prompted permeable nature of lung tissue (Medina et al. 2008).

The in vitro studies reported dose reliant augmentation in epithelium obstacle functioning and reduction in epithelium permeability by prompting Calu-3 lung epithelium cells with the artificial bacterial lipopeptide Pam3CysSK4. This is caused due to improved articulation of the TJ proteins claudin-1 and ZO -1 and a lessen articulation of occluding.

Even though asthma is customarily viewed as a Th2-type inflammatory situation, it has been perceived as a clinically varied illness. The microflora composition of the gut and respiratory system is related to asthma incidents, as indicated by several reports. But it is not yet satisfactorily explained how disturbance of microbiota influences sensitivity to allergic asthma. It is projected that some metabolites formed during the fermentation of dietary fibers like short-chain fatty acids (SCFAs) by commensal suppress allergic airway responses (Trompette et al. 2014).

The Th2 response in the lungs is suppressed by higher serum SCFA, mainly propionate amending DC progenitors by G-protein fixed receptor in reliant way in the bone marrow. Butyrate is the foremost potent immune regulatory metabolite among the SCFAs. Histone deacetylase (HDA) inhibition is the mechanism of action for the butyrate and propionate function, with improvement in the acetylating status of histone in the Foxp3 site (Furusawa et al. 2013; Arpaia et al. 2013) and inducing tolerogenic DCs to augment Treg generation(Arpaia et al. 2013).

The Clostridiaceae family bacteria *Lachnospiraceae* and *Ruminococcaceae* are too recognized for the synthesis of SCFAs by fermented dietary fibers in the colon and thus sustaining epithelial integrity and homeostasis. But how this will helpful for humans, it needs to be confirmed by clinical trials (Sharma and Im, 2018).

13.5 Recent Advances: Clinical and In Vivo Status

In recent years, several experimental studies have investigated the capability of probiotic bacteria to improve the virulent traits of hypersensitivity disorders.

The animate models can be utilized in support of the probiotic impact and their systems of activity. This is found unrealistic in humans inferable from obscure dangers and moral concerns. The impact of such components should take into account during the experimental preliminary plan. The information exploration will encourage the advancement of superior probiotic intercessions and reinforce the proof for probiotic application in the prevention and cure of human beings ailment.

The effect of the human being genotype has likewise been proposed to assume a vital function in the result of probiotic medications, incorporating these acted with regard to allergic diseases. Individual hereditary contrasts and inclination towards inflammatory diseases ought to be thought about while surveying the impacts of probiotics in a clinical setting. The age of an individual and the influence of their gut

microflora should take into consideration for the human being testing. All around elegant study and strong in vivo and in vitro investigation are thus essential to advance definite choice of probiotic species for anticipation and management of allergic illness (Spacova et al. 2018).

To date, in any case, a large portion of the study on probiotic has concentrated on the microflora only as opposed to the interaction between host and microbiota. Additionally, accessible information discards the significance of mycobiome and virome. The existing screening system is centered on the cytokine production efficiency and capability of microbes by using the cell lines or ex vivo isolated peripheral immune cells, even though they do not symbolize phenotypically to gut cells. It is a requirement to develop high-performance screening procedures to ensure the particularity and sufficiency of picked probiotics. The majority of the commercial probiotic preparations are a combination of different bacteria with distinct colony forming units (CFUs). The purpose is learning of the consumer about the period for the viability of a specific strain and number of bacteria in specific dose.

Consequently, experimental testing should be extended to incorporate distinct geological areas. Considering this, it is advantageous not to execute meta-analyses on shared records when diverse strains of bacteria were utilized since the impact can vacillate drastically between the strains. The use of probiotic strains ought not to be permitted except the security and effectors compounds of the probiotics are very well cleared (Sharma and Im, 2018).

13.6 Safety Considerations and Contraindications

Immunomodulatory action may rely upon strain-specific characters so ideal strain might be presented. Probiotics are viewed with a safe, rare short term side effect (Ciorba, 2012). Isolated instances of bacteremia or fungemia have been related to probiotics, though inhabitants information additionally shows that there is no across the board danger of these complications (Snydman, 2008). Microorganisms that are "generally regarded as safe" incorporate species of *Lactobacillus* and *Bifidobacterium* and definite yeast strains. Other bacteria, such as *Enterococcus* and *Streptococcus* strains, are not generally considered as safe, however they are utilized as probiotics (Snydman, 2008). Itself alert ought to be practiced in prescribing probiotics to these populaces. Studies examining probiotics are comparatively short in length, limiting the long term security information and the ability for the real unfavorable circumstance. To make the firm ends, an additional experimental trial examining the safety of probiotics must be led.

The inconsistent outcome may result from the contrast in the cogitation plan, readout, and patient understanding. One significant impediment for an absolute meta-analysis of probiotic studies is the implementation of diverse probiotic species and strains, mainly *Bifidobacterium* or *Lactobacillus* or combination of that (Zuccotti et al. 2015). The administered probiotic doses also change significantly among the study from 10^7 to 10^{10} or more (CFU)/day, and treatment duration may also vary from a while to quite a while (Zuccotti et al. 2015). Nonetheless, the

outcome can vary among experimental set up in any event, though utilizing a similar probiotic strain and a similar direction routine because of the hidden possible significance of host-associated parameters. Along these lines, clinical studies heterogeneousness stays a significant hindrance to the conceptualization of validation-based rules on probiotic execution in allergic hypersensitivity (Forsberg et al. 2016).

Probiotics are susceptible to environmental surroundings such as moisture, heat, light, and oxygen. Customers should take precaution for storing probiotic containing product and adhered to the guidelines shown on the item label. One specific impediment restraint is the inability to indicate probiotic bacterial used for the study, depiction study duplication troublesome. Furthermore, numerous consumer diet complement exclude the particular bacterial strain or dosage of a probiotic on the mark, which makes it difficult for the drug specialist to advocate a product, in any event when a lesson is properly directed to deliver viable outcomes. Albeit numerous experimental testings bolster the protected use of probiotics, more exploration is expected to decide the long lasting safety of these items.

13.7 Future Directions

In current circumstance where the ebb and flow proof was created from hardly any preliminaries with serious extent of heterogeneity, routine utilization of probiotics as an added substance on treatment in subjects with unfavorably susceptible aviation route ailments cannot be suggested.

But the probiotic consumption emerges as a practicable way to diminish the frequency of respiratory tract diseases. Probiotics can affect together innate and adaptive immunity. Knowledge-based strategies supported with experimental data can be applied for successful clinical trials such as selection of optimal probiotic strain, microbe-derived compounds, the duration of regimens, administration forms, doses, and long follow-up time, as well as identification of potential early biomarkers of treatment efficacy. Recently scientist from Ireland, UK, and the USA propose the microbiome, live biotherapeutic product as a predictor of COVID-19 outcomes, for targeted immunomodulation in COVID-19 infection like prevention of virus attachment on host cells as well for prevention or treatment such as use of specific *Lactobacillus* strain as immunostimulatory adjuvant for intranasal vaccination, genetically engineered antigen producing organism. Consequently probiotics has great scope for the allergic airway infections which needs to determine.

References

Abdelhamid AG, Esaam A, Hazaa MM (2018) Cell free preparations of probiotics exerted antibacterial and antibiofilm activities against multidrug resistant *E. coli*. Saudi Pharm J 26 (5):603–607

- Acurcio LB, Sandes SHC, Bastos RW et al (2017) Milk fermented by lactobacillus species from Brazilian artisanal cheese protect germ-free-mice against *Salmonella typhimurium* infection. Benefic Microbes 8(4):579–588
- Akdis CA, Agache I (2014) Global atlas of allergy. Eur Acad Allergy Clin Immunol
- Alvarez-Olmos MI, Oberhelman RA (2001) Probiotic agents and infectious diseases: a modern perspective on a traditional therapy. Clin Infect Dis 32(11):1567–1576
- Anderson RC, Cookson AL, McNabb WC et al (2010) Lactobacillus plantarum MB452 enhances the function of the intestinal barrier by increasing the expression levels of genes involved in tight junction formation. BMC Microbiol 10:316
- Aragón F, Carino S, Perdigón G et al (2015) Inhibition of growth and metastasis of breast cancer in mice by milk fermented with *Lactobacillus casei* CRL 431. J Immunother 38(5):185–196
- Arpaia N, Campbell C, Fan X et al (2013) Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. Nature 504:451–455
- Aurora R, Chatterjee D, Hentzleman J et al (2013) Contrasting the microbiomes from healthy volunteers and patients with chronic rhinosinusitis. JAMA Otolaryngol Head Neck Surg 139 (12):1328–1338
- Azad MB, Coneys JG, Kozyrskyj AL et al (2013) Probiotic supplementation during pregnancy or infancy for the prevention of asthma and wheeze: systematic review and meta-analysis. BMJ 347:f6471
- Baldassarre ME, Di Mauro A, Mastromarino P et al (2016) Administration of a multi-strain probiotic product to women in the perinatal period differentially affects the breast milk cytokine profile and may have beneficial effects on neonatal gastrointestinal functional symptoms. A randomized clinical trial. Nutrients 8(11):677
- Barzegari A, Kheyrolahzadeh K, Khatibi SMH et al (2020) The battle of probiotics and their derivatives against biofilms. Infect Drug Resist 13:659
- Bermudez-Brito M, Plaza-Díaz J, Muñoz-Quezada S et al (2012) Probiotic mechanisms of action. Ann Nutr Metab 61(2):160–174
- Chen YS, Lin YL, Jan RL et al (2010) Randomized placebo-controlled trial of lactobacillus on asthmatic children with allergic rhinitis. Pediatr Pulmonol 45(11):1111–1120
- Ciorba MA (2012) A gastroenterologist's guide to probiotics. Clin Gastroenterol Hepatol 10 (9):960–968
- Cuello-Garcia CA, Brożek JL, Fiocchi A et al (2015) Probiotics for the prevention of allergy: a systematic review and meta-analysis of randomized controlled trials. J Allergy Clin Immunol 136(4):952–961
- Dai C, Zhao DH, Jiang M (2012) VSL# 3 probiotics regulate the intestinal epithelial barrier in vivo and in vitro via the p38 and ERK signaling pathways. Int J Mol Med 29(2):202–208
- de LeBlanc ADM, Dogi CA, Galdeano CM et al (2008) Effect of the administration of a fermented milk containing *Lactobacillus casei* DN-114001 on intestinal microbiota and gut associated immune cells of nursing mice and after weaning until immune maturity. BMC Immunol 9(1):27
- De Moreno de LeBlanc A, Galdeano CM, Chaves S et al (2005) Oral administration of *L. casei* CRL 431 increases immunity in bronchus and mammary glands. Eur J Inflam 3(1):23–28
- De Roock S, Van Elk M, Van Dijk MEA et al (2010) Lactic acid bacteria differ in their ability to induce functional regulatory T cells in humans. Clin Exp Allergy 40(1):103–110
- Dorval E (2015) Probiotics as a treatment for infectious diseases. US Pharm 40(4):20-25
- Fábrega MJ, Rodríguez-Nogales A, Garrido-Mesa J et al (2017) Intestinal anti-inflammatory effects of outer membrane vesicles from *Escherichia coli* Nissle 1917 in DSS-experimental colitis in mice. Front Microbiol 8:1274
- Feleszko W, Jaworska J, Rha RD et al (2007) Probiotic-induced suppression of allergic sensitization and airway inflammation is associated with an increase of T regulatory-dependent mechanisms in a murine model of asthma. Clin Exp Allergy 37(4):498–505
- Food and Agriculture Organization of the United Nations, World Health Organization (2002) Guidelines for the Evaluation of Probiotics in Food. Joint FAO/WHO Working Group Report.

London, Canada: Food and. Agriculture Organization of the United Nations, World Health Organization. ftp://ftp.fao.org/es/esn/food/wgreport2.pdf. Accessed April 30 and May 1, 2002

- Food Safety Department, World Health Organization (2005) Modern food biotechnology, human health and development: an evidence-based study. World Health Organization, Geneva
- Forsberg A, West CE, Prescott SL et al (2016) Pre-and probiotics for allergy prevention: time to revisit recommendations? Clin Exp Allergy 46(12):1506–1521
- Forsythe P, Inman MD, Bienenstock J (2007) Oral treatment with live *Lactobacillus reuteri* inhibits the allergic airway response in mice. Am J Respir Crit Care Med 175(6):561–569
- Fu L, Song J, Wang C et al (2017) Bifidobacterium infantis potentially alleviates shrimp tropomyosin-induced allergy by tolerogenic dendritic cell (DC)-dependent induction of regulatory T cells and alterations in gut microbiota. Front Immunol 8:1536
- Fujimura T, Kinoshita J, Makino I et al (2012) Gastric cancer-state of the art in Japan. Rozhledy v chirurgii: mesicnik Ceskoslovenske chirurgicke spolecnosti 91(6):346
- Fuller R (1989) Probiotics in man and animals. J Appl Bacteriol 66(5):365-378
- Furusawa Y, Obata Y, Fukuda S et al (2013) Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells. Nature 504:446–450
- Galdeano CM, Perdigon G (2004) Role of viability of probiotic strains in their persistence in the gut and in mucosal immune stimulation. J Appl Microbiol 97(4):673–681
- Galdeano CM, Núñez IN, de LeBlanc ADM et al (2011) Impact of a probiotic fermented milk in the gut ecosystem and in the systemic immunity using a non-severe protein-energy-malnutrition model in mice. BMC Gastroenterol 11(1):64
- Galdeano CM, Cazorla SI, Dumit JML et al (2019) Beneficial effects of probiotic consumption on the immune system. Ann Nutr Metab 74(2):115–124
- Goktepe I, Juneja VK, Ahmedna M (2005) Probiotics in food safety and human health. CRC Press, Tylor and Francis
- Guarner F, Khan AG, Garisch J et al (2012) World gastroenterology organisation global guidelines: probiotics and probiotics. J Clin Gastroenterol 46(6):468–481
- He J, Zhang F, Han Y (2017) Effect of probiotics on lipid profiles and blood pressure in patients with type 2 diabetes: a meta-analysis of RCTs. Medicine 96(51):e9166
- Hoggard M, Biswas K, Zoing M et al (2017) Evidence of microbiota dysbiosis in chronic rhinosinusitis. Int Forum Allergy Rhinol 7:230–239
- Hooper LV, Littman DR, Macpherson AJ (2012) Interactions between the microbiota and the immune system. Science 336(6086):1268–1273
- Hummel S, Veltman K, Cichon C et al (2012) Differential targeting of the E-cadherin/beta-catenin complex by gram-positive probiotic lactobacilli improves epithelial barrier function. Appl Environ Microbiol 78:1140–1147
- Hungin APS, Mulligan C, Pot B et al (2013) Systematic review: probiotics in the management of lower gastrointestinal symptoms in clinical practice–an evidence-based international guide. Aliment Pharmacol Ther 38(8):864–886
- Jerzynska J, Stelmach W, Balcerak J et al (2016) Effect of Lactobacillus rhamnosus GG and vitamin D supplementation on the immunologic effectiveness of grass-specific sublingual immunotherapy in children with allergy. Allergy Asthma Proc 37(4):324–334
- Jiang K, Cao S, Cui JZ et al (2013) Immuno-modulatory effect of IFN-gamma in AMD and its role as a possible target for therapy. J Clin Exp Ophthalmol:0071
- Karimi K, Inman MD, Bienenstock J et al (2009) Lactobacillus reuteri–induced regulatory T cells protect against an allergic airway response in mice. Am J Respir Crit Care Med 179(3):186–193
- Kerry RG, Patra JK, Gouda S et al (2018) Benefaction of probiotics for human health: a review. J Food Drug Anal 26(3):927–939
- Koch S, Nusrat A (2012) The life and death of epithelia during inflammation: lessons learned from the gut. Ann Rev Pathol Mech Dis 7:35–60
- Kothari D, Patel S, Kim SK (2019) Probiotic supplements might not be universally-effective and safe: a review. Biomed Pharmacother 111:537–547

- Kumar BV, Vijayendra SVN, Reddy OVS (2015) Trends in dairy and non-dairy probiotic productsa review. J Food Sci Technol 52(10):6112–6124
- Laukens D, Brinkman BM, Raes J et al (2016) Heterogeneity of the gut microbiome in mice: guidelines for optimizing experimental design. FEMS Microbiol Rev 40(1):117–132
- Laura A, Arianna G, Francesca C et al (2019) Hypersensitivity reactions to food and drug additives: problem or myth? Acta Bio Med Atenei Parmensis 90(3):80–90
- Laval L, Martin R, Natividad JN et al (2015) *Lactobacillus rhamnosus* CNCM I-3690 and the commensal bacterium *Faecalibacterium prausnitzii* A2-165 exhibit similar protective effects to induced barrier hyper-permeability in mice. Gut Microbes 6(1):1–9
- Lilly DM, Stillwell RH (1965) Probiotics: growth-promoting factors produced by microorganisms. Science 147(3659):747–748
- Liu J, Chen FH, Qiu SQ et al (2016) Probiotics enhance the effect of allergy immunotherapy on regulating antigen specific B cell activity in asthma patients. Am J Transl Res 8(12):5256
- Majeed M, Nagabhushanam K, Arumugam S et al (2018) Bacillus coagulans MTCC 5856 for the management of major depression with irritable bowel syndrome: a randomised, double-blind, placebo controlled, multi-centre, pilot clinical study. Food Nutr Res 4:62
- Mallina R, Craik J, Briffa N et al (2018) Probiotic containing Lactobacillus casei, Lactobacillus bulgaricus, and Streptococcus thermophiles (ACTIMEL) for the prevention of Clostridium difficile associated diarrhoea in the elderly with proximal femur fractures. J Infect Public Health 11(1):85–88
- Marin ML, Greenstein AJ, Geller SA et al (1983) A freeze fracture study of Crohn's disease of the terminal ileum: changes in epithelial tight junction organization. Am J Gastroenterol 78 (9):537–547
- Marschan E, Kuitunen M, Kukkonen K et al (2008) Probiotics in infancy induce protective immune profiles that are characteristic for chronic low-grade inflammation. Clin Exp Allergy 38 (4):611–618
- Marteau PR, Vrese MD, Cellier CJ et al (2001) Protection from gastrointestinal diseases with the use of probiotics. Am J Clin Nutr 73(2):430s–436s
- Martín R, Chain F, Miquel S et al (2017) Using murine colitis models to analyze probiotics-host interactions. FEMS Microbiol Rev 41(1):S49–S70
- Medina M, Villena J, Salva S et al (2008) Nasal administration of Lactococcus lactis improves local and systemic immune responses against *Streptococcus pneumoniae*. Microbiol Immunol 52 (8):399–409
- Miettinen M, Matikainen S, Vuopio-Varkila J et al (1998) Lactobacilli and streptococci induce interleukin-12 (IL-12), IL-18, and gamma interferon production in human peripheral blood mononuclear cells. Infect Immun 66(12):6058–6062
- Moura JCV, Moura ICG, Gaspar GR et al (2019) The use of probiotics as a supplementary therapy in the treatment of patients with asthma: a pilot study and implications. Clinics 74:e950
- Nielsen DS, Cho GS, Hanak A et al (2010) The effect of bacteriocin-producing *Lactobacillus plantarum* strains on the intracellular pH of sessile and planktonic *Listeria monocytogenes* single cells. Int J Food Microbiol 141:S53–S59
- Niers LE, Timmerman HM, Rijkers GT et al (2005) Identification of strong interleukin-10 inducing lactic acid bacteria which down-regulate T helper type 2 cytokines. Clin Exp Allergy 35 (11):1481–1489
- Núnez IN, Galdeano CM, Carmuega E et al (2013) Effect of a probiotic fermented milk on the thymus in Balb/c mice under non-severe protein–energy malnutrition. Br J Nutr 110(3):500–508
- Orlando A, Linsalata M, Notarnicola M et al (2014) *Lactobacillus* GG restoration of the gliadin induced epithelial barrier disruption: the role of cellular polyamines. BMC Microbiol 14(1):19
- Park MS, Kwon B, Ku S et al (2017) The efficacy of *Bifidobacterium longum* BORI and *Lactoba-cillus acidophilus* AD031 probiotic treatment in infants with rotavirus infection. Nutrients 9 (8):887
- Parker RB (1974) Probiotics, the other half of the antibiotic story. Anim Nutr Healt 29:4-8

- Petersen C, Round JL (2014) Defining dysbiosis and its influence on host immunity and disease. Cell Microbiol 16(7):1024–1033
- Pochard P, Gosset P, Grangette C et al (2002) Lactic acid bacteria inhibit TH2 cytokine production by mononuclear cells from allergic patients. J Allergy Clin Immunol 110:617–623
- Prescott SL, Björkstén B (2007) Probiotics for the prevention or treatment of allergic diseases. J Allergy Clin Immunol 120(2):255–262
- Reid G (2015) The growth potential for dairy probiotics. Int Dairy J 49:16-22
- Resta-Lenert S, Barrett KE (2003) Live probiotics protect intestinal epithelial cells from the effects of infection with enteroinvasive *Escherichia coli* (EIEC). Gut 52(7):988–997
- Ring J (2014) What is allergy. In: Akdis CA, Agache I (eds) Global atlas of allergy. European Academy of Allergy and Clinical Immunology, Zurich, pp 2–3
- Rocha-Ramírez LM, Pérez-Solano RA, Castañón-Alonso SL et al (2017) Probiotic *Lactobacillus* strains stimulate the inflammatory response and activate human macrophages. J Immunol Res 4607491
- Russell JB, Diez-Gonzalez F (1997) The effects of fermentation acids on bacterial growth. In: Advances in microbial physiology, vol 39. Academic Press, pp 205–234
- Russo F, Linsalata M, Orlando A (2014) Probiotics against neoplastic transformation of gastric mucosa: effects on cell proliferation and polyamine metabolism. World J Gastroenterol WJG 20 (37):13258
- Sanders ME, Guarner F, Guerrant R et al (2013) An update on the use and investigation of probiotics in health and disease. Gut 62(5):787–796
- Sanders ME, Benson A, Lebeer S et al (2018) Shared mechanisms among probiotic taxa: implications for general probiotic claims. Curr Opin Biotechnol 49:207–216
- Sankaran-Walters S, Hart R, Dills C (2017) Guardians of the gut: enteric defensins. Front Microbiol 8(647):1–8
- Schmitz H, Barmeyer C, Fromm M et al (1999) Altered tight junction structure contributes to the impaired epithelial barrier function in ulcerative colitis. Gastroenterology 116(2):301–309
- Schreck Bird A, Gregory PJ, Jalloh MA et al (2017) Probiotics for the treatment of infantile colic: a systematic review. J Pharm Pract 30(3):366–374
- Sender R, Fuchs S, Milo R (2016) Revised estimates for the number of human and bacteria cells in the body. PLoS Biol 14(8):e1002533
- Shah NJ, Swami OC (2017) Role of probiotics in diabetes: a review of their rationale and efficacy. Diabetes 5:104–110
- Sharma G, Im SH (2018) Probiotics as a potential immunomodulating pharmabiotics in allergic diseases: current status and future prospects. Allergy, Asthma Immunol Res 10(6):575–590
- Smits HH, Hiemstra PS, Da Costa CP et al (2016) Microbes and asthma: opportunities for intervention. J Allergy Clin Immunol 137(3):690–697
- Snydman DR (2008) The safety of probiotics. Clin Infect Dis 46(2):S104-S111
- So SS, Wan ML, El-Nezami H (2017) Probiotics-mediated suppression of cancer. Curr Opin Oncol 29(1):62–72
- Soyka MB, Wawrzyniak P, Eiwegger T et al (2012) Defective epithelial barrier in chronic rhinosinusitis: the regulation of tight junctions by IFN- γ and IL-4. J Allergy Clin Immunol 130(5):1087–1096
- Spacova I, Ceuppens JL, Seys SF et al (2018) Probiotics against airway allergy: host factors to consider. Dis Models Mech 11(7):dmm034314
- Steelant B, Farré R, Wawrzyniak P et al (2016) Impaired barrier function in patients with house dust mite–induced allergic rhinitis is accompanied by decreased occludin and zonula occludens-1 expression. J Allergy Clin Immunol 137(4):1043–1053
- Stockert K, Schneider B, Porenta G et al (2007) Laser acupuncture and probiotics in school age children with asthma: a randomized, placebo-controlled pilot study of therapy guided by principles of traditional Chinese medicine. Pediatr Allergy Immunol 18(2):160–166
- Sutherland ER, Martin RJ (2007) Asthma and atypical bacterial infection. Chest 132(6):1962–1966

- Szajewska H, Konarska Z, Kołodziej M (2016) Probiotic bacterial and fungal strains: claims with evidence. Dig Dis 34(3):251–259
- Tang RB, Chang JK, Chen HL (2015) Can probiotics be used to treat allergic diseases? J Chin Med Assoc 78(3):154–157
- Toh ZQ, Anzela A, Tang ML et al (2012) Probiotic therapy as a novel approach for allergic disease. Front Pharmacol 3:171
- Toscano M, De Grandi R, Pastorelli L et al (2017) A consumer's guide for probiotics: 10 golden rules for a correct use. Dig Liver Dis 49(11):1177–1184
- Trompette A, Gollwitzer ES, Yadava K et al (2014) Gut microbiota metabolism of dietary fiber influences allergic airway disease and hematopoiesis. Nat Med 20(2):159–166
- Ukena SN, Singh A, Dringenberg U et al (2007) Probiotic *Escherichia coli* Nissle 1917 inhibits leaky gut by enhancing mucosal integrity. PLoS One 2(12):e1308
- Velez EM, Galdeano CM, Carmuega E et al (2015) Probiotic fermented milk consumption modulates the allergic process induced by ovoalbumin in mice. Br J Nutr 114(4):566–576
- Vuotto C, Longo F, Donelli G (2014) Probiotics to counteract biofilm-associated infections: promising and conflicting data. Int J Oral Sci 6(4):189–194
- Walker WA, Iyengar RS (2015) Breast milk, microbiota, and intestinal immune homeostasis. Pediatr Res 77(1–2):220–228
- Winkler P, Ghadimi D, Schrezenmeir J et al (2007) Molecular and cellular basis of microflora-host interactions. J Nutr 137(3):756S–772S
- Zelaya H, Tsukida K, Chiba E et al (2014) Immunobiotic lactobacilli reduce viral-associated pulmonary damage through the modulation of inflammation–coagulation interactions. Int Immunopharmacol 19(1):161–173
- Zhang GQ, Hu HJ, Liu CY et al (2016) Probiotics for prevention of atopy and food hypersensitivity in early childhood: a PRISMA-compliant systematic review and meta-analysis of randomized controlled trials. Medicine 95:e2562
- Zuccotti G, Meneghin F, Aceti A et al (2015) Probiotics for prevention of atopic diseases in infants: systematic review and meta-analysis. Allergy 70(11):1356–1371
- Zyrek AA, Cichon C, Helms S et al (2007) Molecular mechanisms underlying the probiotic effects of *Escherichia coli* Nissle 1917 involve ZO-2 and PKCζ redistribution resulting in tight junction and epithelial barrier repair. Cell Microbiol 9(3):804–816



ARTS, COMMERCE & SCIENCE COLLEGE, MAREGAON (ROAD) Affiliated to SGB Amravati University, Amravati (M.S.) NAAC Accrediated with B+ Grade ISO 9001: 2015 Certified

Ref: RAMAN-2021/Certificate/7468

10-Feb-2021

This is to certify that **S. S. Khandare, M. M. Shukla, M. G. Ingale** have presented a research paper entitled '*Green Synthesis of Silver Nanoparticles Using Apple and Banana Peel Extract, Their Characterization and Optimization*' in the RAMAN-2021 held during 7th to 9th Feb 2021, Organized By Department of Physics, Arts, Commerce and Science College, Maregaon, Maharashtra, India In Association with Department of Physics, P. N. College, Pusad, Maharashtra, India

Convener Dr. N. R. Pawar HOD, Physics ACS College, Maregaon

2013020

Chairman Dr. A. N. Gharde Principal ACS College, Maregaon



ABSTRACT BOOK





INTERNATIONAL E-CONFERENCE ON RECENT ADVANCES IN MATERIAL SCIENCE AND NANOTECHNOLOGY



7th - 9th February 2021

Organised by Department of Physics, Arts, Commerce and Science College, Maregaon, Maharashtra, India &

Department of Physics, P. N. College, Pusad, Maharashtra, India

In Association With

International Journal of Scientific Research in Science and Technology

Print ISSN: 2395-6011 Online ISSN : 2395-602X

Volume 8, Issue 1, January-February-2021

International Peer Reviewed, Open Access Journal

Published By



CONTENTS

Sr. No	Title / Author(s)
1	Advances in Evanescent Wave Based Fiber Optic Intrinsic Biosensors
-	Gajanan G. Muley
2	Role of Nano-phase Luminescent Materials for Environment Friendly Development
-	S. K. Omanwar
	Solar Cell Application of Silicon Decorated Copper Sulfide Nanocomposites
3	S. A. Waghuley
	Study of Conduction Mechanism in Polypyridine - Poly (Vinyl Acetate) Films by
4	Transference Number
	A. V. Kohale, N. D. Kolekar
	Wet Chemical Synthesis, Characterization and Biocompatibility Study of Hydroxyapatite
5	used as Biomaterials
	V. G. Thakare, V. B. Bhatkar, P.A.Wadegaonkar, S. K. Omanwar
6	Enlisting Some Ethnic Plants Species in Ner Region Dist. Yavatmal (M.S.) India
	Chavhan V. N.
	Novel molten salts synthesis and photoluminescence properties of Eu (III) doped Y_2O_3
<u>Z</u>	phosphor
	R. G. Korpe, K. A. Koparkar, N.S. Bajaj, S. K. Omanwar
8	Aldo-keto Gel Synthesis and Photoluminescence Properties of YVO4:Eu ³⁺ Microsphere
Ū	K. A. Koparkar, R. G. Korpe, G. V. Korpe, S. K. Omanwar
9	Solid Waste Management in India : Current Situation And Opportunities
	N. D. Kolekar, A. V. Kohale
10	Thermoluminescence Properties of KAl (SO ₄) ₂ : Eu ³⁺ Phosphors
10	S.R. Bargat, Yatish R. Parauha, G.C. Mishra, S. J. Dhoble
	Locally Rotationally Symmetric Bianchi type-I Cosmological Models with Modified
11	Holographic Ricci Dark Energy in f(T) Theory
	S. D. Katore, S. V. Gore
12	White Light Emission from La ₂ (MoO ₄) ₃ :Dy ³⁺ Phosphor

	Mechanical and Thermophysical Properties of Mg3TH7 (T= Mn, Tc, Re) Complex
83	Hydrides
	Sachin Rai, Navin Chaurasiya, Pramod K. Yadawa
	Preparation and Luminescence Characteristics of Eu2+ Doped SrAL2B2O7 Ceramic
84	Phosphor
	R. S. Palaspagar
	Green Synthesis of Silver Nanoparticles Using Apple and Banana Peel Extract, Their
85	Characterization and Optimization
	S. S. Khandare, M. M. Shukla, M. G. Ingale
	Nanocrystalline Mg0.6Cd0.4Al2O4 Thick Film Gas Sensor for the Detection of
86	LPG, CH4, CO2
	S. V. Agnihotri, V. D. Kapse
	Simple Route Synthesis of 3-Cynocoumarin by Knoevengel Condensation of
87	Benzaldehyde with Ethyl Cynoacetate over Si-Al-MCM-41
	Manish R. Deshpande, Mukund Joshi
00	Structural analysis of Lead Titanate Prepared by Wet Chemical Method
88	A. U. Bajpeyee, N. V. Galande, S. H. Shamkuwar
20	Current and Upcoming Innovations in Spintronics
89	Nikita Korde, Sandeep Waghuley
	Synthesis, Characterization and CO2 Gas Sensing Response of 5% SnO2 Doped
90	Polyaniline Nano Composite
	Hamjade PT, Khaire ND, Motke SG
01	Synthesis and Thermo Acoustical Dynamics of PMMA/Fe2O3 Nanocomposites
91	P. U. Tasalwar, P. D. Dhone, A. R. Bansod, O. P. Chimankar
	Structural Characterisation of conducting PPy/Rhodamine- B dye Composites
92	Synthesized By Simple Chemical Polymerization Method
	M. N. Pawar, N. S. Dixit, S. G. Khobragade, M.S. Dixit
	Solvent Extraction and Spectrophotometric Determination of Cobalt (II) With N,N'-BIS
93	(Salicylaldehyde) Ethylenediamine [Salen]
	S. M. Parkhi, V. P. Dhatrak, A. M. Nannaware
94	Preparation and Photoluminescence Properties of Eu ²⁺ Doped Lithium Alumino-Borate

Green Synthesis of Silver Nanoparticles Using Apple and Banana Peel Extract, Their Characterization and Optimization

S. S. Khandare*, M. M. Shukla, M. G. Ingale

Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India

ABSTRACT

Since last decade, green synthesis of metal nanoparticles such as silver nanoparticles is emerging as a new path to stand against various infections. The present study aims to synthesize silver nanoparticles by a green biological route, using an extract derived from apple (AE) and banana peel waste (BPE), which acts as a reducing and capping agent for reduction of Ag⁺ into Ag⁰ derived from silver nitrate (AgNO₃) showing development of reddish-brown and yellowish-brown colour respectively. Process of synthesis was optimized using several parameters. Optimum concentration of AgNO₃ was found to be for AE: 1.25mM; BPE: 0.75 mM, concentration of extract for AE: 500 µl; BPE: 200 µl, pH was for AE: 9.0 and BPE: 9.0, temperature for AE: 50° C and BPE: 50° C and incubation period for AE: 96 hr; BPE: 24 hr for optimum synthesis of silver nanoparticles. Characterization of the synthesized nanoparticles with UV-Visible spectroscopy reveals a characteristic absorption of surface plasmon resonance (SPR) peak at 422 nm and 422.4 nm respectively. Fourier transform infrared spectroscopy (FT-IR) affirmed the role of AE and BPE as reducing and capping agent of silver ions.

Keywords:- Silver nanoparticles, Green synthesis, Characterization, FTIR.





Green Synthesis of Silver Nanoparticles Using Apple and Banana Peel Extract, Their Characterization and Optimization

S. S. Khandare*, M. M. Shukla, M. G. Ingale

Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India

ABSTRACT

Since last decade, green synthesis of metal nanoparticles such as silver nanoparticles is emerging as a new path to stand against various infections. The present study aims to synthesize silver nanoparticles by a green biological route, using an extract derived from apple (AE) and banana peel waste (BPE), which acts as a reducing and capping agent for reduction of Ag^+ into Ag^0 derived from silver nitrate (AgNO₃) showing development of reddish-brown and yellowish-brown colour respectively. Process of synthesis was optimized using several parameters. Optimum concentration of AgNO₃ was found to be for AE: 1.25mM; BPE: 0.75 mM, concentration of extract for AE: 500 µl; BPE: 200 µl, pH was for AE: 9.0 and BPE: 9.0, temperature for AE: 50° C and BPE: 50° C and incubation period for AE: 96 hr; BPE: 24 hr for optimum synthesis of silver nanoparticles. Characterization of the synthesized nanoparticles with UV-Visible spectroscopy reveals a characteristic absorption of surface plasmon resonance (SPR) peak at 422 nm and 422.4 nm respectively. Fourier transform infrared spectroscopy (FT-IR) affirmed the role of AE and BPE as reducing and capping agent of silver ions. **Keywords:**- Silver nanoparticles, Green synthesis, Characterization, FTIR.

I. INTRODUCTION

Nanotechnology (sometimes abbreviated to "nanotech") is the study of manipulating matter on an atomic and molecular scale. Generally, nanotechnology deals with structures sized between 1 to 100 nanometre in at least one dimension, and involves developing materials or devices within that size (Kahn et al., 2006). Nanomaterials are leading circumference of the rapidly developing field of nanotechnology. They are attracting gradually because of their unique physicochemical properties, determined by their dimensions, shape, composition and crystallinity. They were employed for the treatment of water, in catalysis, field of medicine and biotechnology etc. Among synthetic nanomaterials so

far produced, the metallic nanoparticles (NPs) have distinctive properties like conduction of electricity, chemicals catalysis, high stability for and antimicrobial activities (Muzaffar and Tahir et al.,2018).Among all metal nanoparticles, Silver nanoparticles (AgNPs) are important materials that have been studied extensively, such nanoparticles possess unique electrical, optical as well as biological properties and are thus applied in catalysis, bio sensing, imaging, drug delivery, nano device fabrication and in medicine (P. K. Jain, Huang, El-Sayed, and El-Sayed et al., 2008; Nair and Laurencin et al.,2007). As Silver is a nontoxic, safe inorganic antimicrobial agent that is capable of killing about 650 types of disease causing microorganisms there is an

293

CHEMISTRY RESEARCH AND APPLICATIONS

Benzothiazole Preparation, Structure and Uses



Atakan Heijstek



In: Benzothiazole Editor: Atakan Heijstek ISBN: 978-1-53617-548-6 © 2020 Nova Science Publishers, Inc.

Chapter 3

ESIPT INSPIRED BENZOTHIAZOLE FLUORESCENT MOLECULES

Vikas Patil^{1,*}, Bhavana V. Mohite², Satish V. Patil³ and Sharad Patil⁴

¹University Institute of Chemical Technology, North Maharashtra University, Jalgaon, India ²Department of Microbiology, Bajaj College of Science, Wardha, India ³School of Life Science, North Maharashtra University, Jalgaon, India ⁴Department of Chemistry,SPDM, Arts, Commerce Science College, Shirpur, India

ABSTRACT

Excited state intramolecular proton transfer (ESIPT) based 2substituted benzothiazole fluorescent moleculeshavegainedconsiderable attention in the pastfew years a suseful molecule in high-tech and classical application. It was due to its desirable unique photo-physical properties induced due to the proton transfer in an excited state. The photo-physical

^{*} Corresponding Author's E-mail: vikasudct@gmail.com.





Sarvodaya Shikshan Mandal's

SARDAR PATEL MAHAVIDYALAYA, CHANDRAPUR

(Accredited with A Grade by NAAC with CGPA-3.05)

National Conference on Innovative Science and Technology for Sustainable Future

NCISTSF-2020

Saturday, 14th March 2020







This is to certify that

Dr. S. S. Khandare

Bajaj College of Science, Wardha has participated in National Conference on Innovative Science and Technology for Sustainable Future held on 14th March, 2020. He / She delivered Invited Talk / Chaired Session / Presented a Poster / Oral Paper entitled Decolorization and Degradation of Textile dye Direct Blue 15 by Bacterial Cultures Screened from Textile Industry

This conference was conducted on online platform on 31" May 2020

Dr. S. V. Madhamshattiwar Chief Organizing Secretary NCISTSF-2020

Dr. R. P. Ingole Principal & Chairman of Organizing Committee NCISTSF-2020





Sarvodaya Shikshan Mandal's

SARDAR PATEL MAHAVIDYALAYA, CHANDRAPUR

National Conference on Innovative Science and Technology for Sustainable Future

NCISTSF-2020



Saturday, 14" March 2020





This is to certify that

Mayur Ingale

Bajaj College of Science, Wardha

has participated in National Conference on Innovative Science and Technology for Sustainable Future held on 14th March, 2020. He / She delivered Invited Talk / Chaired Session / Presented a Poster / Oral Paper entitled

Decolorization and Degradation of Textile dye Direct Blue 15 by Bacterial Cultures Screened from Textile Industry

This conference was conducted on online platform on 31" May 2020

Dr. S. V. Madhamshettiwar Chief Organizing Secretary NCISTSF-2020

Dr. R. P. Ingole Principal & Chairman of Organizing Committee NCISTSF-2020



DECOLOURIZATION AND DEGRADATION OF TEXTILE DYE "DIRECT BLUE 15" BY BACTERIAL CULTURES SCREENED FROM TEXTILE INDUSTRY

S. S. Khandare¹, K. C. Sadrani², M. G. Ingale³

1. Associate professor (Microbiology), Bajaj College of Science, Wardha, ksuhas21@gmail.com

2. P.G. Student (Microbiology), Bajaj College of Science, Wardha, komalsadrani24@gmail.com

3. Assistant Professor (Microbiology), Bajaj College of Science, Wardha, mayuringale7@gmail.com

Abstract:

The textile industries use different types of dyes in their processing units which are liberated in natural water bodies through wastewater which causes serious damage to the environment. Chemical and physical method to treat effluent to remove color is expensive hence we study the biological method to degrade Direct Blue 15 dye. Here we isolate and identify four dye decolorizing bacteria by using selective enrichment culture in Bushnell-Haas (BH) medium amended with co-substrate glucose, yeast extract, and mgL⁻¹ Direct Blue 15 dye. The isolates were identified as *Staphylococcus*spp., *Pseudomonas* spp., *Acinetobacter* spp.and *Bacillus* spp. Among these, *Pseudomonas*spp. was found to be the most efficient dye degrader with 72 % dye degradation efficiency. Percent dye degradation efficiency shown by *Staphylococcus*, *Bacillus*, and *Acinetobacter* was 66 %, 64%, and 28 % at optimum temperature of 30^oC and pH 7 respectively. As compared to individual bacteria, enriched bacterial consortium was found to degrade the dye more efficiently with 88 % dye degradation efficiency.

Keywords: Direct Blue 15 dye, Decolorization, Degradation, Textile industry

I. Introduction:

The textile industry plays an important role in the world economy as well as in our daily life, but at the same time, it consumes large quantities of water and generates large amounts of waste water. Dyes have been extensively used in many textile industries worldwide. Dyes are compound that absorb light with wavelength in the visible range, i.e., 400 -700nm. These are composed of a group of atoms called chromophores which are responsible for the dye color [1].The chemical reagents used in the textile sector are diverse in chemical composition ranging from inorganic to organic molecules [2].During the process of dyeing 10-15% dyes remains unused and is discharged in the water bodies with improper treatment. Globally the concentration of these dyes in the water bodies constitutes around 2, 80,000 tons per year. Besides forming toxic compounds these also create anaerobic conditions and unavailability of light to the aquatic life [3].Few of the dyes alone or in combination with hazardous chemicals may turn carcinogenic and may cause various health hazards [4].Dyes are recalcitrant by design and not readily amendable to common treatment methods, imposing a challenge for closed water systems therefore the treatment of dyes is a serious concern. Extensive research in the field of biological azo dye decolourization has shown promising results[5]. Several methods are being employed for removal of these dyes from water bodies. For these purpose biological and nonbiological systems are in effect. The biological systems are more preferred as these are eco-friendly and economical[6].

In the biological methods, the microbes such as bacteria, fungi and algae are being used for thewastewater treatment, which could be a viable option as low-cost and eco-friendly technology. There are various microorganisms found in the contaminated environment, have potential to decolorize and even completely mineralize many dyes from the wastewater efficiently under certain environmental conditions have been reported by various researchers[7]. Among synthetic organic dyes, azo dyes are the most widely used, and they account for 60–70% of the total consumption of dyes. Furthermore, it was proven that most azo dyes and their metabolites can generate toxic, carcinogenic, mutagenic, and teratogenic effects on human health and the environment [8, 9 & 10]. According to the survey of Ecological and Toxicological Association of Dyes and Organic Pigments Manufacturers (ETAD) out of 4000 kinds of dyes, diazo Direct dyes demonstrated highest toxicity[11]. The intermediate aromatic amines and acrylamides generated in the metabolism process of benzamine-derived azo dyes have serious carcinogenic effects [12 & 13]. Therefore, azo-dye wastewater is a major concern, and the removal of azo dyes is extensively explored. Thus, the present study was aimed to isolate and characterize indigenous bacterial strain, capable of decolorizing Directblue 15 azo dye commonly used in dyeing industry of Wardha, Maharashtra, India. Additionally, effect of various environmental parameters on decolorization of the dye by isolated bacterial strains was studied.

II. Materials and Methods:

Sample Collection

The dye effluent was collected from a dyeing industry located in Gol Bazar, Wardha city. The effluent samples were collected in sterilized can and transported to the laboratory and were stored at 4° C till further analysis.

Enrichment, isolation and screening of dye decolorizing bacteria

The isolation was carried out in Bushnell-Haas medium. The medium composition was KH2PO4 - 0.1%, K2HPO4 - 0.1%, MgSO4.7H2O - 0.02%, CaCl2.2H2O - 0.002%, NH4Cl - 0.1%, NH4NO3 - 0.1%, NaCl - 0.01%, and FeCl3-6H2O - 0.005% and the pH wasadjusted to 7.0. For enrichment of dye decolorizing bacteria, 10 mL of the effluent sample was inoculated in 90 mL sterilized Bushnell-Haas medium(BH) supplemented with filter sterilized 100 mgL⁻¹ 1 Direct blue 15 azo dye. After 3 days of incubation (37°C, 150 rpm) 10 mL enriched broth was transferred to fresh dye supplemented BH medium and incubatedat the same condition for another 3 days. Such repeated sub culturingwasperformed for 3 times for the enrichment of putative dye decolorizingbacteria. Serial dilutions of the enriched broth were madeup to 10⁻⁶dilutions and aliquots from each dilution were platedon BH agar medium amended with 100 mg L⁻¹ of dye mixtureand incubated at 37^oC for 48 h. After incubation, morphologicallydistinct colonies were picked up and purifiedthrough repeated streaking on the same medium. The purified bacterialisolates were maintained in nutrient agar slants as pure culture and preserved for long term[14].

Identification of the selected isolates

Bacterial isolates able to grow profusely at 100 mgL⁻¹ of dye concentration were considered as promising dye decolorizer and subsequently, characterized based on their morphological,cultural, physiological and biochemical characteristics[15]. The isolates were then tentatively identified by comparing the test results with Bergey's Manual of Determinative Bacteriology [16].

Dye decolorization assay

Decolorization assay was performed by inoculating the isolates in dye containing BH medium followed by the centrifugation of the broth afterevery 24 hrsandabsorbance of the supernatantwas recorded at 600 nm against a blank.

Analysis of decolorization efficiency

For dye decolorization experiment, Erlenmeyer flasks containing 50 mL of sterilized BH medium (pH 7.0) amended with Direct Blue 15 dye to a final concentration f 100 mg L⁻¹ were inoculated with 10% (v/v) inoculums of each isolate as well as the developed consortium and incubated for 3 days (at 37^{0} C with 150 rpm). Control was maintained without inoculation. After 24, 48 and 72 hrs of incubation the culture broth was centrifuged (10,000g, 15 min at 4^{0} C) and absorbance of the supernatant was recorded at 600 nm. The decolorization activity in terms of (%) decolorization was calculated according to the following formula given by Chen et al. [17]

Decolourization (%) =
$$\frac{\text{lnitial} - \text{final}}{\text{lnitial}} \times 100$$

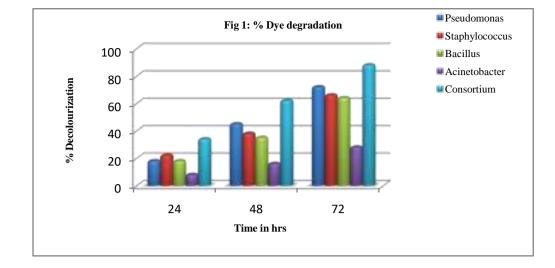
Effect of pH and temperature on the decolorization of dyes

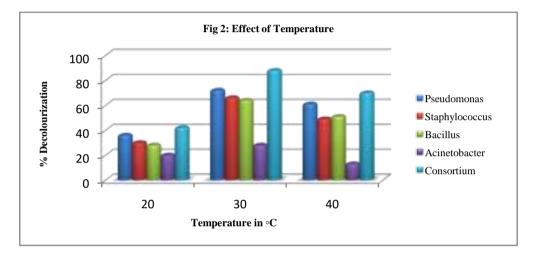
In order to study the effect of pH and temperature, the sterilized BH medium was amended with 100 mg/L of filter sterilized Direct blue 15 dye. The medium was maintained at different pH: 5,6, 7, 8. A volume of 1 mL of overnight culture was inoculated in the flasks and incubated in a shaker at 37^oC. Theeffect of temperature was studied by inoculating overnightculture and incubating in a shaker at 20^oC, 30^oCand 40^oC. The measurement of decolorization of the total dye concentration was performed at an interval of 24 h for 3 days.

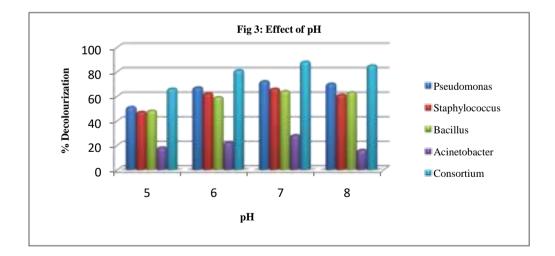
III.Results and Discussion:

In present studyselected bacterial isolates and developed consortium were tested for their ability to decolorize a commonly used textile dye Direct blue 15. In order to isolate dye decolorizing bacteria, the effluent sample was inoculated in BH agar medium amended withDirect blue 15. A total of 4 morphologically distinct bacterial colonies were isolated and screened out by repeated sub culturing method on dye amended BH agar medium. During screening process growth of isolates on dye supplemented BH agar medium in the form of white colonies considered as a positive result for screening of potent dye decolorizers[18]. The selected bacterial isolates were characterized on the basis of their cultural, morphological, physiological and biochemical characteristics as presented in Table 1. All these characteristics were then compared with the Bergey's Manual of Determinative Bacteriology and the isolates were provisionally identified as Psedomonas sp., Staphylococcus sp., Bacillus sp.and Acinetobactersp.After preliminary screening all these isolates were individually tested for their dye degradation efficiency for three days. Among the four isolates, Pseudomonas was found to be the most efficient dye degrader species with percent dye degradation efficiency of 72 % after three days at 30° C optimum temperature and pH 7. While percent dye degradation efficiency for Staphylococcus, Acinetobacter and Bacillussp. were found to be 66 %, 64 % and 28 % respectively. On the other hand enriched bacterial consortium could efficiently degrade the dye up to 88 % at optimum temperature of 30° C and pH 7 in three days. Temperature and pH found to influence dye degradation efficiency greatly.At 20°C Pseudomonassp. showed 36 % dye decolorization. Decolozization efficiency of Pseudomonas sp. increased significantly to 72 % at 30° C. But at40° Cdecolozization efficiency of Pseudomonas sp.was reduced to 61%. Dye degradation efficiency of consortium was also found to increase from 42 % to 88 % after increase in temperature from 20[°] C to 30[°] C. Hence temperature plays crucial role in dye degradation. Similar effect of temperature was also found on other isolates and 30° C was found to be the optimum temperature for all the isolates as well as for consortium. As far as the pH is concerned, 7 pH was found to be optimum for all the isolates as well as consortium. Increase or decrease in pH than 7 results in decrease in percent dye degradation. As compared to individual isolates, bacterial consortium showed significant hike (16 %) in dye degradation.

ISSN NO:2347-6648







Characters	Test	Pseudomonas	S. aureus	Bacillus	Acinetobacter
Characters	Test	sp.	sp.	sp.	sp.
	Gram character	-	+	+	-
Morphology	Shape	Rod	Cocci	Rod	Cocobacilli
	Motility	Motile	Non Motile	Motile	Non Motile
	Indole	-	-	-	-
	MR	-	-	-	-
	VP	-		-	-
Biochemical	Citrate utilization	-	-	+	+
tests	Nitrate reduction	-	-	+	-
	H2S production	-	-	-	-
	Catalase	+	-	+	+
	Oxidase	-	-	+	-
	Urease	+	+	-	-
Sugar	Lactose	-	+	+	-
fermentation	Mannitol	-	-	-	-
	Glucose	-	+	+	+

Table 1: Morphological and biochemical characteristics of isolates.

Conclusion:

The dye decolorizing abilities have been studied over a three days period where we found that 72 hr was most suitable to achieve maximum degradation. The most efficient species were *Pseudomonas* and *Staphylococcus* with 72 % and 66 % dye degradation at optimum temperature of 30° C and pH 7 respectively in three days. On the other hand enriched bacterial consortium could efficiently degrade the dye up to 88 % at optimum temperature of 30° C and pH 7 in three days. As compared to individual bacteria, consortium showed significant hike (16 %) in dye degradation.

References:

[1]VanderZee,F.P.,Bisschops,I.A.E.,Blanchard,V.G.m,Bouwman,R.H.M.,Lettinga,G.andField, J.A, The contribution of biotic and abiotic processes during azo dye reduction in anaerobic sludge, (2003), Water R.37, 3098-3109.

[2]Subhathra M, Prabakaran V, Kuberan T and Balamurugan I. Biodegradation of azo dye from textile effluent by Lysinibacillus sphaericus. Sky Journal of Soil Science and Environmental Management 2013; 2(1): 1-11.

[3] Jin, X.C. Liu, G.Q. Xu, Z.H. Tao, W.Y. Decolorization of a dye industry effluent by *Aspergillusfumigatus* XC6. *Appl Microbiol Biotechnol.*,74:239-43 (2007).

[4] Lucious, S. Reddy, E.S. Anuradha, V. Yogananth, N. Ali, M.Y.S. Vijaya, P. Rajan, R. Parveen, P.M.K. Decolorization of Acid Dyes by *B.cereus* and *P.aeruginosa* isolated from Effluent of Dyeing Industry *Int. J. Pure App. Biosci.*, 2 (3):23-29 (2014).

[5] Cervantes, F.J., F.P. Van Der Zee and G. Lettinga, 2001. Enhanced decolourisation of Acid Orange 7 in a continuous UASB reactor with quinones as redox mediators. Water Science and Technology, 44: 123-128.

[6] Kaushik, P. & Malik, A. Effect of nutritional conditions on dye removal from textile effluent by *Aspergillus lentulus*. *World Journal of Microbiology and Biotechnology*, 26(11):1957-1964 (2010).

[7] Tony, Bella & Goyal, Dinesh & Khanna, Sunil. (2009). Decolorization of textile azo dyes by aerobic bacterial consortium.International Biodeterioration & Biodegradation. 63. 462-469. 10.1016/j.ibiod.2009.01.003.

[8] Raval N.P., Shah P.U., Shah N.K. Adsorptive amputation of hazardous azo dye Congo red from wastewater: A critical review. Environ. Sci. Pollut. Res. 2016;23:14810–14853.doi: 10.1007/s11356-016-6970-0. [PubMed] [CrossRef] [Google Scholar]

[9] Sen S.K., Raut S., Bandyopadhyay P., Raut S. Fungal decolouration and degradation of azo dyes: A review. Fungal Biol. Rev. 2016;30:112–133. doi: 10.1016/j.fbr.2016.06.003. [CrossRef] [Google Scholar]

[10] Martinez-Huitle C.A., Brillas E. Decontamination of wastewaters containing synthetic organic dyes by electrochemical methods: A general review. Appl. Catal.B Environ. 2009;87:105–145.doi: 10.1016/j.apcatb.2008.09.017. [CrossRef] [Google Scholar]

[11] Robinson T., Mcmullan G., Marchant R., Nigam P. Remediation of dyes in textile effluent: A critical review on current treatment technologies with a proposed alternative. Bioresour.Technol. 2001;77:247–255.doi: 10.1016/S0960-8524(00)00080-8. [PubMed] [CrossRef] [Google Scholar]

[12] Cerniglia C.E., Freeman J.P., Franklin W., Pack L.D. Metabolism of azo dyes derived from benzidine, 3,3'-dimethylbenzidine and 3,3'-dimethoxybenzidine to potentially carcinogenic aromatic amines by intestinal bacteria. Carcinogenesis. 1982;3:1255–1260. doi: 10.1093/carcin/3.11.1255. [PubMed] [CrossRef] [Google Scholar]

[13] Castro E., Avellaneda A., Marco P. Combination of Advanced Oxidation Processes and Biological Treatment for the
Removal of Benzidine-Derived Dyes. Environ. Prog. Sustain. Energy. 2014;33:873–885.doi:
10.1002/ep.11865. [CrossRef] [Google Scholar]

[14] Bushnell Haas. J Bacteriol 1941;41:653.

[15] Barrow GI, Feltham RKA. Cowan and steel's manual for the identification of medical bacteria. New York, USA: Camb. Univ. Press; 1993.

[16] Bergey DH, Buchanan RE, Gibbons NE. Bergey's manual of determinative bacteriology. 8th ed. Baltimore: The Williams and Wilkins Co.; 1974.

[17] Chen C, Wu JY, Huang CC, Liang YM, Hwang SCJ. J Biotechnol2003;101:241-52.

[18] Rajee O, Patterson J. Indian. J Microbiol 2011;51(2):159–63.



National Conference on Innovative Research in Science and Technology **NCIRST-2019**

17 & 18 December, 2019

SOUVENIR



Organized by **Department of Chemistry**



Shivaji Nagar, Morshi Road, Amravati, (MS) - 444 603

Shri Shivaji Science College

Accredited "A" Grade with CGPA 3.13 Identified as UGC-CPE, DST-FIST, SGBAU-Lead College

















Scanned by CamScanner

S.N	Title of paper	Authors Pag	
37	Modification And Characterization Of Starch As An Adhesive	Pradip V. Tekade', Manoranjan Patnaik', Bhagyashri U. Tale', Nakul Barwat', Sakshi Kawale', and Harshada Chikhalkar'	
38	Antifungal Activity Of Silver Substituted Copper Ferrite Nanopawder Synthesized By Sol-gel Method		6
39	One Step Synthesis Of Aryl-aeridinesusing Aromatic Acid	Pravin R. Kawle", Rahul	37
40	Studies In Physical Parameters Of P-hydroxy, 3 Methoxybenzaldehyde In Binary Liquid Systems	Gaikwad ² and Hemant Dhande R. A. Thakare A. S. Burghate, S. A. Wadhal ²	37
41	Phytochemical Analysis Of Root Of Cassine Glauca Plant.	Rajesh P. Ganorkar *	38
42	Preparation, Characterization Of G-C ₃ N ₄ -TiO ₁ Nanocomposites And Photocatalytic Activity For Degradation Of Dyes	Radhakrishna S. Sutar, Rani P. Barkul, Meghshyam K. Patil	38
43	Shelf Life Studies Of Pesticidal Formulation Of Bifenthrin 10 Ec In Storage	*Rashmi Urkude ¹ , Sonika R. Kochhar ²	39
44	Determination Of Molar Refraction And Polarizabilityconstant Of Substituted Thiazolyl Schiff's Bases In Binary Solvent Mixtures With Varying Concentration.	Dr. Rupali S. Talegaonkar and Dr. A. S. Bhurghate	39
45	Synthesisand Characterization Of Some N- Substitutedformamidino-n'- Phenyliminothiocarbamide Series	P. V. Raut',S. A. Waghmare [*] *, S. S. Padhen ^e , R. D. Isankar ^d and D.T. Tayade ^d	40
46	Synthesis, Spectral And Thermal Studies Of Metal Complexes Of 2,4-dihydroxyacetophenone Salicylo Hydrazone (dhash)		40
47	Structural Properties Of Chemically Spray Deposited Batio ₃ Thin Films	Syed Ghause Ibrahim [®] and Ashish V. Kadu [®]	41
48	Ac Impedance And Dielectric Studies Of CoFe ₂ O ₄ Nanocomposites Prepared By Sol-gel Techniques	S. M. Ghodsal , A. B. Bodado and G. N. Chaudhari	° 41
49	Investigate Visible Quantum Cutting In Keaf,:gd ² Eu ³⁺ Phosphor	 S.R. Jaiswal, P.A. Nagpure, V.B. Bhatkar and S.K. Omanwar 	43
50	"One Pot Synthesis Of Benzothiazaole Derivaties And Their Characterization"	S. S. Ubarhande, P.R. Padole and B. N. Bera	d• 4
51	Synthesis And Characterization Of Schiff Bases Derived From 4-(5-bromothiophene-2-yl)-6- (4-chloro Pheneyl) Pyrimidin-2-amine.	S. V. Manohare [*] and S. S. Thakare ^b	4
52	Synthesis Of Manganese Chloride Doped In Polyaniline And Its Characterization	Sachin Bangade ** And Vivek M. Raut*	

MODIFICATION AND CHARACTERIZATION OF STARCH AS AN ADHESIVE

Pradip V. Tekade', Manoranjan Patnaik^b, Bhagyashri U. Tale', Nakul Barwat', Sakshi Kawale', and Harshada Chikhalkar'

*Bajaj College of Science (Previously known as Jankidevi Bajaj College of Science), Jamanalal Bajaj Marg, Civil lines, Wardha, Maharashtra, India
*Mahatma Gandhi Institute for Rural Industrialization (MGIRI), Wardha, Maharashtra, India Email: bhagyashritale@gmail.com, pradiptekade@gmail.com

Increasing demand of energy globally and scarcity of petroleum resources has shifted focus of chemical industries to look for alternative raw material resources. Bio-based adhesive are attracting more and more attention in various fields due to their improved environmental footprint and independence from petroleum resources. Though synthetic chemical based resins have better bonding properties, it shows some drawbacks, which are harmful to humans as well as environment. Starch is produced by plants as a way to store the chemical energy that they produce during photosynthesis. The main goal of this study was to use cornstarch and arrowroot starch in the production of environmentally sound adhesives. Arrowroot & corn starch are environmentally friendly products, they can be modified as adhesive. Starch is an ideal material for manufacturing of wood-composite adhesives due to low cost, high free hydroxyl content, easy processing and treatment.

MODIFICATION AND CHARACTERIZATION OF STARCH AS AN ADHESIVE

¹Pradip V.Tekade, ²Bhagyashri U.Tale, ³Nakul Barwat, ⁴Sakshi Kawale, ⁵Harshada Chikhalkar, ⁶Manoranjan Patnaik

¹Associate Professor, ²,³Assistant Professor, ^{4&5}P.G.Students, ⁶Dy. Director

¹⁻⁵ Department of Chemistry, Bajaj College of Science (Previously known as Jankidevi Bajaj College of Science), Jamanalal Bajaj Marg,

Civil lines, Wardha, Maharashtra, India

⁶ Chemical based industries division, Mahatma Gandhi Institute for Rural Industrialization (MGIRI) , Wardha, Maharashtra, India.

Abstract: Increasing demand of energy globally and scarcity of petroleum resources has shifted focus of chemical industries to look for alternative raw material resources. Bio-based adhesive are attracting more and more attention in various fields due to their improved environmental footprint and independence from petroleum resources. Though synthetic chemical based resins have better bonding properties, it shows some drawbacks, which are harmful to humans as well as environment. Starch is produced by plants as a way to store the chemical energy that they produce during photosynthesis. The main goal of this study was to use cornstarch and arrowroot starch in the production of environmentally sound adhesives. Arrowroot & corn starch are environmentally friendly products, they can be modified as adhesive. Starch is considered as a ideal material for manufacturing of wood-composite adhesives because of its low cost, high free hydroxyl content, easy processing and treatment.

I. INTRODUCTION AND LITERATURE SURVEY:

Adhesives play a fundamental role in many modern technologies and adhesive failure have catastrophic consequence. It is necessary to understand the factors responsible for the production of a good durable adhesive bond. Starch is produced from wheat, corn, sweet potato, rice etc. But the corn starch is having a good adhesive and film forming properties. Since starch is renewable, cheap, non-toxic, easily available, biodegradable and hence it has more demand in adhesive industry.

Pure starch powder has white colour with no taste and odour which is insoluble in cold water or alcohol. It is a polymeric carbohydrate, consists a large number of glucose units joined by glycosidic bonds. It consists of two types of molecules; the linear and helical amylose or the branched amylopectin. Depending on a source, starch contains 20 to 25% amylose and 75 to 80% amylopectin [1].

Starch derivatives are used in various industries as thickeners, gelling agents and encapsulating agents for papermaking, as wet-end additives for dry strength, surface sizes and binders, as adhesives (bag, bottle labeling, laminating, envelopes etc.), for warp sizing in textiles, and for glass fiber sizing [2]. Arrowroot becomes thick at a lower temperature than flour & not affected by acidic ingredients as well as freezing [3].

The most commonly available industrial starches are waxy cornstarch, regular corn starch, high-amylose corn starch type V, and high-amylose corn starch type VII, with amylose concentrations of 0, 28, 55, and 70% respectively [4].

Adhesives are substances that are able to make things adhere or stick together without deformation or failure through a process called adhesion. Renewable and biodegradable starch adhesives are topic of interest for research because of its environmental freindliness. Adhesives prepared from starch are most extensively used in corrugated board industry because of its abundant supply, low cost, renewability, biodegradability, and ease of chemical modifications [5].

Properties of good adhesives:

An adhesive is considered to be good if it is able to give complete bonding with good drying rate in presence of adequate heating [6-15]. To realize this, an adhesive needs to have following properties :

- 1. Appropriate viscosity
- 2. High initial tack
- 3. Solids in range of 20-33%
- 4. Consistency in batch glue properties
- 5. Fast setting

Local Organizing Committee

Dr.A.D. Dahegaonkar Dr. S. S. Aswale Dr. R.S. Hajare Dr. G. N. Satpute Dr. P.N. Nasare Dr. N. Gandhare Dr. S.R. Silre Dr. P. M. Shende Dr. K.D. Shahare Dr. S. M. Tekade Prof. D.L. Bhaware Dr. M.P. Dhore Prof. K.V. Bhongale Dr. P.K. Bhute Prof. S.H. Shrirame Dr. Chitra Dhawale Dr.S.D.Deo Dr. S. D. Khamitkar Dr.D.N.Warade Dr. G.D. Kurundkar Dr.P.H. Munjankar Dr. S.B. Kishore Dr.R.B. Sisociva Dr. K.D. Kalaskar Dr.T.S. Morey Dr.A.A. Dhamani Dr.B.S. Gedam Dr. S.S. Bakare Dr.H.S. Tomar Dr. S. Subhash Dr. M.S. Warbha Dr. D. P. Sorwane Dr.L.H. Khalsa Dr. R.R. Kulkami Dr.I.A. Khan, Dr. P. M. Telkhade Dr.W.K. Dange Dr.A.M. Chike Dr.C.M. Bhongale. Dr.A.P.Sawane Dr.S.D. Tade Dr. J. N. Papadkar Dr. Shitai Gomkar Dr. G.D. Deshmukh Dr.U.S. Thool Dr. R.R. Kamdi Dr.A.R. Viruskar Dr. R.B. Dahare Dr.K.G. Rewatkar Dr.L.H. Rohankar Dr. S. B. Kondawar Dr.A.W. Chauhan Dr. B. M. Suryawanshi Dr.A.S. Deshpande Dr. R.A. Mandal Dr.R.A. Gulhane Dr. P.R. Moharkar Dr. LA, Raja Dr. V.D. Bhandakkar Dr. R.V. Tilare Dr. D. M. Pimpaishende Dr.A.D. Tiple Dr. R. M. Thombare Dr. G.T. Kedar Prof. V.S. Dhabarde Prof. Amit Ukey Dr.D.D. Barsagade Dr.A.A. Mistry Prof. L.K. Bedre Dr. C.J. Khune Dr. B. M. Bahirwar Dr. G.A. Wagh Dr.S.R. Gawali Dr. R.P. Mali Dr. R. R. Khirani Dr. B.V. Jadhao Dr. U.P. Manik Dr.K.R. Rao Prof. R.S. Meshram Dr.C.J. Hiware Dr. R.J. Chauhan Dr.V.K. Batra Dr. T.D. Kose Dr. H.S. Jagtap Dr.S.S.Shah Dr.A.J. Khandage Dr.P.N. Jogi Dr. Rajesh Dahegaonkar Dr. S. S. Katkamwar Dr. C.A. Nikhade, Dr.S.V. Madhamshettiwar Dr. Anli Korpenwar Dr. Y.P. Thawari Dr. Shankar Kurkeja Dr. S. S. Sontakke Dr. Mrunal Kale Dr. M.B. Dewase Mr. S. P. Pandav Dr. R. A. Patale Dr. M. B. Wadekar Dr. S. S. Hunge Dr. M. B. Shende

Dr. Wasanti Rewatkar Dr. Sanjay Dudhe Dr. Ajay Agashe Dr. Anil Bhoyar Dr. Saniay Sable Dr. Sudhir Astunkar Dr. C. N. Hanwante Dr. Nirai Khobragade Dr. Anil Chahande Dr. Ashok Mathankar Prof. Vijay Malekar Dr. S.S. Bhutamwar Prof. Prashant Chahare Dr. P.V. Balsaraf Dr. D.Z. Raut Prof. Manohar Bandre Dr. Mangesh Waghade Dr. N.C. Das Dr. S. Deshmuikh Prof. R. Dange Prof. Akkewar Prof. P.B. Ghode Dr. R.P. Dhankar Dr. Lanjewar Dr. Kapoor Dr. V.B. Sakhare Dr. P.B. Dhumane Prof. K.C. Khobragade Prof. S.D. Tummawar Dr.V.P. Dakhane Prof. T.P. Bisen Prof. Ramesh Halame Dr. Sved Abrar A. Dr. Rajkumar Khapekar Dr. N.V. Umate Dr. J.W. Hazare Prof. K.B. Bhute Dr. Vinod Kumar Dr. Pokle Dr. G.R. Avchar Dr. V.G. Mete Prof. B.S. Gedam Dr. Kashyap Prof. Kage Dr. Chepte Prof. Rajurkar Dr. S.S. Zade Dr. Charjan Dr. Thikhe Dr. V. S. Wadhai Dr. U. M. Indurkar

Chief Patrons

- 1) Hon'ble Sau, Nilimatai N. Shinde, President, BSSB, Bhadrawati
- 2) Hon'ble Shri, Nilkanthrao Y. Shinde,
- Ex. MLA& Founder Secretary, BSSB, Bhadrawati
- Hon'ble Prof. T. M. Karde, Former Vice Chancellor, Indiana University, 3) Raipur Chattisgarh, Ex. Professor and Head, PGTD of Mathematics. RTM Nagpur University, Nagpur

Patrons

- 1) Hon'ble Dr. Namdeo V. Kalyankar,
- Vice Chancellor, Gondwana University, Gadchiroli
- 2) Hon'ble Dr. Chandrashekhar V. Bhusari, Pro Vice Chancellor, Gondwana University, Gadchiroli
- 3) Hon'ble Dr. Vivek N. Shinde, Joint Secretary, BSSB, Bhadrawati and Member, Management Council, Gondwana University, Gadchiroli Promoters
- 1) Hon'ble Dr. Vaibhay N. Shinde, Member, BSSB, Bhadrawati
- 2) Hon'ble Dr. Kartik N. Shinde, Member, BSSB, Bhadrawati
- Hon'ble Dr. Vishal N. Shinde, Member, BSSB, Bhadrawati 31

Co-Promoters

- 1) Hon'ble Dr. Ishwar S. Mohurley,
- Registrar, Gondwana University, Gadchiroli.
- 2) Hon'ble Dr. Anil Z. Chitade, Director, Board of Examination and Evaluation, Gondwana University, Gadchiroli,
- 3) Hon'ble Dr. Suresh B. Rewatkar, Dean, Faculty of Science and Technology, Gondwana University, Gadchiroli.

Chairman

Dr. L. S. Ladke

Principal, Nikanthrao Shinde Science and Arts College, Bhadrawati Member - Senate, Gondwana University, Gadchiroli Member - BOE and E. Gondwana University, Gadchiroli Member - Academic Council, Gondwana University, Gadchiroli Chairman - BOS in Mathematics, Gondwana University, Gadchiroli Mobile No. - 9421721895

Convenor

Dr. Aparna B. Dhote Head, Department of Chemistry Mobile No. : 9860139400

Organizing Secretary

Dr. Narendra V. Harney Head, Department of Zoology, Chairman, BOS in Zoology, Gondwana University, Gadchiroli

Co-organizing Secretary

15 1 1 1

Dr. Gajendra R. Bedare Mobile No. 9921975747

Dr. Mohiuddin N. Quadri Mobile No. 9860178791

Mobile No. 9850351565

Dr. Nathu S. Wadhave Mobile No. 9975024641

Prof. Sandeep S. Pradhan Mobile No. 7721954200

National Conference on "RECENT TRENDS IN MATHEMATICAL PHYSICAL, CHEMICAL, LIBRARY AND LIFE SCIENCES"

NCRTMPCLS-2020





Organized by

Department of Mathematics, Physics, Chemistry, Botany, Zoology, Computer Science, Library & IQAC Nilkanthrao Shinde Science and Arts College Bhadrawati - 442902 District - Chandrapur (MS), India.

> Accrediated 'B+' Grade(Third Cycle-2016) with CGPA 2.58 by NAAC, Bangalore.

Institution of Higher Learning, Research and Specialized Studies in the subject Mathematics, Physics, Chemistry, Botany and Zoology. Email - principalnscollege@gmail.com Website - www.nscollege.ac.in | Tele - 07175-265538(O)

National Conference on Recent Trends in Mathematical, Physical, Chemical, Library, Life Sciences - 2020

Orgnized by

Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

EDITORIAL BOARD

Chief Editor

Dr. L.S. Ladke,

Principal, Department of Mathematics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Associate editor in Mathematics & Statistics

Dr. L.S. Ladke

Principal and Head, Department of Mathematics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Dr. S.D. Deo

Associate Professor, PGT Department of Mathematics, Gondwana University, Gadchiroli

Associate editor in Chemistry

Dr. A. B. Dhote

Head, Department of Chemistry, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Dr. S.S. Aswale

Head, Department of Chemistry, L.T. Mahavidyalaya, Wani, District- Yavatmal

Associate editor in Physics

Dr. G. R. Bedare

Department of Physics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Dr. K. N. Shinde

Department of Physics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Associate editor in Botany

Dr. N.S. Wadhave

Head, Department of Botany, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Dr. P.N. Nasare

Department of Botany, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Associate editor in Zoology

Dr. N.V. Harney

Head, Department of Zoology, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Dr. S.R. Sitre

Department of Zoology, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Associate editor in Library and Information Science

Prof. S. S. Pradhan

Head, Department of Library and Information Science, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Article Section

Editorial Board

chemistry

Physics

Zoology

Botany

Library and Information Science

Computer Science

Mathematical Science

Environmental Science

IRJSE IMPACT FACTOR

SJIF Impact Factor 2016: 6.686 2015: 5.642 2014: 4.148 2013: 4.115 ICV 2016 = 55.77

OUR OTHER JOURNAL



International Journal of Life Sciences ISSN:2320-7817 (print) 2320-964X (Online)

National Conference on Recent Trends in Mathematical, Physical, Chemical, Library, Life Sciences - 2020

Special Issue A7 February-2020

First published: 07 February 2020

CHEMICAL SCIENCES

1. Synthesis and Electrical Conductivity of Mn(III), Fe(III), VO(IV), Zr(IV) and UO2(IV) Complexes derived from thiazole Schiff base.

Kelode SR and Jagnit PR

POF

2. Electrical Conductivity of Co (II), Ni(II) and Cu(II), Complexes derived from bidentate thiazole Schiff base.

Kelode SR and Jagnit PR

PDF

3. Design, synthesis, and biological screening of some novel Quinoline-azetidinone derivative: an innovative approach towards the medicinal sciences.

Bodkhe Yogita G and Chowdhary Anupama R

POF

4. Significance of Hydration Process of Sugars and Pseudo-sugar in Sweet Taste Chemoreception at 298 K.

Shah Shaukat Ajim, Ratnakar Lanjewar and Mamta Lanjewar

POF

5. Comparative study of acoustic and thermodynamic property of aqueous solution of cefotaxime sodium and ampicillin sodium by using ultrasonic interferometer.

Hajare RS, Aswale SS and Aswale SR

POF

6. Scatchard analysis of analgin with BSA at various pH and molecular modeling study.

Pisudde Ajay, Tekade Pradip and Thakare Shrikant

POF

7. Assessment of water quality for drinking purposes using correlation study and water quality index.

Umare Vikas D



8. Synthesis, characterisation and antimicrobial activity of some pyrimidine derivatives from chalcones.

Rahatikar GB, Lakhekar SN, Baseer MA

POF

9. Study of Hydration number in binary liquid mixtures.

Ganjare Pravin J, Aswale Sunanda S, Aswale Shashikant R

PDF

10. Solvent Extraction and Spectrophotometric Determination of zinc (II) with N,N'-Bis (salicylaldehyde) ethylenediamine [salen].

Parkhi SM and Nannaware AM<



h1>

Editorial Board

chemistry

Physics

Zoology

Botany

Library and Information Science

Computer Science

Mathematical Science

Environmental Science

IRJSE IMPACT FACTOR

SJIF Impact Factor 2016: 6.686 2015: 5.642 2014: 4.148 2013: 4.115 ICV 2016 = 55.77

OUR OTHER JOURNAL



International Journal of Life Sciences ISSN:2320-7817 (print) 2320-964X (Online)

Scatchard analysis of analgin with BSA at various pH and molecular modeling study.

Pisudde Ajay, Tekade Pradip* and Thakare Shrikant

Department of Chemistry, Jankidevi Bajaj College of Science, Jamnalal Bajaj marg, civil lines, Wardha (India). Email: <u>pradiptekade@gmail.com</u>

Manuscript Details

Available online on <u>http://www.irjse.in</u> ISSN: 2322-0015

Cite this article as:

Pisudde Ajay, Tekade Pradip and Thakare Shrikant. Scatchard analysis of analgin with BSA at various pH and molecular modeling study, *Int. Res. Journal of Science & Engineering,* February, 2020, Special Issue A7 : 26-31.

© The Author(s). 2020 Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

ABSTRACT

Study of the binding of drug with plasma protein by the acoustical properties shows simple and effective method. Analgin is a pain reliever and antipyretic drug. We studied binding of analgin with plasma protein by ultrasonic, FT-IR and molecular modeling techniques. In the present study, we used ultrasonic method for the study of the binding of analgin with BSA which is the novel method for study of binding of analgin with BSA. Study of interaction of analgin with BSA shows successful binding with BSA. Binding of BSA with analgin further confirmed using FT-IR spectroscopy and molecular modeling study. Effect of pH on the binding of analgin with BSA was also studied. The values of the association constant calculated from the Scatchard plot at varying pH 3, 4 and 5 are 0.5012, 0.4994 and 0.5014 respectively. Study of interaction by FT-IR spectroscopy gives the changes in peak positions of amide bands. The amide I changes from 1635 to 1642 cm⁻¹ and amide II 1538 to1556 cm⁻¹. It shows the secondary structure of BSA changes on binding with analgin. binding interaction of analgin with BSA was further confirmed by using molecular modelling study. The energy value obtained (-213.34) shows that the analgin efficiently binds with BSA.

Keywords Ultrasonic study, FT-IR spctroscopy, Scatchard analysis, association constant, molecular modeling study.

National Conference on "RECENT TRENDS IN MATHEMATICAL, PHYSICAL, CHEMICAL, LIBRARY AND LIFE SCIENCES" Certificate This is to certify that Brot. / Dr. / Mist Mes. Nita P. Mohabansi J. 8. Science College, Wardha Dr. has attended National Conference on " Recent Trends in Mathematical, Physical, Chemical Library and Life Sciences' NCRTMPELS - 2020 held on Friday, 7" February, 2020 as Delegate / Research Scholar / P. G. Student, He / She has actively Participated / Delivered Invited Talk / Chaired Technical Session / Presented Paper / Poster entitled" Thermo acoustical studies of molecular interaction of N-[[(25)-1- ethylpyne-- lidin - 2- ye) methyd] - 2 - methony 5in the conference. sulphantaylbenzanamide at different temperature a dke 774/2020 30 ste Dr. Mrs. A. B. Dhote Dr. N. V. Harney Dr. L. S. Ladke Or. Vivek N. Shinde Converior Organizing Secretary Principal jaint Secretary, RSSB WORTMROLS - JORD NCKIMPCLS-2020 Chairman, Organizing Committee Patron HONTMPOLS - NO.KO NCRIMPULS - 1010 Organized by Bhedrawati Shikshan Sanstha, Bhadrawati's CRTMPCLS Nilkanthrao Shinde Science and Arts College 2020 Bhudrawati, District - Chandriepur (M45), India. DGC 2(f) and 12(iii) Status Accredited '8-' Grade(Third Cycle-2016) with CGPA 2.58 by NAAC Bangalore. Institution of Higher Learning, Research and Specialized Studies in the subject. Mathematics, Physics, Chemistry, Botany and Joology, Email - principaliticsllege@gmail.com. Website - aven.nscollege.ec.in / Tele - 07175-265538(0)

Local Organizing Committee

Dr.A.D. Dahegaonkar Dr. S. S. Aswale Dr. R.S. Hajare Dr. G. N. Satpute Dr. P.N. Nasare Dr. N. Gandhare Dr. S.R. Silre Dr. P. M. Shende Dr. K.D. Shahare Dr. S. M. Tekade Prof. D.L. Bhaware Dr. M.P. Dhore Prof. K.V. Bhongale Dr. P.K. Bhute Prof. S.H. Shrirame Dr. Chitra Dhawale Dr.S.D.Deo Dr. S. D. Khamitkar Dr.D.N.Warade Dr. G.D. Kurundkar Dr.P.H. Munjankar Dr. S.B. Kishore Dr.R.B. Sisociva Dr. K.D. Kalaskar Dr.T.S. Morey Dr.A.A. Dhamani Dr.B.S. Gedam Dr. S.S. Bakare Dr.H.S. Tomar Dr. S. Subhash Dr. M.S. Warbha Dr. D. P. Sorwane Dr.L.H. Khalsa Dr. R.R. Kulkami Dr.I.A. Khan, Dr. P. M. Telkhade Dr.W.K. Dange Dr.A.M. Chike Dr.C.M. Bhongale. Dr.A.P.Sawane Dr.S.D. Tade Dr. J. N. Papadkar Dr. Shitai Gomkar Dr. G.D. Deshmukh Dr.U.S. Thool Dr. R.R. Kamdi Dr.A.R. Virußar Dr. R.B. Dahare Dr.K.G. Rewatkar Dr.L.H. Rohankar Dr. S. B. Kondawar Dr.A.W. Chauhan Dr. B. M. Suryawanshi Dr.A.S. Deshpande Dr. R.A. Mandal Dr.R.A. Gulhane Dr. P.R. Moharkar Dr. LA, Raja Dr. V.D. Bhandakkar Dr. R.V. Tilare Dr. D. M. Pimpaishende Dr.A.D. Tiple Dr. R. M. Thombare Dr. G.T. Kedar Prof. V.S. Dhabarde Prof. Amit Ukey Dr.D.D. Barsagade Dr.A.A. Mistry Prof. L.K. Bedre Dr. C.J. Khune Dr. B. M. Bahirwar Dr. G.A. Wagh Dr.S.R. Gawali Dr. R.P. Mali Dr. R. R. Khirani Dr. B.V. Jadhao Dr. U.P. Manik Dr.K.R. Rao Prof. R.S. Meshram Dr.C.J. Hiware Dr. R.J. Chauhan Dr.V.K. Batra Dr. T.D. Kose Dr. H.S. Jagtap Dr.S.S.Shah Dr.A.J. Khandage Dr.P.N. Jogi Dr. Rajesh Dahegaonkar Dr. S. S. Katkamwar Dr. C.A. Nikhade, Dr.S.V. Madhamshettiwar Dr. Anli Korpenwar Dr. Y.P. Thawari Dr. Shankar Kurkeja Dr. S. S. Sontakke Dr. Mrunal Kale Dr. M.B. Dewase Mr. S. P. Pandav Dr. R. A. Patale Dr. M. B. Wadekar Dr. S. S. Hunge Dr. M. B. Shende

Dr. Wasanti Rewatkar Dr. Sanjay Dudhe Dr. Ajay Agashe Dr. Anil Bhoyar Dr. Saniay Sable Dr. Sudhir Astunkar Dr. C. N. Hanwante Dr. Nirai Khobragade Dr. Anil Chahande Dr. Ashok Mathankar Prof. Vijay Malekar Dr. S.S. Bhutamwar Prof. Prashant Chahare Dr. P.V. Balsaraf Dr. D.Z. Raut Prof. Manohar Bandre Dr. Mangesh Waghade Dr. N.C. Das Dr. S. Deshmuikh Prof. R. Dange Prof. Akkewar Prof. P.B. Ghode Dr. R.P. Dhankar Dr. Lanjewar Dr. Kapoor Dr. V.B. Sakhare Dr. P.B. Dhumane Prof. K.C. Khobragade Prof. S.D. Tummawar Dr.V.P. Dakhane Prof. T.P. Bisen Prof. Ramesh Halame Dr. Sved Abrar A. Dr. Rajkumar Khapekar Dr. N.V. Umate Dr. J.W. Hazare Prof. K.B. Bhute Dr. Vinod Kumar Dr. Pokle Dr. G.R. Avchar Dr. V.G. Mete Prof. B.S. Gedam Dr. Kashyap Prof. Kage Dr. Chepte Prof. Rajurkar Dr. S.S. Zade Dr. Charjan Dr. Thikhe Dr. V. S. Wadhai Dr. U. M. Indurkar

Chief Patrons

- 1) Hon'ble Sau, Nilimatai N. Shinde, President, BSSB, Bhadrawati
- 2) Hon'ble Shri, Nilkanthrao Y. Shinde,
- Ex. MLA& Founder Secretary, BSSB, Bhadrawati
- Hon'ble Prof. T. M. Karde, Former Vice Chancellor, Indiana University, 3) Raipur Chattisgarh, Ex. Professor and Head, PGTD of Mathematics. RTM Nagpur University, Nagpur

Patrons

- 1) Hon'ble Dr. Namdeo V. Kalyankar,
- Vice Chancellor, Gondwana University, Gadchiroli
- 2) Hon'ble Dr. Chandrashekhar V. Bhusari, Pro Vice Chancellor, Gondwana University, Gadchiroli
- 3) Hon'ble Dr. Vivek N. Shinde, Joint Secretary, BSSB, Bhadrawati and Member, Management Council, Gondwana University, Gadchiroli Promoters
- 1) Hon'ble Dr. Vaibhay N. Shinde, Member, BSSB, Bhadrawati
- 2) Hon'ble Dr. Kartik N. Shinde, Member, BSSB, Bhadrawati
- Hon'ble Dr. Vishal N. Shinde, Member, BSSB, Bhadrawati 31

Co-Promoters

- 1) Hon'ble Dr. Ishwar S. Mohurley,
- Registrar, Gondwana University, Gadchiroli.
- 2) Hon'ble Dr. Anil Z. Chitade, Director, Board of Examination and Evaluation, Gondwana University, Gadchiroli,
- 3) Hon'ble Dr. Suresh B. Rewatkar, Dean, Faculty of Science and Technology, Gondwana University, Gadchiroli.

Chairman

Dr. L. S. Ladke

Principal, Nikanthrao Shinde Science and Arts College, Bhadrawati Member - Senate, Gondwana University, Gadchiroli Member - BOE and E. Gondwana University, Gadchiroli Member - Academic Council, Gondwana University, Gadchiroli Chairman - BOS in Mathematics, Gondwana University, Gadchiroli Mobile No. - 9421721895

Convenor

Dr. Aparna B. Dhote Head, Department of Chemistry Mobile No. : 9860139400

Organizing Secretary

Dr. Narendra V. Harney Head, Department of Zoology, Chairman, BOS in Zoology, Gondwana University, Gadchiroli

Co-organizing Secretary

15 1 1 1

Dr. Gajendra R. Bedare Mobile No. 9921975747

Dr. Mohiuddin N. Quadri Mobile No. 9860178791

Mobile No. 9850351565

Dr. Nathu S. Wadhave Mobile No. 9975024641

Prof. Sandeep S. Pradhan Mobile No. 7721954200

National Conference on "RECENT TRENDS IN MATHEMATICAL PHYSICAL, CHEMICAL, LIBRARY AND LIFE SCIENCES"

NCRTMPCLS-2020





Organized by

Department of Mathematics, Physics, Chemistry, Botany, Zoology, Computer Science, Library & IQAC Nilkanthrao Shinde Science and Arts College Bhadrawati - 442902 District - Chandrapur (MS), India.

> Accrediated 'B+' Grade(Third Cycle-2016) with CGPA 2.58 by NAAC, Bangalore.

Institution of Higher Learning, Research and Specialized Studies in the subject Mathematics, Physics, Chemistry, Botany and Zoology. Email - principalnscollege@gmail.com Website - www.nscollege.ac.in | Tele - 07175-265538(O)

National Conference on Recent Trends in Mathematical, Physical, Chemical, Library, Life Sciences - 2020

Orgnized by

Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

EDITORIAL BOARD

Chief Editor

Dr. L.S. Ladke,

Principal, Department of Mathematics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Associate editor in Mathematics & Statistics

Dr. L.S. Ladke

Principal and Head, Department of Mathematics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Dr. S.D. Deo

Associate Professor, PGT Department of Mathematics, Gondwana University, Gadchiroli

Associate editor in Chemistry

Dr. A. B. Dhote

Head, Department of Chemistry, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Dr. S.S. Aswale

Head, Department of Chemistry, L.T. Mahavidyalaya, Wani, District- Yavatmal

Associate editor in Physics

Dr. G. R. Bedare

Department of Physics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Dr. K. N. Shinde

Department of Physics, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Associate editor in Botany

Dr. N.S. Wadhave

Head, Department of Botany, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Dr. P.N. Nasare

Department of Botany, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Associate editor in Zoology

Dr. N.V. Harney

Head, Department of Zoology, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Dr. S.R. Sitre

Department of Zoology, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District-Chandrapur

Associate editor in Library and Information Science

Prof. S. S. Pradhan

Head, Department of Library and Information Science, Nilkanthrao Shinde Science and Arts College, Bhadrawati, District- Chandrapur

Article Section

Editorial Board

chemistry

Physics

Zoology

Botany

Library and Information Science

Computer Science

Mathematical Science

Environmental Science

IRJSE IMPACT FACTOR

SJIF Impact Factor 2016: 6.686 2015: 5.642 2014: 4.148 2013: 4.115 ICV 2016 = 55.77

OUR OTHER JOURNAL



International Journal of Life Sciences ISSN:2320-7817 (print) 2320-964X (Online)

0

06/12/2022, 00:57		IRJSE
	of molecular interaction of N-[[(2 moylbenzamide at different temp	
Mohabansi NP and Satone AK		
PDF		
26. A novel blue-greenish emit	ting phosphor KAl1-xPO4F:Tbx3+	- (0.1 x 1.0).
Akojwar Ashish, Naktode PK, Shin	de KN and Kokode NS	
PDF		
27. To determine physical para	meters of different edible oils	
Bhukya PB, Wakulkar AP and Shah	n SA	
PDF		
28. Effect of pH and doses of H	202 on degradation of p-nitoben	zoic acid
Mandavgane Susmita A		
PDF		
29. Abatement of heavy metal	on the surface of newly prepared	adsorbent.
Hunge Sudhir, Rahangdale Pralhad	l, Lanjewar Mamata and Shewate Var	sha
PDF		
30.Study of electrical conducti salicylate-4,4'diaminodiphenyl	vity of thermally stable terpolyme	er resin phenyl
Dewase MB, Singru RN and Siddiq	-	
PDF		
31.Comparative Studies of Dicl Ultrasonically at 303.15k.	hlofenac Sodium and (Paracetame	ol+DichlofenacSodium)
Dhote Aparna B1, Aswale Sunanda	a S2, Aswale Shashikant R2	
PDF		
32. Study of Acoustic Paramete 298.15K.	ers of Rabeprazole Sodium in Diff	erent Solvents at
Dhote AB		
PDF		
33. Acoustical Studies of Molec	cular Interactions in Riboflavin an	d NaOH at 288 K.
Dange Sudhir and Chimankar Omp	orakash	
POF		
34. Spectrophotometric studies cathonic surfactants	s on the interaction of some triph	enylmethane dyes with
Sarkar RD		
PDF		
IRJSE	Important Links	Fallow Us
ISSN: 2322-0015 editorirjse@gmail.com info@irjse.in Mob No. +91 9420775527	Membership Form Copyright Agreement Author guideline	

Visitor Map



2014-2017 | IRJSE | All Rights Reserved

Design by SevaWeb Studio



Thermo acoustical studies of molecular interaction of N-[[(2S)-1ethylpyrrolidin-2-yl] methyl]-2-methoxy-5-sulfamoylbenzamide at different temperature.

Mohabansi NP* and Satone AK

Department of Chemistry, Jankidevi Bajaj College of Science, Jamnalal Bajaj Marg, Civil Lines, Wardha, India. Email: <u>*nitamohabansi15@gmail.com</u>

Manuscript Details

Available online on <u>http://www.irjse.in</u> ISSN: 2322-0015

Editor: Dr. Arvind Chavhan

Cite this article as:

Mohabansi NP and Satone AK. Thermo acoustical studies of molecular interaction of N-[[(2S)-1ethylpyrrolidin-2-yl] methyl]-2-methoxy-5sulfamoyl benzamide at different temperature., *Int. Res. Journal of Science & Engineering*, February 2020, Special Issue A7: 133-137.

© The Author(s). 2020 Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License

(http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

ABSTRACT

Density(ρ),Viscosity(η) and Ultrasonic Velocity (U) of an alcoholic solution of N-[[(2S)-1- ethylpyrrolidin-2-yl]methyl]-2-methoxy-5-sulfamoylbenzamid drug(NSB) 2.5mM,5.0mM and 10mM were measured at 300, 305 and 310K. The resulting data were used to calculate various acoustical parameters ,acoustic impedance (Z), adiabatic compressibility(β), Intermolecular free length (L_f), Wada's Constant (W), Rao's Constant (R), free volume (V_f) , were calculated which provides valuable information regarding drug-alcohol interaction

Keywords NSB, Acoustical parameters, Inter molecular interaction, drug-alcohol interaction.

INTRODUCTION

Ultrasound refers to such high frequency sound waves that they can't be heard. Now a day's Ultra-Sonic technology due to non-destructive nature [1–3], is used in a variety of applications in medicine, biology, industry, materials science agriculture, oceanography, sonochemistry etc. Ultra-sound waves have been used extensively as chemical additives for order to improve the production yield of produced foods, and also useful in the preparation of biomaterials, protein microspheres, polymer and polymer surface modifications, etc. for material chemistry [4-7].

BENEFICIAL MICROBES IN AGRO-ECOLOGY Bacteria and Fungi

Edited by N. Amaresan, M. Senthil Kumar, K. Annapurna, Krishna Kumar, and A. Sankaranarayanan



BENEFICIAL MICROBES IN AGRO-ECOLOGY BACTERIA AND FUNGI

Edited by

N. AMARESAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, Gujarat, India

> M. SENTHIL KUMAR ICAR-Indian Institute of Pulses Research, Kanpur, Uttar Pradesh, India

K. ANNAPURNA Division of Microbiology, ICAR-Indian Agricultural Research Institute, New Delhi, India

KRISHNA KUMAR Pandit Deendayal Upadhyay College of Horticulture & Forestry, Dr. Rajendra Prasad Central Agricultural University, Tirhut College Campus, Dholi, Muzaffarpur, Bihar, India

A. SANKARANARAYANAN C.G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Bardoli, Surat, Gujarat, India





ACADEMIC PRESS

An imprint of Elsevier

СНАРТЕК

19

Azotobacter

Satish V. Patil^{1, 2}, Bhavana V. Mohite³, Chandrashekhar D. Patil⁴, Sunil H. Koli¹, Hemant P. Borase⁵, Vikas S. Patil⁶

 ¹School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India; ²North Maharashtra Microbial Culture Collection Centre
 (NMCC), Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India; ³Department of Microbiology, Bajaj College of Science, Wardha, Maharashtra, India; ⁴PSL Université Paris: EPHE-UPVD-CNRS, USR 3278 CRIOBE, Université de Perpignan, Perpignan Cedex, France; ⁵C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India; ⁶University Institute of Chemical Technology, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India

1. Introduction

Every living organism essentially requires a utilizable source of nitrogen to survive and grow. Utilizable sources of nitrogen exemplified as nitrogen gas and urea. However, the nitrate, nitrites, and ammonia are the most preferable sources by living organisms. Majorly nitrogen is present on earth in gaseous form and (more than 1017 metric tons) out of which about 2% is in free form in the atmosphere. Although such a large quantity of nitrogen is present on the planet, unfortunately, it is not utilized directly by most living organisms.

The abundance of nitrogen in the atmosphere can become usable for living organisms when they are converted into its usable form through the process of biological fixation of nitrogen. This practice is taking place via several routes. The prokaryotes are the major members intricate in the biological way of nitrogen fixation. Prokaryotes fix the nitrogen from the atmosphere by reducing molecular nitrogen into ammonia, which further used for assimilation of amino acids. This process was assumed to provide 200 million tons of nitrogen (N₂) per year (Rascio and Rocca, 2008). The biological nitrogen fixation process is categorized into two types, symbiotic and nonsymbiotic fixation. Nonsymbiotic nitrogen fixation process involves a major genus *Azotobacter, Azomonas, Beijerinckia*, and *Derxia*.

19. Azotobacter

1.1 Azotobacter taxonomy

The perception of family Azotobacteraceae is defined in the Bergey's manual eighth edition as Gram-negative, aerobic heterotrophic bacteria capable of nonsymbiotic nitrogen fixation under normal atmospheric partial oxygen of pressure (Becking, 1974). Based on the numerical taxonomy, Thompson and Skerman (1979) concluded that the family Azotobacteraceae is comprised of "the Gram negative bacterial genera that are non-spore former, free nitrogen fixer and not found similarity with genera of other families."

The foremost distinctive genus *Azotobacter chroococcum* was revealed and described by Beijerinck Martinus in 1901. He designated and labeled the species *Azotobacter chroococcum* as the foremost free-living aerobic nitrogen fixer (Beijerinck, 1901a).

Azotobacter vinelandii was described in 1903 by Lipman and a year after it was named as Azotobacter beijerinckii (Lipman, 1904), in the honor of Beijerinck. Azotobacter nigricans was identified in 1949, by the Russian microbiologist, Nikolai Krasilnikov (Krassilnikov, 1949). Thompson and Skerman (1981) divided it into two subspecies: Azotobacter nigricans subsp. achromogenes and Azotobacter nigricans subsp. nigricans. In 1981, Thompson and Skerman proposed Azotobacter armeniacus (Thompson and Skerman, 1981). The air-tolerant and microaerophilic type Azotobacter salinestris was reported by Page and Shivprasad that was dependent on sodium ions (Page and Shivprasad, 1991).

Azotobacter has its place to the Bacteria kingdom; Proteobacteria phylum; Gamma proteobacteria class; Pseudomonadales order; Azotobacteraceae family and the *Azotobacter* genus. The family Azotobacteraceae covered free nitrogen fixer usually present in soil, water, and sediments (Aquilanti et al., 2004).

Azotobacteraceae is the dominant member of Rhizobacterium family which has its major role in dinitrogen fixation. Nitrogen is the important factor for soil fertility. *Azotobacter* is Gram negative nonendospore forming bacteria; some of the *Azotobacter* species cause the dormant structures known as a cyst, which majorly includes genera of *Azomonas*, *Beijerinckia*, *Derxia*, and *Azotobacter*. The *Azotobacter* shows either motile or nonmotile nature and possesses catalase and oxidase positive nature.

In 1901, Beijerinck, who, studying the Chemolithotrophy, was attracted by nitrogen fixer *Azotobacter* and extensively studied the melanin producing coccus, *Azotobacter chroococcum* and the extensive capsule producing nitrogen fixer *Azotobacter agilis*, which was found in Holland soil.

The Beijerinck isolated the *Azotobacter chroococcum*, the first species of *Azotobacter* from the Holland soil (Beijerinck, 1901b). Thereafter in the succeeding period, numerous other types of *Azotobacter* collection have been isolated from rhizosphere and soil which were categorized under the family Azotobacteraceae, e.g., Lipman (1903a,b, 1904), reported *Azotobacter vinelandii*, *Azotobacter beijerinckii*, respectively, while Krassilnikov (1949) and Dobereiner (1966), isolated and characterized unique *Azotobacter* having specific association with wild grassroots *Paspalum notatum*, i.e., *Azotobacter paspali*. Thompson and Skerman in 1981, reported *Azotobacter armeniacus* and in 1991 *Azotobacter salinestris* by William Page and Shivprasad (1991).

Azotobacter is Gram-negative, blunt to oval short rods with $1.5-2 \mu m$ or more in diameter, having soil as common habitat, besides that aquatic plant rhizospheric, and phylospheric were also the identified habitat. They are generally aerobic and capable to fix the atmospheric nitrogen in presence of suitable carbon sources.

398

1. Introduction

The Azotobacteraceae family was characterized into two main genera *Azotobacter* and Azomonas based on various characters. The genus *Azotobacter* includes six major species, characterized by their atmospheric nitrogen fixation capacity. In the environment or nitrogen deprived medium *Azotobacter* is aerobic in nature but also able to grow under low oxygen tension. Major six species of the genus are identified and studied extensively are

- **1.** Azotobacter chroococcum
- 2. Azotobacter vinelandii
- 3. Azotobacter beijerinckii
- **4.** Azotobacter nigricans
- **5.** Azotobacter armeniacus
- 6. Azotobacter paspali

Besides all above characterized species, Page and Shivprasad (1991) isolated a cyst forming, nitrogen fixer from saline soil surface in Alberta, Canada. The organism showed specific additional characters, i.e., the presence of Na⁺/succinic acid efflux, and it was named as *Azotobacter salinetris*. The organism showed brown-black pigmented colonies on a Burk medium, the brown-black pigmentation is owing to the water-soluble catechol, melanin production. The organism utilizes galactose, mannitol, glucose, fructose, and melibiose as carbon source without producing acid. The organism uses sodium ion as an electron acceptor.

Out of all species, *Azotobacter chroococcum* is very commonly occurring in soil. The genus *Azotobacter* differentiated from Azomonas based on the microcyst formation. *Azotobacter* form microcyst, a dormant spore-like structure, while Azomonas never formed the microcyst. Besides that, *Azotobacter* always has high GC content, i.e., 63–67.5 mol% than the Azomonas have 52–59 mol%.

The typical genus was allocated to the family Azotobacteraceae (Pribram, 1933), but after the 16S rRNA sequencing study, they were shifted to the family Pseudomonadaceae. The phylogenetic study in 2004 discovered that *A. vinelandii* fits the equal clade as *Pseudomonas aeruginosa* (Rediers et al., 2004) bring about with the concept that the genera *Azotobacter*, *Pseudomonas* and *Azomonas* are correlated and might be alternative (Young and Park, 2007).

The taxonomy controversy resulted in usage of immunological attractions present between the several species of Azotobacteraceae family via Immunoelectrophoresis technique. The *Azotobacter chroococcum* strain was immunologically heterogenous than the *Azotobacter vinelandii* and *Azotobacter paspali* strains (Tchan et al., 1983).

Azotobacter has characterized by utilizing a numerical taxonomy into two different phenotypic groups at a similarity level of 79%–85%. One cluster had a colony of white-beige and the other cluster with colonies of yellow-brown (Zulaika et al., 2014).

1.2 Distribution of Azotobacter

They exist in dry deserts, hot steppes, rocky terrain, in dry sands, on foothill summits and in valleys. They are also found in the cold and in an aquatic habitat like the Arctic and Antarctic soils (Garg et al., 2001), thus they present in the soil of diverse topographical areas. The *Azotobacter* population in soil is mostly affected by other soil microbiota. Bagyaraj and Patil (1975) reported the dominant presence of *Azotobacter* in the numerous agronomy crops

rhizosphere for instance sorghum, soybean, ragi, sugarcane, rice, green Gram, and cereals. The *Azotobacter* number was most abundant in black soil compared with red soil and observed reduced in number with an increase in depth while the reduction was additional in black soils (Ramaswamy et al., 1977).

1.3 Habitat and reproduction

The *Azotobacter* can flourish in a nonacidic soil, as well as in the cold climate like the Arctic and Antarctic soil. The cyst forming potential is responsible for resistance to severe environments like drought, solar radiations, ultrasound, UV, and gamma radiation. But the extreme heat is not tolerated by *Azotobacter*.

Azotobacter reproduces by simple division (fission), like all bacteria, and respires aerobically with the generation of energy.

When they form cysts, the cysts cannot reproduce, they survive the adverse conditions, and then when the optimal conditions are achieved again, the cysts germinate and form vegetative cells, which then reproduce via simple cell division.

1.4 Molecular characterization

The *Azotobacter* genus is suitable for the Proteobacteria subclass (Becking, 2006; Tchan, 1984) and includes seven species: *A. chroococcum, A. vinelandii, A. beijerinckii, A. nigricans, A. armeniacus,* and *A. salinestris. A. paspali,* a novel endemic species, was screened from Thailand based on molecular techniques in 2005 (unpublished data). *Azotobacter* is the aerobic-free nitrogen fixer with the heterotrophic mode of nutrition having 63%-67.5% G + C content (Becking, 2006; Setubal et al., 2009), and found presence in the soil, aquatic environment as well as sediments (Tejera et al., 2005; Torres-Rubio et al., 2000). *Azotobacter* contains the highest amount of DNA than the other bacteria, the circular DNA molecule of *Azotobacter* comprises 5,342,073 nucleotide pairs and 4988 genes out of 5043 encode proteins. The Nif genes are the major complex genes. The genome is typical of utmost prokaryotes. The above average amount of DNA is might be due to larger cells of *Azotobacter* than those of other bacteria (Aquilanti et al., 2004).

The *Azotobacter* has 52–67.5 mol% GC content. The DNA content and the number of chromosomes in the cells increase upon aging. *Azotobacter* contains more than 100 copies of a chromosome per cell in old cultures.

The *Azotobacter chroococcum* NCIMB 8003 (ATCC 4412) (Ac-8003) genome, contains 5,192,291 bp making seven circular replicons. The species of *Azotobacter* contains $1.5 \text{ g} \times 10-13 \text{ g}$ of DNA, which is approximately 40 times more than that of *E. coli*. It also has presence of six plasmids (Phadnis et al., 1988). The *Azotobacter* sp. DNA shares several similar properties to *Escherichia coli* genome, as gene type and recognition factors. Nif gene studies of *Azotobacter*, nitrogenase holoenzyme of *Azotobacter vinelandii* indicate that the enzyme active site holds the molybdenum iron-sulfide cluster cofactors (FeMoCo), each carrying two pseudocubic iron-sulfido structures. The *Azotobacter* chromosome has 66.27% G + C content. The metabolic pathways and macromolecular designs of this organism appear well-preserved with genes meant for CO-dehydrogenase, formate dehydrogenase, and a soluble NiFe-hydrogenase (Robson et al., 2015).

400

1. Introduction

NifH gene is the source of taxonomic identification of *Azotobacter* for analysis of their nitrogen fixing genetic potential (Zehr et al., 1995). These NifH genes are also valuable as the markers for the finding and recognition of the genetic diversity of the *Azotobacter* residing in the soil (Ueda et al., 1995; Widmer et al., 1999).

The *A. vinelandii* DJ genome sequence has a 5,365,318 bp single circular genome. *A. vinelandii* has a complement of respiratory proteins for oxygen-sensitive processes. It produces the alginate that guards the organism against spare exogenous oxygen. In accordance with the oxygen availability, the alginate conformation may change by several repetitions of alginate modification genes. The investigation of the genome recognized the genes coding for the three-known oxygen-sensitive nitrogenases and more for oxygen-sensitive enzymes, such as formate dehydrogenase and carbon monoxide dehydrogenase. In this way, *A. vinelandii* could work as a mediator for the formation and elucidation of oxygen-sensitive proteins (Setubal et al., 2009).

The varying strain of *Azotobacter* like *A. vinelandii* strain CA (or OP) (ATCC 13,705, accession no. CP005094) was a nongummy, pigment fabricating, native strain (Bush and Wilson, 1959). *A. vinelandii* strain CA6 (accession no. CP005095) is a mutant strain resulted from strain CA by spontaneous alterations. It was reported that tungstate stops growth and nitrogen fixation by strain CA nevertheless it could inhibit CA6 (Bishop et al., 1980). Similarly, strain CA6 also diminished molybdate uptake (Premakumar et al., 1996) but produce a vast amount of hydrogen gas during nitrogen fixation.

1.5 Nutritional requirement

The *Azotobacter* sp. has metabolic aptitudes of atmospheric nitrogen fixation by conversion to ammonia. The three discrete nitrogenase enzymes make these bacteria of interest for studying the nitrogen fixation and its role in agriculture. The *Azotobacter* sp. has the highest metabolic rate (Jensen, 1954).

Azotobacter is a mixotrophic bacteria showing autotrophy or heterotrophic mode of nutrition by making its own food (by sunlight or by chemical reaction [autotropic] or by getting food from other sources, respectively). Azotobacter often grows green/brown, slimy, and around 7 mm diameter colonies. The fixation of nitrogen requires source of carbon, which could be fulfilled by carbohydrates and sugars, as per, 1 g of glucose is needed in order to fix 10 g of nitrogen.

1.6 The general characteristic of Azotobacter

The *Azotobacter* has specific physiological and morphological characteristics which primarily differentiate it from the other Gram negative and nitrogen fixers (Table 19.1). *Azotobacter* species occur from a range of soil habitat, i.e., slightly acidic to alkaline soil and some species like *Azotobacter paspali* are associated with plant root. But generally, *Azotobacter* species population occurs abundantly in fertile soil; this is due to the *Azotobacter* requirement for high minerals like phosphates.

The *Azotobacter* species are chemoheterotrophic, nitrogen fixers, and motile, other than *Azotobacter beijerinckii* and *Azotobacter nigricans* by peritrichous and polar flagella. They generally produce diffusible and nondiffusible big colonies on the nitrogen-free

401

19. Azotobacter

402

sr. No.	Azotobacter species	Gram nature	Cell shape	Cell size (L × W) μm	Pigment production	Motility
1.	A. vinelandii	-ve	Round-ended rods	$3.0 - 4.5 \times 1.5 - 2.4$	Yellow-green, fluorescent, water-soluble pigment	+ve
2.	A. beijerinckii	-ve	Rods or ellipsoidal	$3.2 - 5.3 \times 1.7 - 2.7$	Yellowish or cinnamon pigment	-ve
3.	A. chroococcum	-ve	Rod-, oval-ovoid-, or coccus	3.0–7.0 × 1.5–2.3	Brown or blackish-brown	+ve
4.	A. paspali	-ve	Long filaments	$7 - 12 \times 1.3 - 1.7$	Yellow-green, fluorescent or red-violet, water-soluble pigment	+ve
5.	A. armeniacus	-ve	Bluntly rounded rods	5.0-5.7 × 1.7-2.0	Diffusible brown-black or red-violet	+ve
6.	A. nigricans	-ve	Bluntly rounded rods	$4.1 - 4.9 \times 1.5 - 2.7$	Yellow nondiffusible pigment	-ve
7.	A. salinetris	-ve	Rods	$2-4 \times 4.5 - 5.0$	Black brown	+ve

TABLE 19.1 Primary morphological characters on Burks medium.

medium containing sugar or alcohol as carbon sources. The colonies are generally smooth, opaque, somewhat convex glistening, though the nature of colony changes accordingly to medium and type of carbon sources used (Thompson and Skerman, 1979), e.g., the colonies with more big size, more transparent and viscous colonies appeared on media containing sucrose and raffinose than the nitrogen-free media with glucose. The species are also characterized by the production of gray-brown, black nondiffusible pigments. The pigment production is also found to be media component dependent, e.g., *Azotobacter chroococcum* produces nondiffusible brown-gray pigment. The pigments production is also found to be the media component dependent. e.g. *Azotobacter chroococcum* produces non diffusible brown-grey pigment on Stainers medium (Pribram, 1933) fortified with the 0.2% gluconate and black diffusible pigment on benzoate fortified medium. But *Azotobacter nigricans* and *Azotobacter vinelandii* produces brown diffusible pigment in presence of benzoate, whereas *Azotobacter vinelandii* produces brown-black pigment.

Azotobacter is chemoheterotrophic utilizing sugars such as glucose, fructose, ethanol, acetate, carbinol fumarate, pyruvate, and other organic acids as a carbon source. It is also able to use various nitrogen compounds but poorly or unable to use nitrate. Azotobacter does not require organic growth factors but requires only minerals, like vanadium and molybdenum, which is an essential component of the nitrogen fixation system.

The combined nitrogen-free medium with suitable carbon source is the preferable condition for the growth of *Azotobacter*. Although the organism is catalase positive and aerobic in nature the reduce or low oxygen tension condition is required for better nitrogen fixation, because dinitrogen fixation is categorized as a reductive process as well as the involvement of the major oxygen labile enzymes, which is get inactivated in presence of oxygen.

The optimum temperature of growth for most of the members of *Azotobacter* is 28–37°C, but another cardinal temperature varies as per the species, e.g., some species of *Azotobacter* require minimum temperature for growth as 14°C, while *A. beijerinckii* and *A. nigricans* have

2. Isolation of the genus

the minimum temperature requirement is of 9°C and *A. armeniacus* required 28°C as minimum growth temperature. Although optimal temperature meant for most of the *Azotobacter* is 32°C, *A. paspali* and *A. vinelandii* have optimum temperature is 37°C. The optimum temperature also found to vary as per strains, for. e.g., some strains of *A. chroococcum* have optimum temperature is 37°C. The temperature tolerance also found to be varying accordingly strain isolated from subtropical and temperature region. It was repairer that all *Azotobacter* survives at 50°C for to 10 min but not any species able to survive within 10 min at 60°C treatment or incubation. Similarly, *Azotobacter nigricans* and *Azotobacter armeniacus* are unable to grow at 37°C. The growth of *Azotobacter* has observed from pH varies from acidic to alkaline i.e., 4.8-8.5.

1.7 Azotobacter phages

As with the other bacteria, the *Azotobacter* species is also reported to be susceptible to bacteriophages. De Jong (1938) given the first idea for presence of bacteriophages for Azotobacter sp. as like with other bacteria. Den Jong observed lysis of Azotobacter cells due to phage infection, although he has not proved it by isolation and characterization of azophage from lysed cultures. But his observation was assumed as the first report on the presence of *Azotobacter* phages. After that, Monsour et al. (1955) isolated Azophages from the soil which have capable of forms plaques on the green pigmented strains of *A. vinelandii*. Hezagi and Jensen (1973), reported that phages showed more lysis of *A. vinelandii* and *A. chroococcum* than *A. beijerinckii*, while other *Azotobacter* were not reported for phage infection.

2. Isolation of the genus

The design of medium for the isolation of *Azotobacter* sp. is based on its basic nature chemoheterotrophy and dinitrogen fixation. Various methods were reported for isolation of *Azotobacter* sp. such as soil paste plate method, and silica gel method. One of the oldest methods, in which soil paste is made and fortified with calcium carbonate, potassium phosphate, and carbon sources like glucose, sucrose, mannitol, and was set in watch glass or gypsum block and kept on a Petri plate containing filter paper and allowed to incubate at 27–30°C for three to up to seven days. After incubation, the slimy colony were raised similarly, and a sieve plate could also be used. The silica gel was fortified with a suitable carbon source on nutrient isolation, and the plates were impregnated or seeded with sieved soil and allowed to incubate at 27–30°C for 48–72 h. The colonies of *Azotobacter* were found to grow around soil particles on silica gel (Becking, 2006).

But currently above methods of isolation are not in use, current isolation methods included use of different nitrogen free solid or agar medium includes Winogradsky (1938) nitrogenfree media, Burk medium (Wilson and Knight, 1952), Ashby's medium (Ashby, 1907), Norris medium (Norris, 1959), and LG medium (Lipman, 1904). These all medium are a general medium for isolation of *Azotobacter* sp. The media composed of all these six methods is somewhat similar vary with only in some carbon sources and percentage of minerals and micro and macronutrient (Table 19.2).

403

19. Azotobacter

404

	Medium				
Burk'sN ₂ -free medium (g/L) (Wilson and Knight, 1952)	Sergei Winogradsky N ₂ free medium(g/L) (Winogradsky, 1938)	LG medium (g/L) (Lipman, 1904)	Ashby's medium (g/L) (Ashby, 1907).	Norris medium(g/L) (Norris, 1959)	
MgSO ₄ 0.20, K ₂ HPO ₄ 0.80, KH ₂ PO ₄ 0.20, CaSO ₄ 0.13, FeCl ₃ 0.00145, sodium molybdate 0.000253, sucrose 20.00	KH ₂ PO ₄ 50.00, MgSO ₄ ·7H ₂ O, 25.00, NaCl, 25.00, FeSO ₄ ·7H ₂ O 1.00, Na ₂ MOO ₄ ·2H ₂ O, 1.00, MnSO ₄ ·4H ₂ O 1.00, pH 7.2	Sucrose 5.00, K_2HPO_4 , 0.20, KH_2PO_4 , 0.60, $MgSO_4$, $7H_2O$ 0.20, $CaCl_2$, $2H_2O$ 0.02, Na_2MoO_4 , $2H_2O$ 0.002, bromothymol blue (5 g/L in 0.2 N KOH), 5 mL FeEDTA (solution 16.4 g/L), 4 mL vitamin solution, 1 mL. DW: 1000 mL with DW. pH 6.0 to 6.2	Mannitol 20.00, K ₂ HPO ₄ 0.200, MgSO ₄ 0.200, NaCl 0.200, K ₂ SO ₄ 0.100, CaCO ₃ 5.000, agar 15.000 final pH 7.0	Glucose 10.00, K ₂ HPO ₄ 1.00, MgSO ₄ 0.20, CaCO ₃ 1.00, NaCl 0.20, sodium molybdate 0.005, FeSO ₄ 0.10, pH 7.0	

 TABLE 19.2
 Common nitrogen-free medium for isolation of Azotobacter.

*The pH was adjusted and autoclaved at 121 °C for 15 min. $Na_2MoO_4 \cdot 2H_2O$ and $FeSO_4 \cdot 7H_2O$ were filtered and sterilized prior to adding into the autoclaved medium

Nevertheless, none of these described media was the perfect medium to isolate specific *Azotobacter* species, although the use of specific enrichment medium help to isolate or enrich specific *Azotobacter* sp., like all above medium, are applicable for getting pure isolated colonies. For further confirmation of *Azotobacter* species, various biochemical and morphological studies are required.

2.1 Preservation of Azotobacter

Among the various methods such as cryopreservation, lyophilization, and immobilization, the simple method of preservations of *Azotobacter* is on nitrogen-free agar medium with sucrose and glucose as carbon sources. This method was found significant, which keep the organism viable for 1–10 years; the only essentially required care is that agar should not be dry. The modern method of preservation like lyophilization was found to be nonsignificant for *Azotobacter* as compared to simple preservation method (Antheunisse, 1973; Lapage et al., 1970). Becking (1961) also found the preservation of nitrogen-free agar medium with cotton paraffin seal at room temperature (RT) or at 4°C showed significant viability of *Azotobacter* up to 3–5 years. Besides these, Thompson (1987) reported successful preservation or maintenance of *Azotobacter* up to 10 years by cryopreservation in liquid nitrogen, but there are some reports of damage of membranes and loss of viability during cryopreservation.

In our laboratory at KBCNMU, Jalgaon, it was observed that the cheapest method of *Azotobacter* preservation is with dry sterile soil. The five-day biomass in cyst induced medium (1% Butanol), was aseptically mixed with dry soil and maintained at RT resulted in maintenance of viable *Azotobacter* for five to six years; similar preservation was also advocated by Vela (1974).

2.2 Some practical methods of preservation

2.2.1 Dry soil preservation

Inoculate pure *Azotobacter* culture in sucrose (1%) and 0.5% butanol containing a medium, incubate at 120 rpm at 30°C for 4–5 days. After incubation, recover biomass aseptically by centrifugation at 10,000 rpm for 10 min. Mix the biomass with dried sterile black cotton soil, seal the bottles with cotton, and preserve at RT.

2.2.2 Agar slope preservation

The simplest procedure among the reported process includes streaking on nitrogen-free medium slant with a pure culture of *Azotobacter*, allow to incubate 30°C for 48 h. Then sealed with molted paraffin, allow cooling for proper sealing and preserving the slant at 4°C.

2.2.3 Immobilization with polymers

The polymer sterile solution (1.0%-1.5%) like polymerlike alginate, Gum Acacia, Carrageenan, and Plyox could be mixed with *Azotobacter* cells, dried aseptically and preserved at 15°C with 40 ± 2% relative humidity for maintaining 80% viability up to 60 days (Rojas-Tapias et al., 2013).

3. Identification of the genus

The organism isolated from the nonspecific enrichment culture only gives the idea that the isolated organism is a free nitrogen fixer, but does not give a confirmed idea that an isolated one is a member or species of *Azotobacter*. Primary confirmatory tests for genus *Azotobacter*, done by the principal morphological test, was the cyst formation potential of the isolate. The cyst formation is the only differentiating simple laboratory test which differentiae *Azotobacter* from other nitrogen fixing organism (Table 19.3).

Organism	Microcyst formation
Azotobacter	Positive
Azomonas	Negative
Beijerinckia	Negative
Derxia	Negative
Azospirullum	Negative
Rhizobium	Negative
Klebsiella	Negative
Pseudomonas	Negative
Azotomonas	Negative

 TABLE 3
 Major differentiating characteristic i.e. Microcyst formation of Azotobacter from another genus

3.1 Cyst formation

Azotobacteraceae cells generally show the vegetative growth of a large, plump and rod-to-oval-shaped cell morphology. But during the life cycle, especially in trace conditions, the vegetative cells give rise to bacteria, a spore-like specialized spherical dormant cell known as a cyst. It has very specific morphology appearing as like spherical cell with an outermost rough layer known as exine; inner to that is a homogeneous thin layer known as in tine, which covers the central body containing nuclear material and globules. The cyst formation process is known encystment process.

Jensen (1954) showed that cyst formation is one of the prime criteria for taxonomic identification of *Azotobacter*. Because the other free nitrogen fixers like *Azotominas*, *Azomonas*, *Derxia*, etc. does not show cyst formation potential. Winogradsky in (1938) reported the induction of cyst formation by specific compounds like ethanol and butanol as a carbon source.

3.1.1 Cysts: a unique character of differentiating Azotobacter sp. from other free nitrogen fixers

Batchinskaya (1935) described a very specialized, spherical form of a cell of Azotobacteraceae. These structures are morphologically very different from the normal vegetative cells. These are nothing but the special form of cell "cyst." Those are with contractile and highly vacuolated structures in the cytoplasm. The central body of these cells covered with a thick capsule-like layer, which has been covered by another thin inner layer. Winogradsky (1938) reported the cyst formation could be induced by fortifying some special chemicals like ethanol, butanol in the nitrogen-free medium. Socolofsky and Wyss (1961), Tchan and New (1984) also reported that the use of 0.3% n-butanol induces the cyst formation in Azotobacter after 5–7 days incubation. Lin and Sadoff (1968) reported that in Azotobacter vinelandii, encystment was induced by Burk's nitrogen-free liquid media added with β -hydroxybutyrate or n-butyl alcohol or crotonate. Nevertheless, butyrate and butyraldehyde do not encourage the encystment. They also observed encystment rate was increased in absence of glucose, β 3-hydroxybutyrate, and in the presence of glucose, cells produce abortive encystment with disorganized exine, releasing viscous material. Layne and Johnson (1964) also reported that the induction of cyst by altering sucrose concentration in Burks medium. They reported induction of 80% cyst formation process after reducing 0.05% sucrose in Burks medium.

Various reports proved that as compared to normal cells of *Azotobacter*, these cysts were resistant to various chemical and physical agents like UV rays and stains (Socolofsky and Wyss, 1961.)

3.1.2 Cyst induction and cyst staining

The confirmation of *Azotobacter* species from other free nitrogen fixers, the isolated pure colonies of organism on nitrogen-free medium is streaked on the special cyst-inducing medium i.e., modified Burks medium containing 0.3% n-butanol or 3% ethanol as a carbon source. For preparations, after autoclaving of the Burk medium, the plates were poured and after solidification, 0.3% n-butanol or 3% ethanol was poured on it and allows diffusing it 2–3 h in freeze at 4°C. Then isolated culture of nitrogen fixer should be streaked, and incubated for 5–7 days at 33°C. The comparative vegetative growth was attained on sucrose containing Burk's medium. The sucrose media suppress the cyst formation. After 5–7 days,

406

3. Identification of the genus

the growth on n-butanol and ethanol medium should be morphologically observed by simple staining and like the bacterial spore; the large size spherical, uniform shape structure cyst was observed. Further confirmation is done by comparison with vegetative cells and cyst staining.

3.1.3 Cyst formation; confirmation by plate assay and staining

The cyst formation also confirmed by simple plate assay in which the organism streaked on Butanol Burk and control Burk medium and incubated for 5–6 days at 30°C (Daniel et al., 2009). After 5 days of incubation, both the plates were sprayed with cyst identification reagent i.e., a solution containing 0.5% fast blue in 5.0% acetic acid. The colonies on Butanol Burk medium show in red because of reaction of fast blue with alkylresorcinol present in cyst layer while colonies on Burk medium, i.e., vegetative cells, do not show any color change.

The simple staining method like Vela and Wyss method (1965) was reported for cyst staining. This includes the following steps: the cyst suspension from Butanol Burk medium was prepared and used for state. A loopful of culture take on a clean glass slide and add few drops of Vela and Wyss reagent (glacial acetic acid, 8.5 mL; Na₂SO₄, 3.25 g; neutral red, 200 mg; light green SF yellowish, 200 mg; ethanol, 50 mL; distilled water, 100 mL; mixed well), kept for 4–5 min, remove the excess stain by blotting paper and observe wet mount under 40x. The vegetative cell appeared green in color. The cells started encystment appeared light yellowish-green. As encystment proceeds, further, the cytoplasm condenses and appears a deeper green. The matured cyst showed a thick and compacted brownish-red exine and clear unstained area of the intine, which covers the deep green stained distinct central body.

These methods of fast blue staining also very significantly differentiate vegetative cells and cysts of Azotobacter. Fast blue-B staining of cysts is based on the principle that fast blue stains the alkylresorcinol lipids, which essentially present in the layers of cysts. The staining is performed with the Azotobacter cells which grown for 5 days in the cyst inducing/butanol containing Burk medium, and loopful of samples of the culture were placed clean glass slide heat fix it and overlay with the with a solution of 0.5% Fast Blue B in 5% acetic acid for 10 min, remove the excess stain by blotting paper and observe wet mount under 40x or air dry it and observe under oil immersion objective. Cyst will appear with the reddish layers with greenish blue central body of cyst. while the vegetative cells will not retain stains and appear with red blue layers. Carbol Fuchsin (Ziehl-Neelsen) staining is also employed for visualisation of cyst.

The cyst formation is also confirmed by UV radiation test and desiccation test. Because, the *Azotobacter* cysts were found to be comparatively more resistant than the vegetative cells to various deleterious agents and environment i.e., ultraviolet irradiation, desiccation, and sonication. It was reported that cyst required twice UV doses than vegetative cells for 90% inactivation. Similarly, cysts are comparatively high resistance to desiccation than the vegetative cells. Although cyst is shown resemblance to the bacterial endospore structure and also extremely resistant to gamma radiation, sonic treatment, and desiccation, it does not comparable for bacterial heat resistance capacity.

3.2 Confirmatory test for identification of Azotobacter sp.

Once the cyst formation ability of organism grown on nitrogen-free media was proved, and then it was assumed that the isolated bacteria growing on any nitrogen-free medium

19. Azotobacter

like Burks, Ashbys, Norris etc. is the member of genus Azotobacter. Further, the exact species level identification of the Azotobacter can be done by various morphological and biochemical tests. These tests include utilization of specific carbon sources, production of diffusible, nondiffusible pigments, tolerance or sensitivity to specific chemical compounds as the basis of various such tests, the specific species of *Azotobacter* were identified. Such differentiating, morphology and biochemical test for different species is as shown in Table 19.4.

3.2.1 Azotobacter chroococcum

408

A. chroococcum is abundantly occurring in the soil. The major morphological structure is cell appear in coccus form. They also show blunt oval avoids cell with $3.0-7.0 \,\mu m$ length \times 1.5–2.3 µm width. Cells remain motile up to 24–48 h only. The major characteristic of A. chroococcum is after aging on Ashby's or Burk medium, it produces yellow-brown nonwater-soluble pigment. Beijerinck (1901b) developed an enrichment method for A. chroococcum known as nutrient solution method which followed by purification on the solid medium for A. chroococcum (Table 19.5).

The solid medium becomes differential because it contains $CaCO_3$ which act buffering agent and maintain pH-7.4-7.5 because this is a favorable condition for growth A. chroococcum. Becking (1961) and Jensen (1965) reported that the soil has pH 7.5 and abundantly contains A. chroococcum. It was observed that soil with pH range 7.0-7.4 has 89%, 6.5–6.9 contain 57%, and 6.0–6.4 have 42% Azotobacter population (Jensen, 1965). Similarly, Jensen and Petersen (1955) advocated that lower pH of nitrogen-free medium is the growth limit factor for A. chroococcum.

For the biochemical identification of A. chroococcum, caproate and caprylate utilization test is a very important test (Table 19.6). Caproate is nothing but the ethyl hexanoate also known as hydroxyl progesterone caproate. It is ester obtained by condensing hexanoic acid and ethanol. It is also present in fruits, flower gives aromas. Among the all Azotobacter species only A. chroococcum and A. vinelandii were able to utilize this ethyl hexanol as carbon or energy sources. Hence, Azotobacter isolate was caproate positive. It indicates that organism may be only A. chroococcum or A. vinelandii. The confirmation was done by caproate and

Sr. No.	Name of organism	Biochemical test
1.	Azotobacter vinelandii	Rhamnose, erythritol, butanol, ethylene glycol 10% sodium benzoate, 0.1% phenol
2.	Azotobacter beijerinckii	Tartarate, α hydroxyl benzoate, D-galaturonate, pH 4.9–5.5
3.	Azotobacter chroococcum	pH 7.0–7.5
4.	Azotobacter paspali	0.5% bromothymol blue in sucrose medium/Sample from rhizosphere of $Paspalum$ notatum
5.	Azotobacter armeniacus	No specific addition of caprylale in Burks medium
6.	Azotobacter nigricans	Citrate, n- valerate
7.	Azotobacter salinetris	Burk medium fortified with sodium salt 1.0%-2.0%

 TABLE 19.4
 Compounds/conditions used in a selective medium.

3. Identification of the genus

Compound	g/L
Glucose	20.0
K ₂ HPO ₄	0.8
MgSO ₄	0.5
KH ₂ PO ₄	0.2
FeCl ₃ •6H ₂ O	0.005
CaCl ₂ •2H ₂ O	0.005
Agar	15
DW	1000 mL
	r
CaCO ₃	20.0
NaMoO ₄ ·2H ₂ O	0.05
рН	7.4-7.6

TABLE 19.5 Enrichment method for Azotobacter chroococcum.

TABLE 19.6 Differentiation characteristic of Azotobacter vinelandii and Azotobacter chroococcum.

Character	Azotobacter vinelandii	Azotobacter chroococcum
Rhamnose	Positive	Negative
Caproate	Positive	Positive
Caprylate	Positive	Negative
Malonate	Positive	Detectable
Mesoinositol	Positive	Negative
Cysten	Negative	Detectable
Glutarate	Positive	Negative
Glycolate	Negative	Positive

caprylic acid/utilization tests, which differentiate *Azotobacter chroococcum* from *A. vinelandii* because of *Azotobacter chroococcum* unable to utilize caprylate as carbon sources (Table 19.2). Besides that, a glycolate utilization test was found to be positive by *A. chroococcum* and negative for *A. vinelandii*. Besides these various tests like malonate, myoinositol, and Rhamnose utilization test confirm the presence of *A. chroococcum*.

3.2.2 Azotobacter vinelandii

It was isolated by Lipman (1903a,b) and recognized as *Azotobacter miscellum* by Cohen and Johnstone in (1964). The organism was first isolated from Vineland, New Jersey and so

409

recognized as *Azotobacter vinelandii* organism showed oval to short rod cell, motile with peritrichous flagella. Cells also show motility up to 24–48 h as with *Azotobacter chroococcum*. The cells show dimension $3.0-4.5 \,\mu\text{m} \log \times 1.5-2.4 \,\mu\text{m}$ width. The *A. vinelandii* shows the specific character of yellow-green fluorescent and water-soluble pigment production. Becking (2006) reported *A. vinelandii* from alkaline soil with pH 8.0–9.5 and rich in sodium chloride, alkaline sea muds, calcareous soil of Indonesia, and various localities of Boliva and South America (Becking, 1961).

Reuszer (1939) observed that addition of benzoate acid, phenolic compound, and benzoate in soil surprisingly replaces the normal *A. chroococcum* flora to *A. vinelandii*.

Reuszer (1939) mention the organism characteristically produced green pigment, on the basis of this observation, Derx (1951) designed special enriched medium for isolation of *A. vinelandii* (Table 19.7).

The Derx medium preparation with a carbon source, ethanol was used for precise isolation of *A. vinelandii* (Table 19.7). Ethanol (10 mL) should be added after autoclaving and at 40°C added in flask just before pouring plate. Besides that, Jensen (1961) also designs the Rhamnose agar medium for isolation of *A. vinelandii*. The medium is based on the principle that other *Azotobacter* sp. except for *Azotobacter vinelandii* generally unable to utilize Rhamnose as a carbon source (Thompson and Skerman, 1979). Although there are very few other strains of *Azotobacter* sp. variants either reported for Rhamnose utilization or nonutilization (Thompson and Skerman, 1979; Claus and Hempel, 1970).

Besides all of these biochemical characterizations, there are various chemical compounds selectively used by *Azotobacter vinelandii*, and hence used for its characterizations such as resorcin, ethylene glycol, and glutarate utilization test. Hence the addition of 0.1% or 0.2% of the above compound in nitrogen-free medium selectively allows *A. vinelandii* isolation from soil and other sources.

Thompson and Skerman (1979) also observed the *Azotobacter vinelandii* exceptionally uses the caproate, caprylate, and mesoinositol as a carbon source. Besides that, they also observed 0.1% phenol in nitrogen-free medium. The medium become a selective medium for isolation of *Azotobacter vinelandii*.

Further, Thompson and Skerman identified selective carbon sources like caproate (C6), caprylate (C8), short chain fatty acids, and among which mesoinositol utilization test become

	0 •
Composition	g/L
Mannitol	5.0
Or ethanol	10 mL
K ₂ HPO ₄	5.0
Sodium benzoate	10.0
pH	7.5-8.0
DW	1000

 TABLE 19.7
 Derx medium for selective screening of Azotobacter vinelandii.

an important test to differentiate the *Azotobacter vinelandii* from *Azotobacter chroococcum*. Because among all species of *Azotobacter*, *Azotobacter chroococcum* and *Azotobacter vinelandii* only utilize caproate, but for further differentiation caprylate or caprylic acid is one most important biochemical test.

Hexanoate is straight-chain saturated fatty acid anion and which is the conjugate base of hexanoic acid or caproic acid. It has also found as a human metabolite and a plant metabolite (Fig. 19.1).

Caprylic acid is the saturated fatty acid made up of octanoic eight-carbon. It is apparently present in the mammal milk, in slight amount as a component of coconut oil, palm kernel oil and even formed during yeast fermentation. It gives unpleasant smell. The caprylic acid or caproate only utilized by *Azotobacter vinelandii* and not by *Azotobacter chroococcum*.

Similarly, the mesoinositol is also utilized by only *Azotobacter vinelandii* not used by *Azotobacter chroococcum*.

3.2.3 Azotobacter beijerinckii

Azotobacter beijerinckii shows morphological similarities with Azotobacter chroococcum. Lipman (1904) isolated Azotobacter beijerinckii and named subspecies acid tolerance. Azotobacter beijerinckii although not viewed as discrete species and assumed as per a pigment deficient strain of *Azotobacter chroococcum* in the Bergey's manual edition restored species level (Buchanan and Gibbons, 1974). Then, on the basis of nonmotile nature of Azotobacter beijerinckii, it was clearly differentiated from Azotobacter chroococcum. After aging, it produces yellowish and cinnamon pigment while Azotobacter chroococcum produces blackish-brown pigment. Azotobacter beijerinckii is only species among Azotobacter, which is nonmotile and can use malonate, propionate benzoate, and D-Galacturonate as a carbon source. These are the important biochemical tests to identify the Azotobacter beijerinckii, from morphologically resemble species i.e., A. chroococcum. Similarly, the strains are exceptionally recognized as amylase positive, hence utilizes starch as a carbon source. Jensen and Petersen (1955) designed a selective medium for A. beijerinckii which have some composition as like nitrogen-free medium but instead of CaCO₃ essentially use CaCl₂ media component and pH of medium maintain slightly acidic i.e., 4.9–5.5. Jensen and Peterson medium for A. beijerinckii is based on their previous finding that all A. beijerinckii strains grow and fix nitrogen at pH 5.1, although they also reported that at alkaline pH more atmospheric

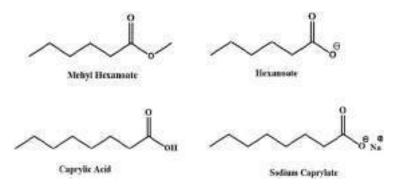


FIGURE 19.1 Structure of unusual carbon sources for *Azotobacter* differentiation.

412

Test	Azotobacter beijerinckii	Azotobacter nigricans
Motility	Negative	Negative
Black, red, violet pigments	Negative	Positive
Malonate	Positive	Detectable
Propionate	Positive	Negative
Benzoate	Positive	Negative
Glycerol	Detectable	Negative
D-galacturonate	Positive	Negative
Glucuronate	Positive	Negative
Glutarate	Negative	Negative
Citrate	Positive	Negative

 TABLE 19.8
 Differentiation characteristic: Azotobacter beijerinckii and Azotobacter nigricans.

nitrogen was fixed by *A. beijerinckii* (Jensen and Petersen, 1955). All strain of *A. beijerinckii* exceptionally produces urease enzyme; hence utilize urea as a nitrogen source (Table 19.8).

3.2.4 Azotobacter nigricans

A. nigricans was originally detected by Krasilnikov from USSR in 1949. The strain shows similarity with A. beijerinckii e.g., both is the only nonmotile species of Azotobacter. Cells are blunt, rounded rods with a dimension of $4.1-4.9 \mu m$ length $\times 1.5-2.7 \mu m$ width. The strain is characterized by various biochemical aspects like unable to utilize malonate, ethanol, pentanol, benzoate etc. To differentiate from Azotobacter beijerinckii benzoate, propionate, malonate, and galacturonic utilization test are the important differentiating biochemical test because A. beijerinckii shows these entire tests positive and A. nigricans gives it totally negative. The confirmation is also possible by testing the glutarate utilization (Table 19.8).

3.2.5 Azotobacter paspali

This species originally isolated by Winogradsky described a method of isolation of *Azotobacter* sp. i.e., silica gel plate containing a mineral solution and the sole carbon source as calcium citrate. It was a character used for identification, as younger filamentous long rods cells with yellowish-green fluorescent or red-violet water-soluble pigment colonies. In 1966 Döbereiner studied it thoroughly by growing it on N₂ free Lipman (1903a,b) and modified medium with sucrose as a sole carbon source and bromothymol blue as an indicator (Table 19.9).

Organic acid production is one of the unique characteristics of *A. paspali*, the sucrose minerals medium contains a pH indicator bromothymol blue, which differentiate colony by yellow color on a blue background after 3–4 days (Table 19.10). Similarly, the large filamentous structure is another characteristic of *A. paspali* which shows 1.12 μ m length and 1.3–1.7 μ m in width. A. paspali is the only *Azotobacter* species which had the specific rhizospheric association. Döbereiner (1970) found that *A. paspali* have specifically for a wild grass *Paspalum notatum* and few other *Paspalum* sp. i.e., *P. virgatum*, *P. plicatulum* etc.

Component	g/L
Sucrose	20.0
K ₂ HPO ₄	0.05
KH ₂ PO ₄	0.15
MgSO ₄ ·7H ₂ O	0.20
CaCl ₂	0.02
CaCo ₃	1.0
$N_2MoO_4 \cdot 2H_2O$	0.02
FeCl ₃ (10% solution)	1 drop
Agar	20
DW	1000 mL
pH	7.0
Bromothymol (0.5% ethanol)	10 mL

 TABLE 19.9
 Döbereiner sucrose mineral medium.

TABLE 19.10 Identification test for Azotobacter armeniacus and Azotobacter paspali.

Test/characters	Azotobacter armeniacus	Azotobacter paspali
Large filamentous cells in young cultures	Negative	Positive
Peroxidase	Detectable	Negative
H ₂ S production from thiosulphate	Negative	Positive
Cysteine	Detectable	Negative
Propionate	Detectable	Negative
Glycolate	Positive	Negative
Malonate	Positive	Negative
Growth at 14°C	Negative	Positive
Peroxidase	Positive	Negative
Nitrate to nitrite	Negative	Negative

3.2.6 Azotobacter armeniacus

This *Azotobacter* species was first isolated from US Armenia in 1964, fully described by Thompson and Skerman (1981). The organism was majorly differentiated by its motile cells, production of brown-black and reddish violet pigment. They also have the ability of esterase

production and citrate utilization as a carbon source. Also utilizes n-valerate and caprylate. A. armeniacus also deferentially characterized by peroxidase production, glycolate and malonate utilization from other *Azotobacter* like *Azotobacter paspali*.

The identification of important *Azotobacter* sp. could be outlined based on simple morphological and biochemical characteristics as shown in Fig. 19.2.

4. Beneficial role of the Azotobacter in agroecology

Azotobacter has played an important role in sustainable agriculture as plant growth promoting properties as well as biocontrol agent production against phytopathogen (Fig. 19.3).

4.1 Mechanism of crop productivity benefit

Azotobacter is the genus of interest for reviewing nitrogen fixation and effect on plant owing to its fast advancement and efficiency of dinitrogen fixation. The bacteria fix the

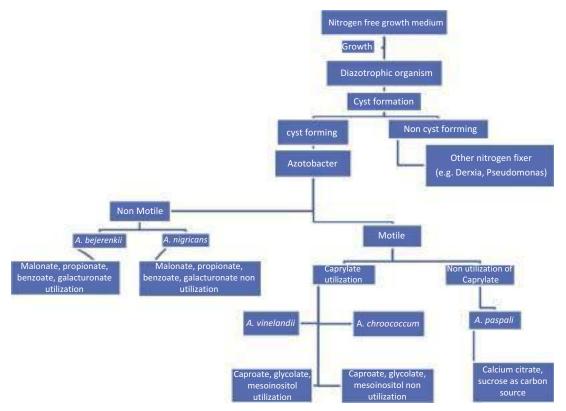


FIGURE 19.2 Key for identification of *Azotobacter* sp. based on important morphological and biochemical characteristics.

I. Bacteria

4. Beneficial role of the Azotobacter in agroecology

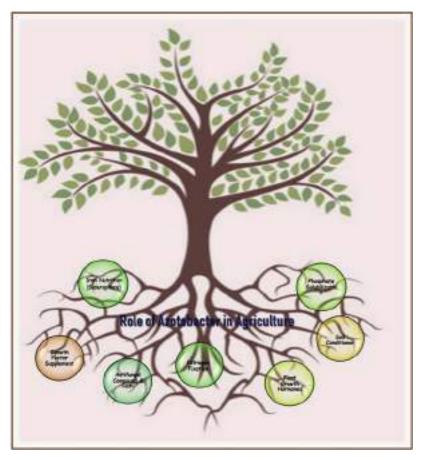


FIGURE 19.3 Role of Azotobacter in sustainable agriculture.

atmospheric nitrogen into ammonia and remaining used for the synthesis of protein, which is further return to the soil after the death of bacteria by mineralization and this will be available for the plant from soil (EI-Lattief, 2016).

The possible mechanism involves dinitrogen fixation by the transformation of atmospheric nitrogen (N₂) gas into ammonia (NH₃). The conversion to a utilizable form of nitrogen, i.e., ammonia is required for biosynthetic pathway and molecular nitrogen cycle (Murcia et al., 1997; Diaz-Barrera and Soto, 2010). The fixation of atmospheric dinitrogen by these heterotrophic aerated shaking cultures is achieved by protection of oxygen labile nitrogenase enzyme by consumption of O_2 (Robson and Postgate, 1980). This is possible with the potential of production of highly active cytochrome oxidases (Jurtshuk et al. 1978, 1981), then superoxide dismutase and catalase (Jurtshuk et al., 1984).

The nitrogen fixation; has several effects on plant growth, as provide the collective molecular nitrogen to the vegetation; the phytohormones formation affecting plant productivity and reduction of nitrate which surges accumulation of nitrogen in the plants. As stated

with the estimate, fertilizer equivalent of 20 kg N/ha for *Azotobacter* is needed (Tandon, 1991) which increase the yield of crop plants about 10%–12% (Jaga and Singh, 2010). The combined incubation of *Azotobacter* with other microorganisms like *Azospirillum*, *Pseudomonas* increased grain yield (Yousefi and Barzegar, 2014; Singh et al., 2015). The inhibitory consequence of excess ammonium ions on nitrogenase synthesis and action as well as ammonium accumulation could be prevented by nitrogen fixation by bacteria and carry fixed nitrogen to plant.

The control of dinitrogen fixation gene expression is able to control ammonia synthesis in response to 2-oxoglutarate via NifA with the balance of nitrogen and carbon (Mus et al., 2017).

Azotobacter vinelandii is the model organism for the study of nitrogen fixation under raised atmospheric oxygen condition. The protection of nitrogenase enzyme in the presence of air is possible due to respiratory and conformational resistance of enzyme and position of the enzyme in a cell. The nitrogenase activity is associated with soil moisture. The increase in soil moisture is associated with the exponential rise in the rate of acetylene reduction (Dighe et al., 2010). The growth augmentation is due to mutual communication of the strains along with growth promoting substance synthesis. This shows that the efficiency outcome is subject to the phases of plant growth (Vikhe, 2014).

4.2 Role of Azotobacter in crop productivity

Azotobacter is the dynamic, free-living heterotrophic nitrogen fixer majorly found in the alkaline or neutral soil habitat. *Azotobacter* works as plant growth promoting rhizobacteria (PGPR) apart from the fixation of molecular nitrogen.

The effect of *Azotobacter* on plant productivity include, increase in seed germination capacity, promote germination (20%–30%) through the making of the plant growth encouraging substance, which causes a decrease in chemical phosphorus and nitrogen by 25% that resulted in stimulation of crop growth. The straight promoting effect PGPR on plant growth comprise of formation and release of subordinate metabolite like growth regulator as well as smoothing uptake of nutrients from the root environment (Glick, 1995; Polyanskaya et al., 2002).

Azotobacter produces various vegetal growth promoting elements, for instance, cytokinin, auxins, and Gibberellic acid, which are the principal constituents for governing plant growth by improvement in nitrogen content and plant mass. These hormonal constituents from the root surface or rhizosphere also affect the growth of the familiarly related developed plants (EI-Lattief, 2016). Hormones enhance phosphate solubilization (Ramon et al., 1972) influence nutrient uptake by increasing phosphatase activity (Hoflich et al., 1994), increases water and mineral uptake (Bashan and Levanony, 1991), production of amino acids and vitamins, the bioactive constituents. The phytohormone synthesis is also associated with the production of extracellular compounds such as riboflavin, Vit B12, biotin, thiamine, pyridoxine, cyanocobalamin, folic acid and pantothenic acid.

The *Azotobacter* number is usually less within the uncultured soils and rhizosphere of the crop plants. The organic matter from the soil and root exudates nourishes the *Azotobacter* and help further to fix atmospheric N (Maryenko, 1964). *Azotobacter* biofertilizer with C: N ratio (20:1) is indicating the stability of biofertilizer.

The *Azotobacter chroococcum* is having a worthy influence on soil fertility and plant nutrition by promoting significant uptake of N and P (Wani et al., 2016). The synthesis and secretion of bioactive compounds like gibberellic acid, vitamin B, biotin, pantothenic acid, nicotinic

acid resulted in boosting the growth of plant root (Rao, 1986). The secretion of ammonia by *Azotobacter* in the rhizosphere is also important in modification of crop nutrient uptake (Narula and Gupta, 1986). The multi-copper protein family bacteria produced polyphenol oxidases (PPOs) and phenol oxidases (POs). The family Azotobacteraceae is also assumed to produce POs (Herter et al., 2011).

4.2.1 Vitamins produced by Azotobacter

Vitamins play a crucial role in the physiological functioning of lives formed by numerous groups of bacteria (Revillas et al., 2000). The vitamin production by *Azotobacter* species is observed in a favorable environment. The B-group vitamins such as pantothenic acid, biotin, niacin, riboflavin are produced by *A. chroococcum* strain H23 (CECT 4435) and *Azotobacter vinelandii* strain ATCC 12837.

4.2.2 Amino acids produced by Azotobacter

At the diazotrophic conditions in glucose-supplemented culture media, *Azotobacter* species produced different amino acids such as tryptophan, lysine, glutamic acid, and methionine (Gonzalez-Lopez et al., 1983). *A. vinelandii* and *A. chroococcum* are recognized for the production of aspartic acid, serine, glutamic acid, glycine, histidine, threonine, arginine, alanine, proline, cysteine, valine, lysine, isoleucine, phenylalanine, tyrosine, methionine, and leucine (Revillas et al., 2005).

4.2.3 Phosphate solubilization

Phosphobacteria is the noteworthy microorganisms for the transformation of phosphorous. The hydrolysis of organic and inorganic phosphorus from insoluble compounds is occurred by phosphate solubilizing bacteria. Thus, P-solubilization efficiency of the microorganisms intended for phosphate nourishment of plant is a very significant character.

4.2.4 Plant growth hormones (IAA, GA)

Several *Azotobacter* species produce Indole acetic acid (IAA; $2.09-33.28 \mu g/mL$) (Spaepen et al., 2007). IAA producing PGPR strains upsurge root length resulting in larger root surface zone, which allows plants to access additional nutrients from the soil. The IAA is accountable for the division and differentiation of plant cells and tissues as well as stimulation of root elongation (Ahmad et al., 2008).

Azotobacter chroococcum is the signature model for its role in plant nourishment and its influence on soil richness by synthesis of plant development hormone. A. vinelandii cells can biosynthesize at least three molecules the intracellular polyester poly- β -hydroxybutyrate (PHB), the extracellular polyesaccharide alginate, and catechol compounds (siderophores).

4.2.5 Biopolymers as a soil conditioner

The biopolymers in the rhizosphere have different natural functions such as self-adhesion of cells into biofilms, surface adhesion, the creation of defensive barriers, water retention about roots, and nutrient accumulation. The biopolymer has its role as a soil modifier to build up slope stability, lessen transport of objects in overflow water, cut the passage of heavy metals, and generation of dust (Larson et al., 2012).

Biopolymers strengthen the soil by ecofriendly way and required in low concentration. The polysaccharide biopolymer has hydroxyl groups on its surfaces that encourage hydrogen bond formation with water molecules to make hydrophilic nature that allowing the formation of viscous hydrogels or hydrocolloids.

In drought condition, the biopolymer can create hydrogen bonding toward clay particle or subsidiary ionic bonding with clay particle in existence of earth metals. The direct and indirect bonding primes the creation of a steady biopolymer-clay matrix, which resulted in a substantial rise in soil cohesion. The mixing of biopolymers with coarse and clay particles is therefore anticipated to offer best firming possessions, owing to the mixture of amplified motorized resistance among rough particles, and a cementation outcome among biopolymer-clay matrices (Chang et al. 2016).

Azotobacter species (A. beijerinckii, A. chroococcum, A. vinelandii) produce extracellular (alginate) and intracellular (Poly- β -hydroxybutyrate) polymer (Haleem Khan et al., 2015). Pseudomonas and Azotobacter species produced alginates (Remminghorst and Rehm, 2006). Alginate is a straight-chain polymer composed of a varying number of (1–4)- β -D-mannuronic acid and a-L-guluronic acid, its C-5-epimer has an extensive variety of applications such as a stabilizer, thickener, emulsifier and gelling agent in food, and in addition to textile and pharmaceutical industries. The polyhydroxyalkanoate (PHAs) is intracellular polyesters polymer.

The bacterium *Azotobacter vinelandii* produce the alginate and PHAs as important polymers with an excess of carbon source and limiting phosphorous and oxygen. Alginate is produced as extracellular polysaccharide in *A. vinelandii* and *P. aeruginosa*, while PHAs is involved in cyst differentiation in *A. vinelandii* (Sadoff, 1975). The formation of the cyst is the result of the intracellular accumulation of PHA/PHB inside the cytoplasm outlined by lipoprotein double wall in the environment of excess carbon and limited nitrogen, phosphorus, or oxygen condition. Once the carbon source is exhausted, cysts are oxidized to work as energy source quickly with the involvement of PHB depolymerases enzyme (da Silva and Garcia-Cruz, 2010).

A. vinelandii also produces the intracellular polyester PHB (polymer of the polyhydroxyalkanoates family), which is a biodegradable and biocompatible thermoplastic and benefited as a supplementary for majority plastics such as polyethylene and polypropylene. The bacteria can accumulate PHB polyester intracellularly as both a carbon and energy reserve material. PHB is made up of around 150 diverse hydroxyl alkanoic acids (Schroth and Hancock, 1982).

The other important function that is achieved with PHB in *A. vinelandii* is the protection of nitrogenase enzyme by ensuring the bacterial respiration even in absence of exogenous carbon and energy source by avoiding decrease of oxygen and maintaining respiratory function (Page et al., 1992; de Almeida et al., 2004).

In our Lab at KBC North Maharashtra University, Jalgaon, *Azotobacter* biopolymer was exploited for various benefits like improved germination, water holding capacity, soil porosity, organic content as soil conditioner along with its role as bioflocculent, for toxic heavy metal and dye removal (Patil et al., 2010, 2011; Mohite and Patil, 2014; Mohite et al., 2017) (Fig. 19.4).

Azotobacter is renowned for the production of diverse forms of subsidiary metabolites, for instance, plant growth hormones (IAA, nicotine, and gibberellins), amino acids (Thiamine),

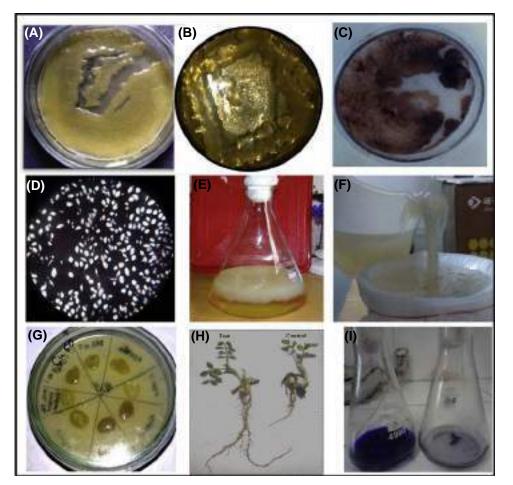


FIGURE 19.4 *Azotobacter* biopolymer and its explored applications (At KBC NMU laboratory), (A), (B), and (C) Different *Azotobacter* sp. on nitrogen-free medium, (D) *Azotobacter* with its biopolymer sheath by negative staining, (E) and (F) *Azotobacter* biopolymer production and recovery; (G), (H), and (I) *Azotobacter* biopolymer applications for heavy metal accumulation, plant growth promotion, and dye removal, respectively.

vitamins (Riboflavin), siderophores, antifungal substance (Myresiotis et al. 2015). The growth encouraging effects like effect on the shoot and root length, germination of seeds is due to the growth encouraging elements like IAA, gibberellic acid (GA), and nicotinic acid (Ahmad et al., 2005).

These subordinate metabolites inspire the growth promoting effect by expelling auxins, vitamins, amino acids, provision of iron to plants by siderophores and poly hydroxyl buty-rate (PHB) for large-scale production of alginic acid. The phytohormone production capacity is dispersed among plant-related bacteria, about 80% of plant rhizosphere bacteria can make plant growth encouraging elements.

4.3 Azotobacter as biocontrol agent

4.3.1 Antifungal compounds

The antibiotic production is among the most focused biocontrol mechanisms for fighting against phytopathogens. *Azotobacter* can offer protection from drought and generates antifungal compounds that are responsible for inhibition of the growth of soil-borne fungi such as *Alternaria, Curvularia, Aspergillus, Helminthosporium,* and *Fusarium* (Mali and Bodhankar, 2009). The *Azotobacter* species produce diverse types of antibiotics like 2,3-dihydroxybenzoic acid, azotochelin, aminochelin, azotobactin and protochelin (Kraepiel et al., 2009).

Azotobacter species worked as biocontrol agents against various plant pathogens (Mali and Bodhankar, 2009; Agrawal and Singh, 2002). The species of Arthrobacter and Azotobacter inhibited *F. verticillioides* root colonization which has suppressed fumonisin B-1 creation by *A. armeniacus. F. oxysporum* causes infection to several crops such as chilli, and pigeon pea. The *A. vinelandii* showed a maximum zone of inhibition (40 mm) against *F. oxysporum* confirming the antifungal activity (Cavaglieri et al., 2005; Bhosale et al., 2013).

The growth of various pathogenic fungi in the rhizosphere is inhibited by the antibiotic secreted by *Azotobacter chroococcum* (Subba Roa, 2001). *Azotobacter* inhibited the growth of *Rhizoctonia solani* by producing an antifungal antibiotic which inhibits it (Vikhe, 2014). *Azotobacter* sp. could produce antifungal compounds opposing the pathogens alike *Tricho- derma* sp., *Alternaria* sp., *Fusarium* sp. (Bjelić et al., 2015).

4.3.2 HCN production

In addition to the production of antibiotics, some of the rhizobacteria accomplished production of HCN, which is a subordinate metabolite of volatile nature that inhibits the microbes and affect the crop growth and development. It is a powerful inhibitor of various metal containing enzymes, particularly copper encompassing cytochrome C oxidases. HCN synthetase enzyme is responsible for formation HCN from glycine. The enzyme is present in association with the plasma membrane of rhizobacteria.

4.3.3 Siderophore production by Azotobacter

Siderophores are low molecular weight complexes formed by fungi and bacteria as iron (Fe) chelating agents. Under the iron deficient condition, at neutral to alkaline pH, different bacteria produce the siderophores (Sharma and Johri, 2003). *Azotobacter* excretes siderophores under deprivation of iron as *A. vinelandii* (Page and Von Tigerstrom, 1988), which can combine to iron and form sturdy complexes which are later transferred into the cell by extreme precise transporters (Page et al., 2003). The reported siderophore for *A. vinelandii* is the azotochelin (bis(catechol)), aminochelin (monocatechols), the 2,3-dihydroxybenzoic acid and protochelin (tris(catechol)) and the yellow-green fluorescent pyoverdine-like azotobactin (Kraepiel et al., 2009). *A. vinelandii* generate minimum five diverse siderophores having an antibacterial effect such as 2, 3- dihydroxybenzoic acid, azotochelin (bis-catechol), protochelin (tris-catechol), aminochelin (monocatechols), and the yellow-green fluorescent pyoverdine-like azotobactin.

The pathogenic microorganism proliferation could be prevented by siderophore produced by *Azotobacter* by confiscating Fe^{3+} in the vicinity of the root.

References

Siderophore producing *Azotobacter* can prevent the proliferation of pathogenic microorganisms by requisitioning Fe^{3+} in the locality of the root. Although the plant can use the iron with the help of bacterial siderophores, the whole concentration is perhaps too little to pay significant iron uptake by the plant.

In modern agriculture practices, *Azotobacter* could support to reduce the chemical fertilizers practice. The urea adaptive nature of *Azotobacter* facilitates the plant growth improvement by combined inoculation of chemical and bacterial fertilizer (Shrivastava et al., 2015).

5. Outlook

Azotobacter is identified for its potential in diverse fields. India is well known for its geographical and biological diversity, and there are tremendous chances to get the more potent and versatile *Azotobacter* strains, which will act as a potential candidate for agriculture, fermentation, and other industrial applications. Hence, the present chapter focuses on simple morphological and biochemical keys and techniques for screening of *Azotobacter* which is the need for the economic laboratory studies in developing country like India.

Acknowledgment

Authors are thankful to Late Prof. Jan-Hendrick Becking (the pioneer Dutch biologist) and **Prof. Bernard Glick** (University of Waterloo, Canada) for the inspiration and encouragement.

The corresponding author, SVP is kindly acknowledging the Department of Biotechnology, New.

Delhi for the Indo-US Foldscope Major Research Project grant (Grant No. BT/IN/Indo-US/Foldscope/39/2015). And Co-author, BVM acknowledge the Science and Engineering Research Board (SERB) under the Start Up Research Grant, Young Scientist (YSS/2015/001722) for providing financial support.

References

- Agarwal, S.K., Singh, S.S., Verma, S., Kumar, S., 2000. Antifungal activity of anthraquinone derivatives from Rheum emodi. J. Ethnopharmacol. 72 (1–2), 43–46.
- Agrawal, N., Singh, H.P., 2002. Antibiotic resistance and inhibitory effect of *Azotobacter* on soil borne plant pathogens. Indian J. Microbiol. 42 (3), 245–246.
- Ahmad, F., Ahmad, I., Khan, M.S., 2005. Indole acetic acid production by the indigenous isolates of Azotobacter and fluorescent Pseudomonas in the presence and absence of tryptophan. Turkish J. Biol. 29 (1), 29–34.
- Ahmad, F., Ahmad, I., Khan, M.S., 2008. Screening of free-living rhizospheric bacteria for their multiple plant growth promoting activities. Microbiol. Res. 163 (2), 173–181.
- Antheunisse, J., 1973. Viability of lyophilized microorganisms after storage. Antonie Van Leeuwenhoek 39 (1), 243-248.
- Aquilanti, L., Favilli, F., Clementi, F., 2004. Comparison of different strategies for isolation and preliminary identification of *Azotobacter* from soil samples. Soil Biol. Biochem. 36 (9), 1475–1483.

Ashby, S.F., 1907. Some observations on the assimilation of atmospheric nitrogen by a free living soil organism.— Azotobacter chroococcum of Beijerinck. J. Agric. Sci. 2 (1), 35–51.

Bagyaraj, D.J., Patil, R.B., 1975. Azotobacter research in Karnataka. Curr. Res. 4, 181-184.

- Bashan, Y., Levanony, H., 1991. Alterations in membrane potential and in proton efflux in plant roots induced by *Azospirillum brasilense*. Plant Soil 137 (1), 99–103.
- Batchinskaya, A.A., 1935. Structure and development of Azotobacter. Trudy vsesoyuznogo nauchno isledovotel. Skogo Instituta Selkskokhozyaistvennoi Mikrobiologya 6 (1), 3.

- 422
- Becking, J.H., 2006. The family Azotobacteraceae. In: Dworkin, M., Falkow, S., Rosenberg, E., Schleifer, K.-H., Stackebrandt, E. (Eds.), The Prokaryotes. Springer, New York, pp. 759–783.
- Becking, J.H., 1961. Studies on the nitrogen-fixing bacteria of the genus *Beijerinckia*: II. Mineral nutrition and resistance to high levels of certain elements in relation to soil type. Plant Soil 297–322.
- Becking, S., 1974. Family Azotobacteraceae. In: Bergey's Manual of Systematic Bacteriology, vol. 1. Williams and Wilkins, London, pp. 219–230, 1984.
- Beijerinck, M.W., 1901a. Uber oligonitrophile mikroben. Zbl. Bakt. Abf. II 7, 561.
- Beijerinck, M.W., 1901. On oligonitrophilous bacteria. Proc. Koninklijke Nederl. Akademie van Wetenschappen 3, 586–595.
- Buchanan, R.E., 1925. General Systematic Bacteriology, first ed. Villiem and Wilkins, Co, p. 194.
- Bhosale, H.J., Kadam, T.A., Bobade, A.R., 2013. Identification and production of Azotobacter vinelandii and its antifungal activity against Fusarium osporum. J. Environ. Biol. 34, 177–182.
- Bishop, P.E., Jarlenski, D.M., Hetherington, D.R., 1980. Evidence for an alternative nitrogen fixation system in *Azotobacter vinelandii*. Proc. Natl. Acad. Sci. U.S.A. 77 (12), 7342–7346.
- Bjelić, D.D., Marinković, J.B., Tintor, B.B., Tančić, S.L., Nastasić, A.M., Mrkovački, N.B., 2015. Screening of Azotobacter isolates for PGP properties and antifungal activity. Zbornik Matice srpske za prirodne nauke (129), 65–72.
- Buchanan, R.E., Gibbons, N.E. (Eds.), 1974. Bergey's Manual of Determinative Bacteriology, eighth ed. Williams & Wilkins Co., Baltimore, Md, p. 21202.
- Bush, J.A., Wilson, P.W., 1959. A non-gummy chromogenic strain of Azotobacter vinelandii. Nature 184 (4683), 381.
- Cavaglieri, L.R., Andrés, L., Ibáñez, M., Etcheverry, M.G., 2005. Rhizobacteria and their potential to control *Fusarium verticillioides*: effect of maize bacterisation and inoculum density. Antonie Van Leeuwenhoek 87 (3), 179–187.
- Chang, I., Im, J., Cho, G.C., 2016. Introduction of microbial biopolymers in soil treatment for future environmentallyfriendly and sustainable geotechnical engineering. Sustainability 8 (3), 251.
- Cohen, G.H., Johnstone, D.B., 1964. Extracellular polysaccharides of Azotobacter vinelandii. J. Bacteriol. 88 (2), 329-338.
- Claus, D., Hempel, W., 1970. Specific substrates for isolation and differentiation of *Azotobacter vinelandii*. Arch. Mikrobiol. 73 (1), 90–96.
- Daniel, S., Odon, V., Yanet, R., Soledad, M., Miguel, C., Guadalupe, E., 2009. Isolation and characterization of *Azotobacter vinelandii* mutants impaired in alkylresorcinol synthesis: alkylresorcinols are not essential for cyst desiccation resistance. J. Bacteriol. 191 (9), 142–148.
- da Silva, A.N., Garcia-Cruz, C.H., 2010. Biopolymers by azotobacter vinelandii. In: Biopolymers. In Tech., pp. 413–438
- de Almeida, A., Ruiz, J.A., López, N.I., Pettinari, M.J., 2004. Bioplásticos: una alternativa ecológica. Química Viva 3 (3).
- De Jong, L.D.D., 1938. Das Verhalten von Azotobacter chroococcum unter abnormen Lebensbedingungen. Arch. Mikrobiol. 9 (1–5), 223–252.
- Derx, H.G., 1951. L'accumulation spécifique de l'Azotobacteragile Beijerinck et de l' Azotobacter vinelandii Lipman. Proc. Koninklijke Nederl. Akademie Wetenschappen - Ser. B Palaeontol. Geol. Phys. Chem. Anthropol. 54, 624–634.
- Diaz-Barrera, A., Soto, E., 2010. Biotechnological uses of Azotobacter vinelandii: current state, limits and prospects. Afr. J. Biotechnol. 9 (33).
- Dighe, N.S., Shukla, D., Kalkotwar, R.S., Laware, R.B., Bhawa, S.B., Gaikwad, R.W., 2010. Nitrogenase enzyme: a review. Der Pharm. Sin. 1 (2), 77–84.
- Döbereiner, J., 1970. Further research on *Azotobacter paspali* and its variety specific occurrence in the rhizosphere of *Paspalum notatum* Flugge. Zentralblatt fürBakteriologie, Parasitenkunde, Infektionskrankheitenund Hygiene, Abt. 2 (124), 224–230.
- Döbereiner, J., 1966. *Azotobacter paspali* sp. n., uma bactéria fixadora de nitrogênio na rizosfera de Paspalum. Pesqui. Agropecu. Bras. 1 (1), 357–365.
- EI-Lattief, E.A., 2016. Use of *Azospirillum* and *Azotobacter* bacteria as biofertilizers in cereal crops: a review. Int. J. Res. Eng. Appl. Sci. 6 (7), 36–44.
- Garg, S.K., Bhatnagar, A., Kalla, A., Narula, N., 2001. In vitro nitrogen fixation, phosphate solubilization, survival and nutrient release by *Azotobacter* strains in an aquatic system. Bioresour. Technol. 80 (2), 101–109.
- Glick, B.R., 1995. The enhancement of plant growth by free-living bacteria. Can. J. Microbiol. 41 (2), 109-117.
- Gonzalez-Lopez, J., Salmeron, V., Moreno, J., Ramos-Cormenzana, A., 1983. Amino acids and vitamins produced by Azotobacter vinelandii ATCC 12837 in chemically-defined media and dialysed soil media. Soil Biol. Biochem. 15 (6), 711–713.

- Haleem Khan, A.A., Naseem, B., Vardhini, V., 2015. Synthesis of extracellular and intracellular polymers in isolates of *Azotobacter* sp. Int. J. Renew. Energy Technol. 4 (12), 231–233.
- Hegazi, N.A., Jensen, V., 1973. Studies of *Azotobacter* bacteriophages in Egyptian soils. Soil Biol. Biochem. 5 (2), 231–243.
- Herter, S., Schmidt, M., Thompson, M.L., Mikolasch, A., Schauer, F., 2011. A new phenol oxidase produced during melanogenesis and encystment stage in the nitrogen-fixing soil bacterium *Azotobacter chroococcum*. Appl. Microbiol. Biotechnol. 90 (3), 1037–1049.
- Höflich, G., Wiehe, W., Kühn, G., 1994. Plant growth stimulation by inoculation with symbiotic and associative rhizosphere microorganisms. Experientia 50 (10), 897–905.
- Jaga, P.K., Singh, V., 2010. Effect of biofertilizer, nitrogen and sulphur on sorghum-mustard cropping system. In: Proceedings of National Seminar on Soil Security for Sustainable Agriculture Held at College of Agriculture, Nagpur (*MS on February 27-28, 2010*) "Xxii Savetovanje O Biotehnologiji" Zbornik Radova, Knjiga, vol. 1, p. 2017.
- Jensen, H.L., 1954. The Azotobacteriace. Bacteriol. Rev. 18, 195-214.
- Jensen, H.L., 1965. Non-symbiotic nitrogen fixation. 436–480. In: Bartholomew, W.V., Clark, F.E. (Eds.), Soil Nitrogen, Monograph 10. American Society of Agronomy, Madison.
- Jensen, V., 1961. Rhamnose for detection and isolation of Azotobacter vinelandii Lipman. Nature 190 (4778), 832-833.
- Jensen, V., Petersen, E.J., 1954. Studies on the Occurrence of *Azotobacter* in Danish Forest Soils. 95–110. Royal Veterinary and Agricultural College Yearbook 1954. Kandrup & Wunsch, Copenhagen.
- Jensen, V., Petersen, E.J., 1955. Taxonomic studies on *Azotobacter Chroococcum* Beij. And *Azotobacter Beijerinckii*. In: Royal Veterinary and Agricultural College, Yearbook 1955. Kandrup and Wunsch, Copenhagen, pp. 107–126.
- Jurtshuk Jr., P., Mueller, T.J., Wong, T.Y., 1981. Isolation and purification of the cytochrome oxidase of Azotobacter vinelandii. Biochim. Biophys. Acta Bioenerg. 637 (2), 374–382.
- Jurtshuk, P., Liu, J.K., Moore, E.R., 1984. Comparative cytochrome oxidase and superoxide dismutase analyses on strains of *Azotobacter vinelandii* and other related free-living nitrogen-fixing bacteria. Appl. Environ. Microbiol. 47 (5), 1185–1187.
- Jurtshuk, P., Mueller, T.J., McQuitty, D.N., Riley, W.H., 1978. The cytochrome oxidase reaction in Azotobacter vinelandii and other bacteria. In: Functions of Alternative Terminal Oxidases, pp. 99–121.
- Kraepiel, A.M.L., Bellenger, J.P., Wichard, T., Morel, F.M.M., 2009. Multiple roles of siderophores in free-living nitrogen-fixing bacteria. Biometals 22 (4), 573.
- Krassilnikov, N.A., 1949. Opredelitelv Bakterii I Actinomicetov. Akademii Nauk SSSR Moscow, p. 328.
- Lapage, S.P., Shelton, J.E., Mitchell, T.G., Mackenzie, A.R., 1970. Chapter II culture collections and the preservation of bacteria. In: Methods in Microbiology, vol. 3. Academic Press, pp. 135–228.
- Larson, S.L., Newman, J.K., Griggs, C.S., Beverly, M., Nestler, C.C., 2012. Biopolymers as an alternative to petroleumbased polymers for soil modification; ESTCP ER-0920: treatability studies (No. ERDC-TR-12-8). Eng. Res. Dev. Cent. Vicksburg Ms Environ. Lab 1–34.
- Layne, J.S., Johnson, E.J., 1964. Natural factors involved in the induction of cyst formation in *Azotobacter*. J. Bacteriol. 87 (3), 684–689.
- Levine, N.D., 1975, 21202. xxvi+ 1246 pp. \$45.00. J. Protozool. In: Buchanan, R.E., Gibbons, N.E. (Eds.), 1974. Bergey's Manual of Determinative Bacteriology, vol. 22. Williams & Wilkins Co., Baltimore, Md, p. 7 (1).
- Lin, L.P., Sadoff, H.L., 1968. Encystment and polymer production by *Azotobacter vinelandii* in the presence of β-hydroxybutyrate. J. Bacteriol. 95 (6), 2336–2343.
- Lipman, J.G., 1903a. Experiments on the transformation and fixation of nitrogen by bacteria. Rep. NJ St. Agric. Exp. Stn 217–285.
- Lipman, J.G., 1903b. Nitrogen-fixing Bacteria (Ph.D. thesis). Cornell University, Ithaca, New York. June 1903. New Jersey State Agricultural Experiment Station, 16th Annual Report. State Printers. New Jersey.
- Lipman, J.G., 1904. Soil bacteriological studies. Further contributions to the physiology and morphology of the members of the Azotobacter group. Rep. New Jersey State Agric. Exp. Stn. 25, 237–289.
- Lipman, J.G., 1905. Further contributions to the physiologie and morphologie of members of the Azotobacter group. Ann. Rep. New Yersey State Agricult. Exper. Stat 25, 237. Seventeenth Annual Report over 1904. State Printers. New Jersey.
- Mali, G.V., Bodhankar, M.G., 2009. Antifungal and phytohormone production potential of Azotobacter chroococcum isolates from Groundnut (Arachis hypogea L.) rhizosphere. Asian J. Exp. Sci. 23 (1), 293–297.
- Maryenko, V.G., 1964. Dokt TSHA 99 (2), 399-406, 357-360.

- Mohite, B.V., Patil, S.V., 2014. A novel biomaterial: bacterial cellulose and its new era applications. Biotechnol. Appl. Biochem. 61 (2), 101–110.
- Mohite, B.V., Koli, S.H., Narkhede, C.P., Patil, S.N., Patil, S.V., 2017. Prospective of microbial exopolysaccharide for heavy metal exclusion. Appl. Biochem. Biotechnol. 183 (2), 582–600.
- Monsour, V., Wyss, O., Kellogg Jr., D.S., 1955. A bacteriophage for Azotobacter. J. Bacteriol. 70 (4), 486.
- Murcia, R., Rodelas, B., Salmeron, V., Martínez-Toledo, M.V., González-López, J., 1997. Effect of the herbicide simazine on vitamin production by *Azotobacter chroococcum* and *Azotobacter vinelandii*. Appl. Soil Ecol. 6 (2), 187–193.
- Mus, F., Tseng, A., Dixon, R., Peters, J.W., 2017. Diazotrophic growth allows Azotobacter vinelandii to overcome the deleterious effects of a glnE deletion. Appl. Environ. Microbiol. 83 (13), e00808–e00817.
- Myresiotis, C.K., Vryzas, Z., Papadopoulou-Mourkidou, E., 2015. Effect of specific plant-growth-promoting rhizobacteria (PGPR) on growth and uptake of neonicotinoid insecticide thiamethoxam in corn (*Zea mays L.*) seedlings. Pest Manag. Sci. 71 (9), 1258–1266.
- Narula, N., Gupta, K.G., 1986. Ammonia excretion by Azotobacter chroococcum in liquid culture and soil in the presence of manganese and clay minerals. Plant Soil 93 (2), 205–209.
- Norris, J.R., 1959. The isolation and identification of Azotobacter. Lab. Pract. 8, 239–243.
- Nosrati, R., Owlia, P., Saderi, H., Rasooli, I., Malboobi, M.A., 2014. Phosphate solubilization characteristics of efficient nitrogen fixing soil *Azotobacter* strains. Iran. J. Microbiol. 6 (4), 285.
- Page, W.J., Shivprasad, S., 1991. Azotobacter salinestris sp. nov., a sodium-dependent, microaerophilic, and aeroadaptive nitrogen-fixing bacterium. Int. J. Syst. Evol. Microbiol. 41 (3), 369–376.
- Page, W.J., Von Tigerstrom, M., 1988. Aminochelin, a catecholamine siderophore produced by Azotobacter vinelandii. Microbiology 134 (2), 453–460.
- Page, W.J., Kwon, E., Cornish, A.S., Tindale, A.E., 2003. The csbX gene of *Azotobacter vinelandii* encodes an MFS efflux pump required for catecholate siderophore export. FEMS Microbiol. Lett. 228 (2), 211–216.
- Page, W.J., Manchak, J., Rudy, B., 1992. Formation of poly (hydroxybutyrate-co-hydroxyvalerate) by Azotobacter vinelandii UWD. Appl. Environ. Microbiol. 58 (9), 2866–2873.
- Patil, S.V., Salunke, B.K., Patil, C.D., Salunkhe, R.B., 2011. Studies on amendment of different biopolymers in sandy loam and their effect on germination, seedling growth of *Gossypium herbaceum* L. Appl. Biochem. Biotechnol. 163 (6), 780–791.
- Patil, S.V., Salunkhe, R.B., Patil, C.D., Patil, D.M., Salunke, B.K., 2010. Bioflocculant exopolysaccharide production by Azotobacter indicus using flower extract of Madhuca latifolia L. Appl. Biochem. Biotechnol. 162 (4), 1095–1108.
- Phadnis, S.H., Dimri, G.P., Das, H.K., 1988. Segregation characteristics of multiple chromosomes of *Azotobacter* vinelandii. J. Genet. 67 (1), 37–42.
- Polyanskaya, L.M., Vedina, O.T., Lysak, L.V., Zvyagintsev, D.G., 2002. The growth-promoting effect of *Beijerinckia mobilis* and *Clostridium* sp. cultures on some agricultural crops. Microbiology 71 (1), 109–115.
- Pribram, E., 1933. Klassification der Schizomyceten. F. Deuticke, Leipzig, pp. 1–143.
- Premakumar, R., Jacobitz, S., Ricke, S.C., Bishop, P.E., 1996. Phenotypic characterization of a tungsten-tolerant mutant of Azotobacter vinelandii. J. Bacteriol. 178 (3), 691–696.
- Ramaswamy, P.P., Mathan, K.K., Nair, K.S., 1977. Azotobacter population in red and black soils of Tamil Nadu (India). Mysore J. Agric. Sci. 11, 364–366.
- Ramon, A., Barea, J.M., Callao, V., 1972. Phosphate dissolving and nitrogen fixing microorganism and its possible influence on soil fertility. Agrochimica 16, 345–350.
- Rao, D.L.N., 1986. Nitrogen Fixation in Free Living and Associative Symbiotic Bacteria. In: Soil Microorganisms and Plant Growth. Oxford and IBH Pub., New Delhi, pp. 116–140.
- Rascio, N., La Rocca, N., 2008. Jorgensen Editors-In-Chief: Sven Erik, Fath Brian. Biological Nitrogen Fixation, pp. 412–419.
- Rediers, H., Vanderleyden, J., De Mot, R., 2004. Azotobacter vinelandii: a Pseudomonas in disguise? Microbiol.-SGM 150, 1117–1119.
- Remminghorst, U., Rehm, B.H., 2006. Bacterial alginates: from biosynthesis to applications. Biotechnol. Lett. 28 (21), 1701–1712.
- Reuszer, H.W., 1939. The effect of benzoic acid compounds upon the abundance of microorganisms, including *Azotobacter* organisms, in a soil. Proc. Third Comm. Int. Soc. Soil Sci. A 151–160.

- Revillas, J.J., Rodelas, B., Pozo, C., Martinez-Toledo, M.V., López, J.G., 2005. Production of amino acids by Azotobacter vinelandii and Azotobacter chroococcum with phenolic compounds as sole carbon source under diazotrophic and adiazotrophic conditions. Amino Acids 28 (4), 421–425.
- Revillas, J.J., Rodelas, B., Pozo, C., Martínez-Toledo, M.V., González-López, J., 2000. Production of B-group vitamins by two Azotobacter strains with phenolic compounds as sole carbon source under diazotrophic and adiazotrophic conditions. J. Appl. Microbiol. 89 (3), 486–493.
- Robson, R.L., Postgate, J.R., 1980. Oxygen and hydrogen in biological nitrogen fixation. Annu. Rev. Microbiol. 34 (1), 183–207.
- Robson, R.L., Jones, R., Robson, R.M., Schwartz, A., Richardson, T.H., 2015. Azotobacter genomes: the genome of Azotobacter chroococcum NCIMB 8003 (ATCC 4412). PloS One 10 (6), e0127997.
- Rojas-Tapias, D., Ortiz-Vera, M., Rivera, D., Kloepper, J., Bonilla, R., 2013. Evaluation of three methods for preservation of Azotobacter chroococcum and Azotobacter vinelandii. Univ. Sci. 18 (2), 129–139.
- Sadoff, H.L., 1975. Encystment and germination in Azotobacter vinelandii. Bacteriol. Rev. 39 (4), 516.
- Schroth, M.N., Hancock, J.G., 1982. Disease-suppressive soil and root-colonizing bacteria. Science 216 (4553), 1376–1381.
- Setubal, J.C., dos Santos, P., Goldman, B.S., Ertesvåg, H., Espin, G., Rubio, L.M., Valla, S., Almeida, N.F., Balasubramanian, D., Cromes, L., Curatti, L., 2009. Genome sequence of *Azotobacter vinelandii*, an obligate aerobe specialized to support diverse anaerobic metabolic processes. J. Bacteriol. 191 (14), 4534–4545.
- Sharma, A., Johri, B.N., 2003. Growth promoting influence of siderophore-producing *Pseudomonas* strains GRP3A and PRS9 in maize (*Zea mays* L.) under iron limiting conditions. Microbiol. Res. 158 (3), 243.
- Shrivastava, R., Shrivastava, A.K., Dewangan, N., 2015. Combined application of Azotobacter and Urea to improve growth of rice (Oryza sativum). IOSR J. Environ. Sci. Toxicol. Food Technol. 1 (3), 67–72.
- Singh, R., Babu, S., Avasthe, R.K., YadavG, S., Chettri, T.K., Phempunadi, C.D., Chatterjee, T., 2015. Bacterial inoculation effect on soil biological properties, growth, grain yield, total phenolic and flavonoids contents of common buckwheat (*Fagopyrum esculentum* Moench) under hilly ecosystems of North East India. Afr. J. Microbiol. Res. 9 (15), 1110–1117.
- Socolofsky, M.D., Wyss, O., 1961. Cysts of Azotobacter. J. Bacteriol. 81 (6), 946.
- Spaepen, S., Vanderleyden, J., Remans, R., 2007. Indole-3-acetic acid in microbial and microorganism-plant signaling. FEMS Microbiol. Rev. 31 (4), 425–448.
- Subba Roa, N.S., 2001. An appraisal of biofertilizers in India. In: Kannaiyan, S. (Ed.), *The Biotechnology of Biofertilizers* (Doctoral Dissertation. Narosa Pub. House, New, p. 375.
- Tandon, H.L.S., 1991. Role of sulphur in plant nutrition. Fertil. News 36 (69), 79.
- Tchan, Y.T., New, P.B., 1984. Azotobacteraceae, 219-225. In: Krieg, N., Holt, J.G. (Eds.), Bergey's Manual of Systematic Bacteriology, vol. I. Williams & Wilkins Co., Baltimore.
- Tchan, Y.T., 1984. Genus 1. Azotobacter Beijerinck 1907, 567. In: Bergey's manual of systematic bacteriology, 1, pp. 220–229.
- Tchan, Y.T., Wyszomirska-Dreher, Z., New, P.B., Zhou, J.C., 1983. Taxonomy of the Azotobacteraceae determined by using Immunoelectrophoresis. Int. J. Syst. Evol. Microbiol. 33 (2), 147–156.
- Tejera, N., Lluch, C., Martinez-Toledo, M.V., Gonzalez-Lopez, J., 2005. Isolation and characterization of Azotobacter and Azospirillum strains from the sugarcane rhizosphere. Plant Soil 270 (1), 223–232.
- Thompson, J.P., Skerman, V.B.D., 1979. Azotobacteraceae: The Taxonomy and Ecology of the Aerobic Nitrogen-Fixing Bacteria. Academic Press Inc, (London) Ltd.
- Thompson, J.P., Skerman, V.B.D., 1981. Azorhizophilus, new genus. Azorhizophilus Paspali, pp. 215-218.
- Thompson, J.P., 1987. Cryopreservation of *Azotobacteraceae* in liquid nitrogen. MIRCEN J. Appl. Microbiol. Biotechnol. 3 (3), 323–336.
- Torres-Rubio, M.G., Valencia-Plata, S.A., Bernal-Castillo, J., Martínez-Nieto, P., 2000. Isolation of *Enterobacteria*, Azotobacter sp. and Pseudomonas sp., producers of indole-3-acetic acid and siderophores, from Colombian rice rhizosphere. Revista latinoamericana de microbiología 42 (4), 171–176.
- Ueda, T., Suga, Y., Yahiro, N., Matsuguchi, T., 1995. Remarkable N₂-fixing bacterial diversity detected in rice roots by molecular evolutionary analysis of nifH gene sequences. J. Bacteriol. 177 (5), 1414–1417.
- Vela, G.R., Wyss, O., 1965. Radiation resistance of soil Azotobacter. J. Bacteriol. 89 (5), 1280-1285.
- Vela, G.R., 1974. Survival of Azotobacter in dry soil. Appl. Microbiol. 28 (1), 77-79.
- Vikhe, P.S., 2014. Azotobacter species as a natural plant hormone synthesizer. Res. J. on Recent Sci. 3, 59–63 (IVC).

- 426
- Wani, S.A., Chand, S., Wani, M.A., Ramzan, M., Hakeem, K.R., 2016. Azotobacter chroococcum–a potential biofertilizer in agriculture: an overview. In: Soil Science: Agricultural and Environmental Prospectives. Springer, Cham, pp. 333–348.
- Widmer, F., Shaffer, B.T., Porteous, L.A., Seidler, R.J., 1999. Analysis of nifH gene pool complexity in soil and litter at a Douglas fir forest site in the Oregon Cascade Mountain Range. Appl. Environ. Microbiol. 65 (2), 374–380.
- Wilson, P.W., Knight, S.C., 1952. Experiments in Bacterial Physiology, vol. 49. Burguess, Minneapolis, USA.
- Winogradsky, S., 1938. Etudes sur la microbiologie du sol et des eaux. Annales de l'Institut Pasteur 60, 351–400 (Platenos. V, VI, and VII).
- Wyss, O., Neumann, M.G., Socolofsky, M.D., 1961. Development and germination of the *Azotobacter* cyst. J. Cell Biol. 10 (4), 555–565.
- Young, J.M., Park, D.C., 2007. Probable synonymy of the nitrogen-fixing genus *Azotobacter* and the genus Pseudomonas. Int. J. Syst. Evol. Microbiol. 57 (12), 2894–2901.
- Yousefi, A.A., Barzegar, A.R., 2014. Effect of Azotobacter and Pseudomonas bacteria inoculation on wheat yield under field condition. Intl. J. Agric. Crop Sci. 7 (9), 616.
- Zehr, J.P., Mellon, M., Braun, S., Litaker, W., Steppe, T., Paerl, H.W., 1995. Diversity of heterotrophic nitrogen fixation genes in a marine cyanobacterial mat. Appl. Environ. Microbiol. 61 (7), 2527–2532.
- Zulaika, E., Shovitri, M., Kuswitasary, N.D., 2014. Numerical taxonomy for detecting the *Azotobacter*ial diversity. In: The 8th Korean-Asean Joint Symposium on Biomass Utilization and Renewable Energy.

AS PER THE SYLLABUS OF BAJAJ COLLEGE OF SCIENCE (AUTONOMOUS) WARDHA

A TEXT BOOK OF CONCISE CHEMISTRY B.Sc. II

A Pragati Edition

- TEKADEMOHABANSIDADUREBANSINGE
- BARWAT
- BORKAR

SEMESTER

OTHER USEFUL BOOKS

A Text Book of Elements of Chemistry -Tekade, Mohabansi, Dadure, Bansinge, Barwat, Borkar A Text Book of Chemistry II -Pagariya, Padole, Chandak, Murhekar, Wagh, Thakare A Text Book of Chemistry IV -Dahikar, Dupare, Thakare, Banewar, Sangole, Gadpayale -Mandlik, Rathod, Rajput, Patil, Thakare A Text Book of Chemistry VI -S.K. Agarwal, Keemti Lal UGC Advanced Inorganic Chemistry -Indrajit Kumar Organometallic Compounds -S. Pimplapure Inorganic Polymer Chemistry -Pimplapure, Jain, Soni, Sahay **Coordination Chemistry** Jagdamba Singh, L.D.S. Yadav UGC Advanced Organic Chemistry Jagdamba Singh, L.D.S. Yadav **Organic Synthesis** Organic Polymer Chemistry -Jagdamba Singh, R.C. Dubey -Jain, Soni, Pimplapure Heterocyclic Compounds -Alka L. Gupta Polymer Chemistry --S. Pimplapure, Rashmi Jain Chemistry of Natural Products Organic Reaction Mechanism -Kusum Sharma -Kusum Sharma Designing Organic Synthesis A Text Book of Nanochemistry -Kusum Sharma Organic Spectroscopy Principles, Problems and Their Solutions -Jagdamba Singh, Jaya Singh Flow Through Nuclear Chemistry -Malik, Kumari, Sabharwal The Textbook of Stereochemistry -S. Agarwal, D. Jangid A Systematic Approach to Statistical Thermodynamics Vakil Poddar Solid State Chemistry -J.N. Gurtu, A. Gurtu Green Chemistry -J.N. Gurtu, Kusum Sharma UGC Advanced Physical Chemistry -J.N. Gurtu, A. Gurtu **Bio-physical Chemistry** -Alka L. Gupta Analytical Chemistry Alka L. Gupta Instrumental Methods of Chemical Analysis -H. Kaur An Introduction to Chromatography -H. Kaur Spectroscopy -H. Kaur

PRAGATI PRAKASHAN, MEERUT

Pragati Bhawan, 240, W.K. Road, Meerut.
 © 0121-2640642, 2643636, 4007643
 Support@pragatiprakashan.in
 www.pragatiprakashan.in



CONCISE OF SCIENCE (AUTOMONION OF

AND DESCRIPTION & AND DESCRIPTION OF AND ADDRESS OF ADD

Denne fin P. Hennel unel

Dr. M. M. Dation

TEV. Star 4.34

C Authors-A Text Book of Concise Chemistry-IV Sem

PRAGATI PRAKASHAN

Educational Publishers

Head Office : PRAGATI BHAWAN, 240, W. K. Road, Meerut-250 001 Phone : 0121-2643636, 2640642, 4007643 www.pragatiprakashan.in e-mail pragatiprakashan.in support@pragatiprakashan.in First Edition 2020

ISBN : 978-93-89961-30-0

Price : Rs. 230/-

Published by A.K. Mittal for Pragati Prakashan, Meerut - 250 001 and Photocomposing by : Pragati Prakashan, Meerut Printed at Arihant Electric Press, Meerut

Pragati's

BAJAJ COLLEGE OF SCIENCE (AUTONOMOUS) WARDHA

A TEXT BOOK OF **CONCISE CHEMISTRY**

(B.Sc. II Year, IV Semester Students)

an

Dr. P. V. Tekade M. Sc. B. Ed. SET. NET. GATE Ph. D.

> Dr. K. M. Dadure M. Sc. M.Phil Ph. D.

Mr. N. A. Barwat M. Sc. SET. NET

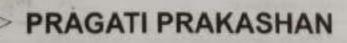
Dr. Mrs. N. P. Mohabansi M. Sc. B. Ed. SET Ph. D.

> Mr. M. D. Bansinge M. Sc. NET

Dr. P. G. Borkar M. Sc. SET, NET, GATE, Ph. D. Post Doc.

Department of Chemistry Bajaj College of Science, WARDHA

port sol. D. Bansinge



Contents

		Unit-I	
Α.	Chemistry of Lanthanides and	Actinides	
B.	Errors in Chemical Analysis		1-8
		Unit-II	9-25
A.	Coordination Compounds		-
B.	Oxidation and Reduction		26-72
		Unit-III	73-99
Ald	ehvdes and Ketones		100-115
	And the second s	Unit-IV	100.173
Α.	Carboxylic Acids		116-123
B.	Carboxylic acid derivatives		126-131
		Unit-V	
Α.	Adsorption		132-146
Β.	Catalysis		147-160
		Unit-VI	
Ele	ctrochemistry I		161-171
	gration		172-188
		Practical Course	
	Inorganic	Chemistry Experiments	189-196
Α.	Gravimetric Analysis		
	(i) Estimation of Ba2* as BaS	O ₀	
	(ii) Estimation Ni2 as Ni-DN	1G	
B.	Chromatographic separation	of binary mixtures (at least Two)	containing Cu(II)
	Co(II) and Ni (II) ions by paper	r chromatography and determinati	on of Rf values.
	Organic	Chemistry Experiments	197-208
0	rganic qualitative analysis	and the second	
	Preliminary examination		
	Element Detection		
	Functional group Detection		
	Preparation of derivative.		202.221
	Physica	l Chemistry Experiments	209-222
1	To verify the Freundlich adso	protion isotherm of acetic acid on a	charcoal.
2	To verify the Langemuide a de	auchion insthorm of acetic acid OR	CHarcoan
3	. To determine the strength	of the given strong acid conduct	tometrically using
	standard alkali colution		
4	. To determine the strength	of the given weak acid conduc	tometrically using
	Standard alkali calution		
	5. To determine the strength	of strong acid and a week acid i	n a given mixture
	COnductrimoterization to a star		
	6. To determine the solubility	and solubility product of a spa	ringly soluble sal
	CONLINE COMPANY IN	and solubility product of a spa	
	7. To determine the ionization of	onstant of weak acid conductomet	rically.
	and an and a state of the	unstant of weak acid conductomet	includy.

MODERN TEACHING OF ICT & CLIMATE CHANGE

NAMRATA SHARMA BHUMESH KUMAR GUPTA

Modern Teaching of ICT & Climate Change

This edited book contains chapters/articles contributed by participants of 121st Orientation Programme

Chief Editor Namrata Sharma, Bhumesh Kumar Gupta

RIGI PUBLICATION

All right reserved

No part of this book may be reproduced in any form, by Photostat, Microfilm, xerography, or any other means or incorporated into any information retrieval system, Electronic or Mechanical, Without the Written Permission of the copyright owner.

Modern Teaching of ICT & Climate Change

By

Namrata Sharma, Bhumesh Kumar Gupta

Copyright[©] Namrata Sharma, Bhumesh Kumar Gupta 2019 Originally published in India

ISBN: 978-93-88393-96-6 (Paperback) Published by RIGI PUBLICATION

777, Street no.9, Krishna Nagar Khanna-141401 (Punjab), India Website: www.rigipublication.com Email: info@rigipublication.com Phone: +91-9357710014, +91-9465468291

CONTENTS

1.	Environment And Sustainable Development Through Higher Education Author : Sudhir Ramchandra Tiple	7
2.	National Integration: A Debate Author : Ghode Rakesh Bharat	18
3.	Social Disparity In Indian Higher Education Evidence From All India Survey On Higher Education Author : Akarsh Arora	23
4.	Use Of ICT In Teaching Learning Process Author : Varsha Baswaraj Kharobe	38
5.	An Analytical Study Of Science Textbooks Developed By National Council Of Educational Research And Training, New Delhi For The Class VI To VIII With Reference To The Environmental Education Related Content Author : Shamim Aara Hussain	45
6.	Formative Assessment :Classroom Assessment Techniques Author : Y. Vijaya Lakshmi	62
7.	RUSA In Quality Enhancement Of Higher Education In Maharashtra State: Outcomes And Future Plan Author : Jaya Rambhau Somatkar	69
8.	Ict: An Advanced Techniques In Higher Education Author : Chandrashekhar Shalik Sutar	79
9,	Global Warming And Agriculture Sector Author : Ganesh Laxman Dhote	85

10.	Environment Conservation And Sustainable Development In India	91
	Author : Vinod Madhao Barde	
11.	Role Of Human Values In Higher Education Author : Virendra Balvir Turkar	98
12.	Environmental Issues Of Global Scenario Author : Shailesh Kashinath Bhagat	106
13.	Climate Change: Causes, Effect And Its Solutions Author : Sumit Dnyaneshwar Rokade	114
14.	Climate Change Due To Human Being And Role Of Higher Education Author : Yogesh Sureshrao Sakhare	122
15.	Preparation Of Nickel Zinc Ferrite Thin Films For Sustainable Development By Supercapacitor Application Author : Dattatraya Kisan Pawar	130
16.	Impact Of Global Warming And Climate Change On Biodiversity Author : Shailesh Shivdas Bhaisare	139
17.	Role Of Tet In Primary And Secondary Education Of Remote Area Author : Mahesh Kishanrao Kulkarni	147
18,	Emerging Trends In Teacher Education Author : Himanshu Ganeshbhat Acharya	158
19.	सूचना और संचार प्रौद्योगिकी आधारित शिक्षण Author : Vipin Kumar Singh	162

20.	Characteristics Of World-Class Universities Author : Robin Thomas	173
21.	ICT Based Teaching : Atriangular Relationship Between Teacher, Learner And Content Author : Vikas Bakshi	182
22.	Taxation As A Tool For Social Equality (Special Reference To Goods And Service Tax) Author : Anjali Pal	191
23.	Social Equality And Empowering Women Strengthen As A Accounting Profession: An Overview Author : Mahendra Kaithwas	199
24.	Outcome Based Education (Obe) With Special Reference To Indian Higher Education System Author : Jaya Kameriya	205
25.	Role Of Information And Communication Technology (ICT) In Higher Education Author : Rajesh Sethi	216
26.	Innovation In Teaching Methods Author : Jectendra Talreja	221
27.	A Study Of Strength, Weakness, Opportunity And Challenges (SWOC) Analysis Of Choice Based Credit System (CBCS) In Higher Education In India Author : Vijay Grewal	228
28.	ICT For Imparting Digital Education: Emerging Paradigm In Higher Education In India Author : Jyoti Yadav	235

ENVIRONMENT AND SUSTAINABLE DEVELOPMENT THROUGH HIGHER EDUCATION

Sudhir Ramchandra Tiple, Assistant Professor Jankidevi Bajaj College of Science, Wardha (M.S.) sudhir.tiple@gmail.com

Abstract:

Sustainable development has become an important concept for securing future of the world. Indian problems such as growing population, migration of people from rural area to urban area, increasing use of resources is endangering the passing of environmental benefits to next many generations to come. The higher education is the tool and solution for sustainable development in India as the awareness about the environmental hazards can be communicated to the lemans by the method of education. India or other — countries of world is working on sustainable development through higher education since 1972. This paper is to review the progressive development of environment and sustainable development through higher education. Many findings have been used to see the progressive development. It is suggested that the serious approach is to be made in order to preserve natural resources for future generation and education can be the best solution.

Keywords: Sustainable development, Higher education, Environment

1. Introduction:

Sustainable development has become an important concept for securing future of the world and India. (Sathaye, Shukla, and Ravindramath 2001) Indian pr4oblems such as growing population, migr_ation of people from ruml area to urban area, increasing use of resources are endangering the passing of environmental benefits to next many generations to come (Sathaye et al. 2001). Thus sustainable development is very quintessential need of today 's India. But it is very difficult to attain a sustainable development without considering the higher education. (Banga Chbokar 2010; Chbokar 2010; Dutz 2007) Higher education is the tool and solution for sustainable development in India as the awareness about the environmental hazards can be communicated to the lemans by the method of educating the one who belongs to the people and society most (Misra 2012; Srivastava and Rehman 2006) Hence it is essential to know about the education first and then the Sustainable develo pment will be discussed. Also the various initiatives by India and rest of world will be discussed. At the last the benefits and shortcoming of higher education in sustainable development will be characterized.

2. Education

Education in India is mostly categ orised in two different categories as Basic education and the Higher education.

a. Basic Education:

ⁿ standard in The basic education starts from the kinder garden to the 10 India. In India primary education is free as per the 86th Amendment of the Constitution of India. The amendment was made by then Prime Minister of India Shri. Atal Bihari Vajpeyi. As per this amendment primary education is . made free and compulsory for children of one age groups 6 to 14 years as their fundamental right under "Sarva Shiksha Abhiyan". (MHRD, 2015) (Kainth 2006; Rao 2009) In 1963, the NCERT major criterion was to make aware the student from their childhood age about the sensible use of natural resources. The syllabus consists of units like Air, Water and Weather, Rocks, Soils and Minerals, which clearly deals with environmental study. The Kothari commission (1964 -66) sugges ted that basic education should contain details of Environmental education and coordination of it with the life needs and desire of the people and the nation-(Agarwal 2007; Tilak 2018) The report recommended that the aims of teaching scien ce should build a proper understanding of the concepts, principles, facts, and procedure of the physical biological environment. The Review Committee (1975) recommended that there should be one book for environmental studies for classes III, IV and V (Agarwal 2007) The courses for environmental studies was designed in a way that it include both the natural and social environment. Thus a balance was made between one society and one environment. According to the National Curriculum Framework of 2005 (Research and (India) 2005) at the primary stage, EVS was given special importance.

b. Higher Education

The higher education consists of the education which students acquire during under graduation, post graduation, doctorate and post doctorate studies. The Supreme Court of India had acknowledged the significance of environmental education at all levels and it was made mandatory to have a course on Environment at the undergraduate level to sensitise the youth towards the environmental issues and concerns. As per the Supreme Court direction, the University Grants Commission introduced six months of compulsory environmental course in all the universities and colleges during the academic year 2004-05. (Rajagopulan 2015)

3. Environment and Sustainable Development

To find the solution we must first know what sustainable development actually is sustainable development? We will have to strike balance between the different types of needs of Indian people and one environment. The distribution should be in a way that the need does not a profound impact on one environment. Thus sustainable development can be defined as economic development that is conducted without depletion of the natural resources. In other words Sustainable development is development that meets the need s of the present, without upsetting the ability of future generations to meet their own needs. Sustainable development and social development go hand in hand hence proper linkage between Sustainable development and social development is needed. Sustainable development comprises of the need of the current generation and the future generation in an extent to which are environment can be used for the development of social development of current generation so that future generation will not get affected by the use of the environment. Education is a factor that can influence sustainable development in a broader extend. Higher Education can be a path for proper functioning of our proper development of human being along with conserving the same for the future gener ation. The sustainable development ideas were formulated in chronological order as follow(Kumar, Kumar, and Vivekadhish 2016; Misra 2012; United N ations 2016):

- In 1972 the first coin was tossed about sustainable development which was discussed in UN conference on human environment held at Stockholm.
- It was in 1987 a report "our common future" was published by UN world Commission on human environm ent and development. This report also called as Brundtland report was the first to define sustainable development

which says "meeting the need of present without compromising the ability of future generations to meet their own needs".

- Although the conference was held in 1972 it was in 1990 when universities of different parts of the world where requested by the International Association of Universities (IAU) to promote sustainable development by the method of Higher Education.
- In 1992 Brazil hosted UN Earth summit the main objective of the summit was to give a thought on the environmental problems and how to resolve by multidisciplinary approach mechanism.
- A major development took place in 1993 when Kyoto declaration on sustainable development was adopted by IAU.
- Hereafter there were several other meetings such as in year 2000, the chief of various countries gathered in UN to discuss about environmental concern.
- 7. The major improvement was done in 2002 on sustainable development in a conference held at Johanne sburg by declaring decade 2005 -2014 as decade for education for sustainable development. UNESCO was the agency which was looking after sustainable development and higher educationKyoto declaration (Ganapati and Liu 2009; Suthaye et al. 2001; Sotamaa 2009) was a major breakthrough to find the linkage between higher education and sustainable development. This declaration was mainly promising the following points:
- 1. The socio-economical benefits combined with eco-cultural benefits
- Accommodating sustainable development with the higher education and research
- 3. Suggest a different way of thinking
- 4. Development is directly related to the human race.
- To deliver welfare education as to create sensible citizen with respect to sustainable development.
- To form various collaborations between different institution for sustainable future

The Kyoto declaration had helped to encourage the rural population to know about sustainable development. It also wants local universities to provide information on sustainable development to the local rural people. Thus universities can help to remove the misunderstanding of the relation b etween

cultural myths and sustainable development. This will be helping local people to overcome the myths and superstition regarding the misuse of resources which are in great danger. The Kyoto declaration also realised that the available resources on earth should be categorised in different types depending upon the danger they are facing due to the over utilisation of resources. This data was shared with universities so that they can interpret and find solutions by the method of project and research wor k in higher education, thus Universities can create a method to overcome these problems. Thus Kyoto declaration has helped universities to research in sustainable development and find the reaction of their research on society. It is emphasising on consider ation of International Association of universities (IAU) and implementation of the declaration done by IAU. The IAU had emphasized on universities to work on environmental literacy that is to find a way to convey environment and sustainable development to public at large. These will develop relation and cooperation between the different segments of society and will increase collective handwork for safeguarding the natural resources for future generation.

The Kyoto declaration recommendation for universities is as follow:

- a. Each University will have their own Action Plan which will ensure that the institutions working under it will be committed to follow the principle and practices of sustainable development and in return it should communicate the same to its students, employee and public which is directly or indirectly attached to the instructions.
- It should not only educate others but also follow the sustainable development practices in its own campus.
- It should engage several teachers in their academic staff to teach sustainable development.
- d. It should encourage their staff and students to protect the environment and follow the procedures adopted by the University in accordance with the environmental and sustainable policies of the university and the institutions.
- It should try to implement interdisciplinary (intra -university) work for the functioning of program related with sustainable development.
- To improve ethical obligation of the stakeholders who are working on sustainable development.
- g. University students and staff should work hand in hand and they should overcome the forces which lead to environmental misuse.

- h. University can also promote Inter University collaborations for the betterment of the environment by utilising the best available facilities of each university.
- University can build a partnership with various other Institutions which are involved in developing innovative Technologies which will help in making environment better than previous.

4. Sustainable Development Goal (SDG)

The SDG (Le Blanc 2015; Bongaarts 2016; Rasul 2016; United Nations 2016)term was tossed when leaders of 193 countries came together in 2015 for the betterment of the future. After analysing the present scenario of the food shortage, water scarcity due to low rainfall, the unnecessary wars, poverty etc everywhere in the world. Hence to overcome this situation and to provide peoples around the world a better future, these countries created a plan called the Sustainable Development Goals (SDGs). This set of 17 goals is designed to get solution for poverty, health, hunger, etc and protect the adverse effects of climate change. Although it seems to be an adventurous plan but previous data of 15 years shows that due to the collective efforts of leaders around the word, the poverty has decreased to 50 percent in 2015 from 2000. Hence, all leaders around the world started to work on the SDGs one among them is United Nations Development Program (UNDP). UNDP is one of the leading organizations working to fulfill the SDGs by the year 2030. The 17 Goals as per their Goal Number are as follow:

1. No Poverty

This goal was set to end the extreme poverty by 2030. Although the world poverty was halved during 2000 to 2015 but as per survey around 80 crore peoples are still living in extreme poverty all over the word. The extreme poverty is considered when the livelihood of an individual is around Rs. 100 per day (\$1.25). Hence the goal is to overcome this by 2030.

2. Zero Hunger

This goal is set to end hunger by 2030 by confirming food security for each individual. Also, improvement of nutrition value and promotion of sustainable agriculture through organic farming is considered.

3. Good Health and Well-being

This goal is set to ensure the healthy lives to all classes and age of the human being.

4. Quality Education

The goal 4 is to bring equality in quality education. Everyone is liable for the learning throughout life. Hence every individual should have an environment for quality education throughout life.

5. Gender Equality

The goal is to achieve gender equality and empower the girls and women in every society around the globe.

6. Clean Water and Sanitation

The goal is to ensure sustainable development in water management and provide sanitation to each and every person of the world.

7. Affordable and Clean Energy

To ensure affordable, modern, reliable and sustainable energy to all is the primary target of this goal.

8. Decent Work and Economic Growth

This goal is set to promote sust anable economic growth for all that is everyone should get productive employment and decent work.

9. Industry, Innovation and Infrastructure

The goal is to build durable infrastructure, promote sustainable industrialization and innovation.

10. Reduced Inequality

The goal is reduce inequality within countries and support the developing countries and under develop countries to make up with the developed countries. 11. Sustainable Cities and Communities

The goal is to make cities sustainable for the safety and living values.

12. Responsible Consumption and Production

The goal is to make the utilization of any commodity in an equal manner. Hence emphasis is given on responsible consumption as well as responsible production.

13. Climate Action

There is a need of urgent action on the climate change to sustain the available resources for the future generations. The goal is set to take control on the fast accelerating climate change.

14. Life below Water

The sea is explored at a very high rate, hence the goal is set to conserve oceans. Also one goal is to ensure sustainably use the oceans, seas and marine resources.

15. Life on Land.

To protect, restore and encourage sustainable use of earthly ecosystems is one of part of this goal. It is also emphasizes on management of forests, prevent description, and the most important one is to halt and reverse land degradation and halt biodiversity losses.

16 Peace and Justice Strong Institutions

Peace is the need for sustainable development of the society and it can be achieved through the equality in justice delivery for all and equality in institutions at all levels

17. Partnerships to achieve the Goal

The last goal is to strengthen the means of implementation and refresh the global partnership for sustainable development.

5. India and SDG's

The Univ ersity ranking agency, Times Higher Education (THE) of United Kingdom released a major ranking to put various universities of the world in order of the extent of work done in the field of SDG. This ranking has considered the 11 SDGs which are having a direct impact of higher education on it. The SDGs which are considered to evaluate universities are Goal 3 to 5. Goal 8 to 13, Goal 16 and 17. The universities of Japan had the most representation in the world ranking list followed by the United States and Ru ssia. The rankings are given on the overall of 11 SDGs as well as individual SDGs. The list has been topped by different nations in different categories. But considering India with rest of the countries it is found that the contribution of India is very le-3.84 India is not standing even in top 100 of any of the 11 SDG (Kumar et al. 2016) categories. A joint report of NITI Aavog and United Nation was released in December 2018 which shows the SDG India Index for the Indian states and Union Territories in executing 2030 SDG goals. The report had emphasized on the critical progress of India o n SDG in the next decade as the population is increasing with an accelerated rate and also due to the high economic growth. Due to this reason it has become important to implement the SDG in a firm way. But the situation found in the ranking shows a very d ifferent picture, the institutions which are representing India on a global platform such as Central universities, NITs, IIMs and even IITs do not find any place in the the University Impact rankings . But on the other hand some private institutions have found their places in this ranking which are:

- 1. JSS Academy of Education and Research, TamilNadu
- 2. Amrita Vishwa Vidyapeetham
- 3. Annamalai University
- 4. Christ University, Bengaluru
- 5. Jamia Millia Islamia
- 6. KIIT University
- 7. KLE University
- 8. Manipal Academy of Higher Education
- 9. Pondicherry University
- 10. PSG College of Technology

This is a big question for the developing India whether our leading institutes are in reality accountable campuses that propagate sustainable development in higher education.

6. Conclusion

Sustainable development is a very important factor for the preservation of the environment for the future generation. This idea is to be percolated in the minds of the common people so that the excessive amount of utilization of the natural resources could be made un der control. Along with the environmental studies in basic education, higher education it should also be given a higher concern in environment and sustainable development. Higher education is the only way to create awareness about the sustainable developme – nt. International community had already considered higher education as an important facture in achieving Sustainable Development Goals (SDGs). But India is par away from progress done by the international community. Hence India needs to develop a strong policy for the environment and sustainable development through higher education.

References:

- Agarwal, Pawan. 2007. "From Kothari Commission to Pitroda Commission." Economic and Political Weekly 554–57.
- Le Blanc, D avid. 2015. "Towards Integration at Last? The Sustainable Development Goals as a Network of Targets." Sustainable Development 23(3):176–87.
- Bongaarts, John. 2016. "World Health Organization Health in 2015: From MDGs, Millennium Development Goals, to SDGs, Sustainable Development Goals." *Population and Development Review* 42(3):575–575.
- Chhokar, K. B. 2010. "Higher Education and Curriculum Innovation for Sustainable Development in India." *International Journal of Sustainability* in Higher Education 11(2):141–52.
- Dutz, Mark. 2007. Unleashing India's Innovation: Toward Sustainable and Inclusive Growth.
- Ganapati, Sukumar and Liguang Liu. 2009. "Sustainable Development in the Clean Development Mechanism: The Role of Designated National Authority in China and India. " Journal of Environmental Planning and Management 52(1):43–60.

- Kainth, Gursharan Singh. 2006. "A Mission Approach to Sarva Shiksha Abhiyan." Economic and Political Weekly 3288–91.
- Kumar, Sanjiv, Neeta Kumar, and Saxena Vivekadhish. 2016. "Millennium Development Goals (MDGS) to Sustainable Development Goals (SDGS): Addressing Unfinished Agenda and Strengthening Sustainable Development and Partnership." Indian Journal of Community Med icine 41(1):1.
- Misra, Harekrishna. 2012. "E-Governance and Millennium Development Goals: Sustainable Development Perspective in Rural India." Proceedings of the 6th International Conference on Theory and Practice of Electronic Governance.
- Rajagopalan, Ragh avachari. 2015. Environmental Studies: From Crisis to Cure. Oxford University Press.
- Rao, Vasanta Srinivasa. 2009. "Lack of Community Participation in the Sarva Shiksha Abhiyan: A Case Study." *Economic and Political Weekly* 61–64.
- Rasul, Golam. 2016. "Managing the Food, Water, and Energy Nexus for Achieving the Sustainable Development Goals in South Asia." *Environmental Development* 18:14–25.
- Research, National Council of Educational and Training (India). 2005. National Curriculum Framework 2005 . National Council of Educational Research and Training.
- Sathaye, Jayant, P. R. Shukla, and N. H. Ravindranath. 2001. "SPECIAL SECTION: CLIMATE CHANGE AND INDIA Climate Change, Sustainable Development and India: Global and National Concerns." *Current Science* 90(3).
- Sotamaa, Yrjö. 2009. "The Kyoto Design Declaration: Building a Sustainable Future." Design Issues 25(4):51–53.
- Srivastava, Leena and I. H. Rehman. 2006. "Energy for Sustainable Development in India: Linkages and Strategic Direction." *Energy Policy* 34(5):643–54.
- Tilak, Jandhyala B. G. 2018. "The Kothari Commission and Financing of Education." Pp. 255–82 in Education and Development in India. Springer.
- United Nations. 2016. "Sustainable Development Goals: Metadata." Sastainable Development Goals (March):1–526.

FOLDSCOPE AND ITS APPLICATIONS



CHIEF EDITOR DR. ARUN DEV SHARMA

Foldscope and its Applications

EDITOR-IN-CHIEF Dr. Arun Dev Sharma

ISBN No: 978-93-85835-68-1



Published By: National Press Associates, New Delhi

Foldscope and its Applications

EDITOR-IN-CHIEF

Dr. Arun Dev Sharma

Editors

Dr.GayatriGurjar Dr.ChTulasiMastanamma Dr.Indu Sharma Dr.ShobhaAjeetWaghmode Dr. SG Kulkarni Dr.BharathiPrakash Dr.DharmeshHarwani Dr.Mousmisaikia Dr.Mahipal Singh Shekhawat Dr.Anupmaharshal W Dr. KG Sabarinathan Dr M. Gomathy Dr KG Sabarinathan

© 2019. National Press Associates, New Delhi

All rights reserved 2019. No part of this book may be reproduced or Transmitted in any form or by any means of electronic or mechanical including photocopy, recording or any information stored in a retrieval system, without the prior written permission of the publisher.

ISBN No: 978-93-85835-68-1

Price: 800/-

The Responsibility for the facts or opinions expressed in the papers are entirely of the Authors. Neither the College nor the Publisher is responsible for the same.

Printed in India

.

National Press Associates

Admin Office: C-24, Ground Floor, PanchsheelVihar, Malviya Nagar, New Delhi-110017, India Regional Office: #79, Guru AngadDev Nagar, Flower Enclave, Ludhiana (PB), India. Branch Office: C-104, AnuroopSoceity, Vartak Nagar, Thane (West)-400606, Maharashtra, India Email:info@npajournals.orgwww.npajournals.org

PREFACE

A Foldscope, a low-cost science tool, is an optical microscope that can be assembled from simple components, including a sheet of paper and a lens. It was developed by Dr. Manu Prakash and designed to cost less than US\$1 to build. It is part of the "frugal science" movement which aims to make cheap and easy tools available for scientific use in the developing world. The Department of Biotechnology (DBT), Government of India and the Prakash Lab at Stanford University, USA signed an agreement to bring the Foldscope to India to encourage curiosity in science. It is being used as a teaching tool for the students in biology, chemistry, physics and many other streams. Keeping these facts in the background, the editors and authors of the book have tried to compile their research and review outlook about Foldscope usage and its various applications. The aim of this book is to facilitate the adoption of Foldscope as an educational and research tool by students, teachers, scholars, scientists and the general people. Many authors who are also Project Investigators and recipients of the Foldscope research grant acknowledge Department of Biotechnology, Government of India. The authors hope that this book will not only provide pleasant reading but also practical knowledge which can be utilized by the user of this book in the area of Foldscope microscopy.

EDITOR IN CHIEF

Dr Arun Dev Sharma

FISCA,FPASc, FSAWR, HOD, Associate Prof, PG Dept of Biotechnology, Lyallpur Khalsa College, Jalandhar, 144001, Punjab, India

CONTENTS

APPLICATION OF THE FOLDSCOPE TO IDENTIFY, CHARACTERIZE INDIGENO SPECIES AND GENERATE A DNA BARCODE DATABASE	US
Anupma Harshal W, Sagarika Vivek Damle, Sharon K., Anand Sharma	1
FOLDSCOPE: AN EFFICIENT PORTABLE TOOL FOR PLANT BIOLOGISTS, EDUC AND SOCIETY	CATION
Arun Dev Sharma, Priya Nischal	12
PUBLIC PARTICIPATION IN MITIGATING WATER BORNE DISEASES USING FOLDSCOPE AS A TOOL IN TRIBAL REGION OF DAKSHINA KANNADA Bharathi Prakash, G D Khedkar, S P Jeevan, K.E. Prakash, Vaniprabhu	18
FOLDSCOPE AS A STUDY TOOL FOR POLLEN GRAINS	
Ch. Raghumani Singh, Y. Sunitibala Devi	24
FOLDSCOPE: AN EDUCATIONAL CUM RESEARCH TOOL USED IN IDENTIFICAT MICROORGANISMS FROM WASTE WATER	FION OF
Dharmendra Rathod	29
FOLDSCOPE VIEW OF FLORAL AND FAUNAL DIVERSITY IN THE THAR DESER RAJASTHAN	T OF
DharmeshHarwani, JyotsnaBegani, SwetaBarupal, JyotiLakhani	32
FOLDSCOPE AS A TOOL TO CREATE AWARENESS ABOUT HYGIENE AND INVOLVEMENT OF MICROBES AMONG STREET FOOD VENDORS AND CONSUM Tulasi Mastanamma, Hima Bindu, Sunitha Das	/IERS 39
FOLDSCOPES AS USER-FRIENDLY TOOL ON BONE MARROW STEM CELLS Dr. A. Mangala Gowri	48
SCREENING FOR THE PRESENCE OF MICROBES IN VEGETABLE AND WATER SAMPLES FROM TWO DISTRICTS OF MANIPUR, INDIA, USING FOLDSCOPE Pukhrambam Grihanjali Devi, Indira Yumnam, N. Romabati Devi	55
STUDY OF ZEBRAFISH EMBRYOGENESIS USING FOLDSCOPE Gayathri N., Priti Dubey, NitinWasnik	60
A GLIMPSE INTO THE PLANT WORLD THROUGH FOLDSCOPE Madhavi G Kanade, Gayatri S Gurjar	65
FOLDSCOPE AND PHYLLOSPHERE MICROORGANISMS Gomathy, M, Sabarinathan, K.G, Subramaniyam, K.S, Kalaiyarasi, VandJeyshree M	76
CERVICAL CANCER &IT'S DIAGNOSIS BY FOLDSCOPE Hari Lakshmi Chikkala, K. Vijaya Rachel	82

.

BIOFILM PRODUCING ORGANISMS AND THEIR ANTIBIOTIC RESISTANCE: A FOLDSCOPE APPROACH	
Indu Sharma, Gayatri Gogoi, Parijat Hazarika	87
FOLDSCOPE AS A RESEARCH TOOL AND ITS APPLICATIONS Jayateertha R Diwan, KashappaChikkanaragund	95
STANDARDIZATION OF FOLDSCOPE IN COMPARISON WITH A MICROSCOPE TO IDENTIFY E. COLI FOR FURTHER USE IN FIELD SETTINGS	0
Jeyakumar Angeline, Swapnil Godbharle, Bibek Raj Giri, Juana Hatwik	101
FOLDSCOPE: A PRIMARY TOOL FOR DETECTION OF BIOACTIVE COMPOUNDS PLANT CELLS	
Kuldeep D. Shekhaliya, Jigna G. Tank, Rohan V. Pandya	105
STUDY OF MORPHOLOGY, HISTOLOGY, POWDER CHARACTERISTIC OF SOME BY USING FOLDSCOPE AS A RESEARCH TOOL	
Juvatkar PV, Dr. Kale M. K., Khan N1, Gorde N, Waghulde S, Gokhale S	117
FOLDSCOPE AS A TOOL TO SCREEN PARASITIC INFECTIONS IN WILD ANIMAL TAMIL NADU	
M.Palanivelrajan, K.Manoj Dhanraj, C. Sreekumar, K. Senthilkumar	128
FOLIAR MICROMORPHOLOGICAL ANALYSIS OF OXALIS CORNICULATA L. USIN FOLDSCOPE	
Mahipal S. Shekhawat, Manokari M., Priyadharashini S., Cokul Raj M., Kannan N	135
AN EXPERIMENTAL INVESTIGATION ON THE QUALITY OF AIR AND GROWTH MICROORGANISM IN VARIOUS CONSTRUCTION MATERIALS AT VARIOUS CONDITIONS AND IN VARIOUS COOKING VESSELS	OF
E. Muthu Kumaran, Manikannan Mathayan	143
VERSATILITY OF THE PAPER MICROSCOPE: FOLDSCOPE, MIRACLE MICROSCO Moirangthem Bhubaneshwari Devi, Dhananjoy Singh Chingangbam	OPE 147
MICROSCOPIC CHARACTERIZATION OF BOTANICALS HAVING ANTI-MALARIA PROPERTIES USED BY THE TRIBAL COMMUNITIES OF ASSAM	AL
Pranab Borah and Mousmi Saikia	154
A FOLDSCOPIC STUDY OF ARSENIC RESISTANCE BACTERIA IN GROUNDWATE SAMPLES OF LAKHIMPUR, ASSAM (INDIA)	
Mridul Buragohain	163
FOLDSCOPE: LOW COST PAPER MICROSCOPE FOR ENVIRONMENTAL MONITO Munmi Gogoi, Dr. Dilip Saikia	DRING 169
CAPTURING DIVERSE, IMPRESSIVE IMAGES USING FOLDSCOPE Prarthana.J, Narayana	175

.

FOLDSCOPE: A NEW AGE EXPLORATORY EDUCATIONAL TOOL	
Lalit Mohan, Keshav Goyal, Shaubhik Anand, Muskan Mittal, Shrabani Snigdha, Tavleen Ba	jwa,
Kusum R Gupta, Rakesh Kumar Gupta and Prerna Diwan	188
EXPLORATION OF ARBUSCULAR MYCORRHIZAL FUNGI (AMF) IN THE RHIZOS OF FIVE VARIETIES OF <i>COCOS NUCIFERA</i> L. DURING SUMMER SEASON IN POL TALUK, COIMBATORE DISTRICT, TAMILNADU, INDIA	
Nivedha. D, R. Gokilavani and H. Rehanabanu	194
A COMPARATIVE STUDY ON USE OF FOLDSCOPE AND COMPOUND FOLDSCOP BIOLOGICAL SAMPLE THE OBSERVATION	
S. T. V. Raghavamma	201
FOLDSCOPE: AN INNOVATIVE TOOL TO STUDY AGRICULTURALLY IMPORTAN MICROBES.	T
Sabarinathan K. G, M.Gomathy, D. Arun Kumar and R. Kannan	206
FOLDSCOPE: A VERSATILE TOOL TO STUDY THE PUPPET MASTERS OF RHIZOSPHERIC AND AQUATIC MICROBIOME	212
Satish V. Patil ,Bhavana V. Mohite	212
DETECTION AND DIAGNOSIS OF FISH PATHOGENS USING FOLDSCOPE ON FRESHWATER FISHES IN THE WATER BODIES OF THE CHIRANG DISTRICT OF ASSAM, INDIA	
Sewali Pathak	219
DIAGNOSIS OF NATURE'S KIDNEY WITH THE AID OF FOLDSCOPE	
ShamimRahman	225
GREEN SYNTHETIC APPROACH CORESHELL NANOMATERIALS: AS AN ANTIMICROBIAL AGENT.	
ShobhaA.Waghmode	231
MAPPING MICROSCOPIC BIODIVERSITY IN AQUATIC ECOSYSTEMS USING PAI MICROSCOPE 'FOLDSCOPE'	PER
Maya Murdeshwar, SujataDeshpande, Siddhi Parab, Jennifer Tellis	238
MICROSTRUCTURAL EXAMINATION OF WASTE E- WASTE REINFORCED POLY MATRIX COMPOSITE BY NOVEL MICROSCOPY	MER
Swanand Gajanan Kulkarni	252

.

Dhananjaya Pratap Singh Vijai Kumar Gupta • Ratna Prabha *Editors*

Microbial Interventions in Agriculture and Environment

Volume 1 : Research Trends, Priorities and Prospects



Dhananjaya Pratap Singh Vijai Kumar Gupta • Ratna Prabha Editors

Microbial Interventions in Agriculture and Environment

Volume 1 : Research Trends, Priorities and Prospects



Contents

1	50 Years of Development of Beneficial Microbes for SustainableAgriculture and Society: Progress and Challenges Stillto Be Met—Part of the Solution to Global Warmingand "Hothouse Earth"1Gary E. Harman
2	Metabolomics Approaches in Microbial Research:Current Knowledge and Perspective Towardthe Understanding of Microbe PlasticityPaulo R. Ribeiro, Rhaissa R. Barbosa, and Catherine P. de Almeida
3	Is PGPR an Alternative for NPK Fertilizers in SustainableAgriculture?51Éva Laslo and Gyöngyvér Mara51
4	Soil: Microbial Cell Factory for Assortment with BeneficialRole in Agriculture63Pratiksha Singh, Rajesh Kumar Singh, Mohini Prabha Singh,Qi Qi Song, Manoj K. Solanki, Li-Tao Yang, and Yang-Rui Li
5	Insights into the Unidentified Microbiome: CurrentApproaches and Implications93Ratna Prabha, Dhananjaya Pratap Singh, and Vijai Kumar Gupta
6	Interactions in Soil-Microbe-Plant System: Adaptationto Stressed Agriculture131Stefan Shilev, Hassan Azaizeh, Nikolay Vassilev, Danail Georgiev,and Ivelina Babrikova
7	Microbe-Mediated Tolerance in Plants Against Biotic and Abiotic Stresses

8	Arbuscular Mycorrhizal Colonization and Activation of Plant Defense Responses Against Phytopathogens
9	Microbes as Resource of Biomass, Bioenergy, and Biofuel
10	Microbe-Mediated Reclamation of Contaminated Soils: Current Status and Future Perspectives
11	Plant Growth-Promoting Rhizobacteria (PGPR)and Fungi (PGPF): Potential Biological Control Agentsof Diseases and Pests.281Pankaj Prakash Verma, Rahul Mahadev Shelake, Suvendu Das,Parul Sharma, and Jae-Yean Kim
12	Biofortification: A Promising Approach Toward Eradication of Hidden Hunger
13	Microbes in Foods and Feed Sector
14	New Age Agricultural Bioinputs
15	Microbial Bio-production of Proteins and Valuable Metabolites 381 Abiya Johnson, Prajkata Deshmukh, Shubhangi Kaushik, and Vimal Sharma
16	2,4-Diacetylphloroglucinol: A Novel Biotech Bioactive Compound for Agriculture. 419 Raksha Ajay Kankariya, Ambalal Babulal Chaudhari, Pavankumar M. Gavit, and Navin Dharmaji Dandi
17	Coral Reef Microbiota and Its Role in Marine Ecosystem Sustainability

х



New Age Agricultural Bioinputs

14

Bhavana V. Mohite, Sunil H. Koli, Hemant P. Borase, Jamatsing D. Rajput, Chandrakant P. Narkhede, Vikas S. Patil, and Satish V. Patil

14.1 Introduction

Nitrogen-based biofertilizers are significant bioinputs, but according to current environmental changes and ever-increasing food demand, it is the need of time to popularize more efficient bioinputs for soil. These bioinputs will help to fight against problems like an unpredictable monsoon, global warming, and decreasing soil fertility, and indiscriminate use of agrochemicals.

Besides chemical fertilizers, organic soil conditioners, the application of phosphate solubilizers, nitrogen fixers, and *Trichoderma*, *Verticillium*, *Metarhizium* like versatile biocontrolling agents are the common strategies of soil conditioning. In the past 50 years, there is tremendous work published on nitrogen fixers and phosphate solubilizers. The results of these findings directed to the exploitation of common biofertilizers like *Azotobacter* and *Rhizobium* as a nitrogen fixer and other organic inputs. In addition to above, phosphate, zinc, sulphur, potassium solubilizers are a

H. P. Borase

V. S. Patil

S. V. Patil (🖂)

North Maharashtra Microbial Culture Collection Centre (NMCC), Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India

© Springer Nature Singapore Pte Ltd. 2019

B. V. Mohite · S. H. Koli · J. D. Rajput · C. P. Narkhede

School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India

School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India

C. G. Bhakta Institute of Biotechnology, Uka Tarsadia University, Surat, Gujarat, India

University Institute of Chemical Technology, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India

School of Life Sciences, Kavayitri Bahinabai Chaudhari North Maharashtra University, Jalgaon, Maharashtra, India

D. P. Singh et al. (eds.), *Microbial Interventions in Agriculture and Environment*, https://doi.org/10.1007/978-981-13-8391-5_14

significant part of current agricultural practices. Although these practices proved beneficial to uphold soil fertility and other agronomical problems like pest attack and plant susceptibility to various infections, physiological problems due to the change in the atmosphere need some novel strategies or additional bioinputs.

There are various significant bioinputs like the application of 1-aminocyclopropane-1-carboxylic acid (ACC) enzyme and phytase producing microorganisms and bacterivorous flora. These are which were reported, but unfortunately remain as neglected practices by Indian farmers. The following three major bioinputs are need of time to use as new soil bioinputs in modern agricultural practices:

- 1. Use of ACC oxidase and deaminase producer bioinputs
- 2. Use of phytase producer
- 3. Use of bacterivorous soil microbes

The central idea of this chapter is presented in Fig. 14.1, which represents the ability of major modern agricultural bioinputs.

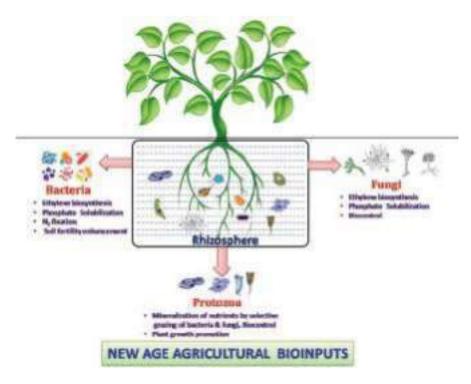


Fig. 14.1 Schematic representation for the new age agricultural bioinputs

14.2 Application of ACC Oxidase and Deaminase Producer Bioinputs

14.2.1 ACC and ACC-Degrading Enzymes

The Yang cycle produces 1-aminocyclopropane-1-carboxylic acid (ACC) and ACC oxidase and deaminase (ACCO and ACCD) (Yang and Hoffman 1984). Shang Yang unlocked the mystery of freshness of fruit, flowers, defoliation, and ripening of fruits by proposing a continuous biochemical cycle known as the Yang cycle. The Yang cycle biosynthesizes ethylene in plants. Ethylene is important in host–pathogen interactions, seed germination, flowering, and fruit ripening. It establishes the central role of methionine in ethylene synthesis. Yang's study proved the genesis of S-adenosylmethionine as a transitional compound which is further converted into ACC and then ethylene (Fig. 14.2).

ACC is the signaling molecule of a plant, easily transported through intra- and intracellular tissues over short and long distances.

ACC is a cyclic α -amino acid with a three-membered cyclopropane ring merged to an α -carbon atom of the amino acid (Fig. 14.3) and chemical formula C₄H₇NO₂ with a molar mass of 101.0 g/mol⁻¹. ACC is considered an essential intermediate that regulates ethylene biosynthesis. The enzyme ACCO is a member of the oxidoreductase class, which is responsible for the transformation of

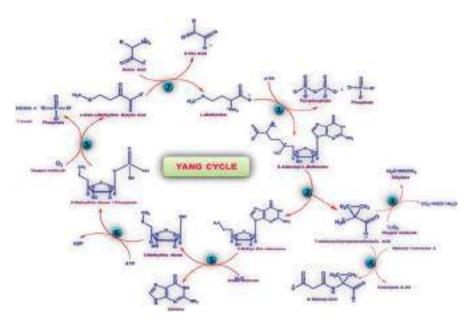


Fig. 14.2 Yang cycle for ethylene biosynthesis. Cycle path: (1) SAM synthetase, (2) ACC synthase, (3) ACC oxidase, (4) ACC N-malonyltransferase, (5) MTA nucleosidase, (6) MTR kinase, and (7) transaminase, (S) spontaneous reaction

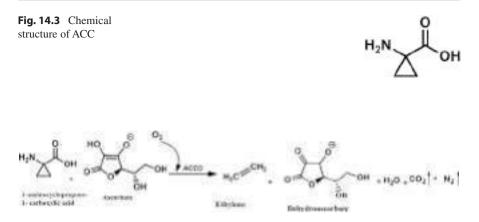


Fig. 14.4a Transformation of ACC to ethylene with ACCO

1-aminocyclopropane-1-carboxylate to ethylene with carbon dioxide, water, and other by-products (Fig. 14.4a).

In drought stress conditions, ethylene synthesis is rapidly increased (Morgan and Drew 1997). Ethylene is the one of the marker compounds of drought conditions and is also known as stress ethylene. Nitrogen fixation and nodulations are influenced by the various effects of high ethylene synthesis through water and temperature stress, like reduction of transpiration rate by closing stomata to regulate the abscisic acid pathway (Tanaka et al. 2005; Tamimi and Timko 2003; Penmetsa and Cook 1997; Guinel 2015). Hence, if the ACCO is regulated, then the natural synthesis of ethylene is regulated. Various researchers advocated that various rhizospheric microbes also control the ethylene level in a plant by deaminating ACC diffused through root cells and seeds (Finlayson et al. 1991; Penrose and Glick 2001; Penrose and Glick 2003).

14.2.2 Aminocyclopropane-1-Carboxylic Acid Oxidase (ACCO)

Aminocyclopropane-1-carboxylic acid oxidase is an enzyme recognized to fight against the consequences of drought in plants. It was well documented that drought affects various biochemical, morphological, and physiological activities of plants, e.g., turgor pressure, transport of soil nutrients, nutrient transport to root, nutrient diffusion through root mass, and a run of water-soluble nutrients such as silicon, manganese, and sulphate. Besides these, it leads to oxidative stress, which causes a decrease in chlorophyll synthesis, membrane deterioration, and protein degradation in plants (Hsiao 2000; Selvakumar et al. 2012; Sgherri et al. 2000; Rahdari et al. 2012).

14.2.3 Aminocyclopropane-1-Carboxylic Acid Deaminase (ACCD)

ACCD is the enzyme synthesized in the cytoplasm of bacteria. It is a multimeric sulfhydryl enzyme having a monomeric subunit with molecular weight of 35–42 KD (Glick et al. 2007). ACCD catalyses ACC conversion and produces α-ketoglutaric acid and ammonia (Fig. 14.4b). It was reported that D-serine and D-cysteine (D-amino acids) also act as a substrate for ACCD. Previously, the optimum temperature and pH for ACC deaminase were reported as 30–35 °C and 8.5 (Jacobson et al. 1994; Honma and Shimomura 1978; Jia et al. 1999). But currently, there is significant research going on to screen a versatile ACC deaminase producer who has a broad temperature and pH range (Xuguang et al. 2018). Various bacteria were reported for the production of ACCD, e.g., *Enterobacter cloacae, Pseudomonas putida, Pseudomonas* sp., *Alcaligenes, Hansenula, Rhizobium, Sinorhizobium* sp., *Pseudomonas chlororaphis, Rhizobium leguminosarum*, and *Bacillus subtilis* (Klee et al. 1991; Glick 1995; Belimov et al. 2007; Tittabutr et al. 2013; Ma et al. 2004; Duan et al. 2009). Similarly, some fungi and yeast were also reported for ACCD production, e.g., *Penicillium citrinum* (Minami et al. 1998; Jia et al. 1999).

Glick (1995) described the role and importance of some plant growth-enhancing Rhizobacterium in the management of drought pressure and various physiological activities of plants. Glick (1995) illustrated that ACC is produced in more quantity during drought stress and exudated outside of the root cells. The plant growthinducing bacteria around the roots are recognized for its versatile activity and utilize the ACC exudate by ACC deaminase, and to keep the balance in internal and external ACC level, internal ACC is transported outside of the root. This process reduces the amount of ACC required for the biosynthesis of ethylene inside plant cells. Hence, if such ACCD-producing Rhizobacterium is present around the rhizospheric area of vegetation in a drought condition, ethylene production is suppressed, further leading to restrain inhibitory stress; ethylene causes defoliation, inhibition of root elongations, and nodulation transpiration (Glick et al. 2007). The presence of ACCD-producing microbes in soil proved their significance in a variety of plant growth-promoting activities, e.g., the existence of ACCD producer enhances the nitrogen fixations by inducing the normal process of root nodule organization in drought or temperature stress conditions.

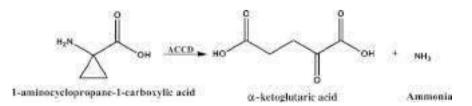


Fig. 14.4b Conversion of ACC to ethylene with ACCD

14.3 Application of Phytase Producer

14.3.1 Importance of Phosphorous

Phosphorous (P) is the next main macronutrient required for plant growth after nitrogen. It accounts for about 0.2% of dry weight of a plant. It makes vital biomolecules like nucleic acids, ATP, and phospholipids, and ultimately plant growth is inhibited without the supply of this nutrient. It also has a role in the regulation of the metabolic pathway and enzyme-catalyzed reactions. Phosphate affects germination and seed maturity and eventually plant development. Plant development comprising of root, stem, and stalk is dependent on phosphate. Phosphate has a role in the formation of seed and flower, which ultimately has an effect on crop development and yield (Khan et al. 2009). It has a remarkable function in N fixation in legumes, energy metabolism, membrane synthesis, photosynthesis, respiration, enzyme regulation, crop value, and abiotic and biotic stress resistance. No atmospheric source of phosphate could be made available to plants (Ezawa et al. 2002), and soils normally contain trace quantities of available phosphate (predominantly as HPO₄²⁻ and $H_2PO_4^{-}$) that is readily available for plant uptake. Phosphate addition in the soil in the form of fertilizers fulfills the plant requirement (Richardson et al. 2009). The unavailability of phosphate in soluble form is a vital factor (Xiao et al. 2011) that restricts the agricultural production worldwide (Ramaekers et al. 2010). Both organic and inorganic phosphate accumulate in soil and consequently not available for plant consumption. Inorganic phosphate is fused through chemical adsorption and precipitation, while immobilization of organic phosphate occurs in soil organic matter (Sharma et al. 2012).

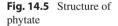
Even phosphatic fertilizers fail due to their conversion to an insoluble form like calcium phosphate and aluminum phosphate (>70%) (Mittal et al. 2008). Phosphate is available in low quantity in soil (1.0 mg kg⁻¹ soil); additionally, it becomes unavailable by reacting with reactive metals like Al³⁺ in acidic, calcareous, or normal soils (Gyaneshwar et al. 2002; Hao et al. 2002). Crop plants can, therefore, make use of only a little bit of phosphorus, which eventually results in reduced crop performance (Reddy et al. 2002). The high percentage of an insoluble type of phosphate leads to eutrophication, while frequent use of phosphate causes soil infertility and rapid depletion of nonrenewable phosphate reserves. The outcome of this event would be the lake's biological death i.e. cyanobacterial blooms, hypoxia, and death of aquatic animals due to depleted bioavailable oxygen and buildup of nitrous oxide. (Vats et al. 2005). In the plant, a range of morphological and physiological changes was observed due to deficiency in phosphate, which consecutively affects plant growth, productivity, and survival (Tran et al. 2010), and hence are a significant pin down for the agriculture industry worldwide.

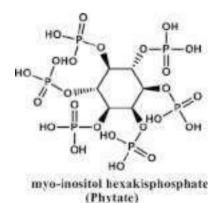
Hence, effective phosphorous utilization is crucial for the sustainable expansion and prevention of undesirable environmental effects (Scholz et al. 2015). The translation of a phytate–phosphate compound in the soil in crop accessible orthophosphate would mitigate phosphate-related obstacles.

14.3.2 What Is Phytate?

Phytate is a significant storage compound of phosphorus in seeds. Eighty percent of the total seed phosphorus is made by phytate, which accounts for 1.5% of seed dry weight (Raboy and Dickinson 1987). The myo-inositol hexakisphosphate is a phosphate salt of myo-inositol having all six hydroxyl groups substituted by phosphate residues (Fig. 14.5). The myo-inositol 1,2,3,4,5,6-hexakis (dihydrogen) phosphate is commonly called myo-inositol hexakisphosphate, or phytate, which is a collection of the organic form of phosphorus compounds found widely in nature. The prefix "hexakis" designates that the phosphates are not internally connected and the compound is formed by a polydentate ligand, which binds with more than one metal atom coordination site. Each phosphate group is in ester form within an inositol ring and binds entirely with 12 protons (Bohn et al. 2008; Cao et al. 2007).

Phytate usually presents as a salt of monovalent and divalent cations (Fe²⁺, Mn²⁺, K^+ , Mg^{2+} , and Ca^{2+}) and formed in seeds at the stage of ripening. In phytic acid, the negatively charged phosphate sturdily binds with positively charged metallic cations resulting in an insoluble complex and restricting the accessibility of nutrients. Phytic acid and its derivatives are accountable for various cellular events such as signaling, RNA export, endocytosis, DNA repair, and vesicular cell trafficking (Bohn et al. 2008; Frias et al. 2003). In plants, phytate is the prime storage type of inositol phosphate. The plant root has 30% phosphorus fractions, while seeds and cereal grains have 80% phosphorus (Lott et al. 2000; Turner et al. 2002; Haefner et al. 2005). Two pathways are considered for the biosynthesis of phytate: lipiddependent and lipid-independent. The synthesis of phytic acid starts from myoinositol via a series of phosphorylation steps. In the former route, phytate is attained by the successive phosphorylation of Ins(1,4,5)P3 (inositol 1,4,5-triphosphate) and Ins(1,3,4)P3 (inositol 1,3,4-triphosphate). The subsequent compound is released from PtdIns(4,5)P2 (phosphatidylinositol 4,5-biphosphate) by the effect of a specific phospholipase C. The intracellular location of the intermediates of phytic acid biosynthesis is not fully explored.





Organic phosphate in rhizosphere has a high affinity to soil particles by precipitation and adsorption and hence it creates deprived accessibility to the plant as it cannot be desorbed (Menezes-Blackburn et al. 2013). Phytic acid is degraded in seed germination by a precise assembly of enzymes called phytases.

14.3.3 Phytase Enzyme

Phosphorus deficiency results from the phytase secretion of a variety of plant roots (Minggang et al. 1997). The distinct phosphatases phytases (myo-inositol hexakisphosphate phosphohydrolase) sequentially hydrolyze the phosphomonoester bonds from phytic acid, thereby liberating lower inositol phosphates and inorganic phosphate (Singh et al. 2011). These catalysts commence phytic dephosphorylation at various positions on the inositol ring, and it produces diverse isomers of lower inositol phosphates (Turk et al. 2000).

14.3.4 Structure and Mechanism of Action of Phytase

Phytase (myo-inositol hexakisphosphate phosphohydrolase) is a homodimaeric enzyme (EC 3.1.3.26 and EC 3.1.3.8) (Hegeman and Grabau 2001; Guimarães et al. 2004). Phytases carried out the subsequent release of inorganic phosphorus from phytic acid. Phytases act hydrolytically to break the phosphate ester bond of phytate and release inositol phosphates and phosphorus with other essential nutrients, which are required for plant absorption (Angel et al. 2002) (Fig. 14.6). Phytases are involved in the dephosphorylation of inositol-6-phosphate and high-order inositol hexakisphosphate hydrolyze sequentially to form lower-order esters like inositol monoesters (Hayes et al. 1999; Vats and Banerjee 2004). The inositol penta- and hexakisphosphate (phytate) hydrolyzing enzymes are of interest because they constitute a high percentage of the whole organic phosphate (Turner et al. 2002).

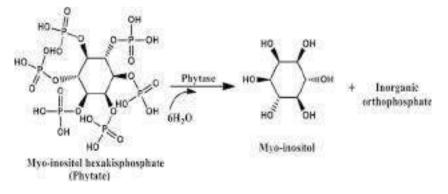


Fig. 14.6 Phytase action on phytate

The phytase protein has substrate binding and catalyzation conserved domains. The substrate binding domain is present at the N-terminal with RHGxRxP conserved sequence for substrate binding. The C-terminal catalyzation theme comprises of particular HD components. The "pocket" structure is framed by the connection of residues in the motif (Mullaney et al. 2000). The substrate restricting site with RHGxRxP arrangement responds with the substrate and frames the chemical substrate complex. The phosphate groups are then released from the substrate by the HD element (Li et al. 2010).

Phytate hydrolysis occurs in two stages: the nucleophilic attack and protonation. The histidine in the dynamic site of the catalyst caused a nucleophilic assault to the fragile phosphoester bond of phytate and caused the protonation by the aspartic acid of the leaving cluster (Li et al. 2010). The β -propeller alkaline phytases lack the RHGXRXP sequence motif, and hence it needs calcium thermostability as well as enzyme activity to produce the IP3 (inositol triphosphate) (Kim et al. 1998a; Mullaney and Ullah 2003).

Phosphatases cause hydrolysis of 60% of the total organic phosphate. The highest quantity of phosphate was released by phytases from phytate (Bünemann 2008). The release of orthophosphate from soil natural phosphate is effective in microbes as well as in plants. Plant phytases have been distinguished in roots and root exudates during the early stage of seed germination; they frequently show a poor action, making them inefficient for hydrolyzing soil phytic acid as well as phosphorous usage (Hayes et al. 1999; Richardson et al. 2009) and thus suggest that the microbial catalyst demonstrates superior, effective liberation of phosphorous (Tarafdar et al. 2001).

14.3.5 Categorization of Phytases

Phytases are assembled by their enzyme action, pH action, and the initiation site of dephosphorylation of phytate. They are categorized into 3-phytases (EC 3.1.3.8), 5-phytases (EC 3.1.3.72), and 6-phytases (EC 3.1.3.26) on account of the initial hydrolysis position of phytate according to IUPAC-IUBMB (Bohn et al. 2008), which were subsequently alienated into alkaline and acid phytases (Jorquera et al. 2008). The three-dimensional structure and catalytic mechanism cause classification into four classes: histidine acid phytases (HAP) (EC 3.1.3.2), cysteine phytase or purple acid phosphatase (PAP) (EC 3.1.3.2), beta-propeller phytase (BPP) (EC 3.1.3.8), and protein tyrosine phosphatase (PTP)-like phytases (Li et al. 2010), which have recently been characterized (Lei et al. 2007). HAPs and BPPs are the most well-known and contemplated phytases. Various bacterial, fungal, and plant phytases have a place with the HAP family, while BPP has all the earmarks of being the prevalent phytase in *Bacillus* species (Greiner et al. 2007; Huang et al. 2009). These two most important categories have a different catalytic activity that results in distinct end products. While HAPs catalyze the hydrolysis of PA in myo-inositol and Pi, BPP activity results in the creation of the inositol-triphosphates – either Ins(1,3,5)P3 or Ins(2,4, 6)P3 (Greiner et al. 2007; Kerovuo et al. 2000).

As per the optimum pH, acid phytases, for the most part, incorporate HAP, PAP, and PTP-like phytases, though alkaline phytases include just BPPs from *Bacillus* species (Singh and Satyanarayana 2015; Tye et al. 2002). Alternatively, carbon position of dephosphorylation initiation resulted in phytases grouping into 3-phytase (myo-inositol hexakisphosphate 3-phosphohydrolase), 6-phytase (myo-inositol hexakisphosphate 5-phosphohydrolase).

The categorization of phytase into EC 3.1.3.8, EC 3.1.3.26, and EC 3.1.3.72 (myo-inositol-hexaphosphate phosphohydrolases) was organized on the background of protein sequencing, and successive dephosphorylation (George et al. 2007) of P occurs at three and six positions, correspondingly. The labeling basis is the three- and six-bond position of myo-inositol 6-phosphate. The 3-phytases (EC 3.1.3.8) are present in filamentous fungi like *Aspergillus* sp. and 6-phytases (EC 3.1.3.26) are found in plants, e.g., wheat.

14.3.6 Reserve of Phytase

Phytases can be formed by microorganisms, plants, and animals. Wheat, rice, soybeans, barley, peas, corn, and spinach are examples of plant sources. Microorganisms like bacteria, fungi, and yeast are the real source of phytase found in the blood of vertebrates such as fish and reptiles (Gupta et al. 2015; Bohn et al. 2008). Among the phytases from microorganisms, attention is focused on *Aspergillus* sp. because of its high production and extracellular activity (Gupta et al. 2015). To circumvent this obstacle the sole strategy is the application of phytases which hydrolyze the phytate and increase availability of P to plants. Commercially available phytase addition is costly and time-consuming, and hence the maintenance of rhizospheric phytase producer is important. Another engineering approach involves incorporation of genes behind phytase production from microbes into transgenic plants. However, there is a range of constraints for phytase engineered crop plants like loss of seed viability, yield, vulnerability for ecological pressure, and rejection of genetically modified organisms (GMOs) (Reddy et al. 2017).

14.3.7 Microorganisms Producing Phytase

Phytases of microbial origins are of rigorous significance among plants, animals, and microorganisms owing to the ease of genetic manipulation and large-scale production (Adhya et al. 2015). Microorganisms are the key drivers in the soil, which regulates phytate mineralization. The occurrence of microorganisms in soil rhizo-sphere may balance plants inability to procure P directly from phytate. In microorganisms, bacteria, yeast, and fungi have been effectively researched for extracellular phytase action (Pandey et al. 2001). A single phytase cannot address the issues of business and ecological applications (Bakthavatchalu et al. 2013). Microbial

phytases are investigated mainly from fungi of a filamentous type such as Aspergillus ficuum (Gibson 1987), Mucor piriformis (Howson and Davis 1983), Aspergillus fumigatus (Pasamontes et al. 1997), Cladosporium sp. (Quan et al. 2004), and Rhizopus oligosporus (Casey and Walsh 2004). Phytase production by different bacteria has been described, viz., *Bacillus* sp. (Kim et al. 1998b; Choi et al. 2001), Citrobacter braakii (Kim et al. 2003), Pseudomonas sp. (Richardson & Hadobas 1997), Escherichia coli (Greiner et al. 1993), Raoultella sp. (Sajidan et al. 2004), and Enterobacter (Yoon et al. 1996). The anaerobic rumen bacteria, mainly Selenomonas ruminantium, Prevotella sp., Megasphaera elsdenii, and Mitsuokella multiacidus (Richardson et al. 2001b) and Mitsuokella jalaludinii (Lan et al. 2002), have also been investigated for phytases. The γ -proteobacteria group possesses the phytase production potential among the majority of soil bacteria. Fungi have extracellular phytases, while bacteria produce cell-linked phytases. Bacillus (Choi et al. 2001; Kerovuo et al. 1998; Kim et al. 1998a; Powar and Jagannathan 1982; Shimizu 1992) and *Enterobacter* (Yoon et al. 1996) are the only bacterial genera having extracellular phytase activity. The phytase activity of Selenomonas ruminantium and Mitsuokella multiacidus (D'Silva et al. 2000) is outer membrane linked, while Escherichia coli produces the periplasmic phytase enzyme (Greiner et al. 1993).

B. subtilis is as a competent of phytase producer owing to its nonpathogenic and safe nature for industrial-level phytase production. This microorganism has numerous additional advantageous properties like organic acid production and antibiosis for phosphate solubilization in the soil. Currently, *Aspergillus* and *E. coli* are the commercial phytase producers. Among the various organisms reported, the inhabitant *E. coli* enzyme demonstrates the maximum phytase activity.

Phytases from bacterial sources are a genuine option in contrast to fungal enzymes because of their specificity to the substrate, protection from proteolysis, and effective catalytic action (Konietzny and Greiner 2004). Bacillus phytases are exceptionally effective due to its higher thermal stability and neutral pH. The Bacillus phytase has stringent specificity for a substrate for the calcium-phytate complex effective for application in the environment (Farhat et al. 2008; Fu et al. 2008). Nevertheless, owing to inefficient enzyme production methods for *Bacillus* sp., it could not be produced at commercial scale as only a few strains have been significantly commercialized for phytase production (Zamudio et al. 2001). Lactobacillus sanfranciscensis is the main sourdough lactic acid bacteria that demonstrated a significant level of phytate degrading action (De Angelis et al. 2003). The HAP are specifically produced from Aspergillus sp. like A. terreus, A. ficuum, and A. niger (Wyss et al. 1999), while the alkaline phytases are produced from Bacillus amyloliquefaciens (Idriss et al. 2002) and Bacillus subtilis (Kerovuo et al. 2000). Escobin-Mopera et al. (2012) had purified phytase from Klebsiella pneumoniae 9-3B. Rhizobacteria can mineralize phytate and may enhance P uptake of plants in soils (Patel et al. 2010). A better and substitute resource of phytase is continuously searched by screening new organisms that may produce novel and effective phytases. The ultimate aim is to produce phytase cost-effectively with optimized conditions for industrial application.

14.3.8 Why Do Bacteria Produce Phytase?

Bacterial phytase production is an inducible complex regulatory mechanism. Phytase synthesis control is different in various bacteria. Phytase production is not a condition for balanced bacterial growth, but it is the response to an energy or nutrient constraint. Phytase formation takes place when bacterial cells face environmental variations prior to the commencement of growth or when actively growing culture faces a stressful condition. The metabolic regulation by signal transduction is also a mechanistic role (Zamudio et al. 2002).

14.3.9 Parameters Affecting the Activity of Phytases

The soil environment presents extreme difficulties like denaturation, degradation, adsorption, and dilution to extracellular chemicals (Wallenstein and Burns 2011). The constancy of extracellular and intracellular enzymes is variable. Stability is portrayed more in extracellular than intracellular proteins and is credited by glycosylating disulfide bonds that alter thermal soundness, an expansive pH scope of action, and some protection from proteases. Some are stabilized by binding with humic substances and clay minerals (Quiquampoix and Burns 2007). Biological and physicochemical procedures influence phytase action. The former causes changes in enzyme creation rates leading to isoenzyme generation and changes in microbial network synthesis, while the latter causes changes in absorption desorption responses, substrate dissemination rates, and enzyme degradation rates (Wallenstein et al. 2009). Essential elements influence the action of enzyme include the amount and kind of substrate (Fitriatin et al. 2008), type of solvent, pH, temperature, the existence of an inhibitor and activator, the quantity of the enzyme, and the reaction product (Sarapatka 2002).

14.3.9.1 Effect of Substrate on Phytase Action

Phytase action shifts with various substrates. The different substrates include 1-naphthyl phosphate, 2-glycerolphosphate, glucose-6-phosphate (Escobin-Mopera et al. 2012), 2-glycerolphosphate, fructose-6-phosphate, calcium phytate, sodium phosphate. phytate, p-nitrophenyl phosphate, ß-glycerol adenosine-5'guanosine-5'-triphosphate (GTP), monophosphate (AMP), adenosine-5'diphosphate (ADP), adenosine-5'-triphosphate (ATP), and nicotinamide adenine dinucleotide phosphate (NADP) (Farouk et al. 2012; Bakthavatchalu et al. 2013). Phytases are categorized as substrate particular and nonparticular acid phosphatases (Rossolini et al. 1998; Rodríguez and Fraga 1999).

14.3.9.2 Effect of pH on Phytase Action

The activity of phytases relies on the pH and temperature. Plant phytases have less pH and thermal stability than microbial phytases. The optimum pH for phytase activity is 5.0–8.0, hence classified as acid or alkaline phytases, respectively (Konietzny and Greiner 2002). The optimum pH for fungal phytases is 4.5–6.5 with

80% activity; for example, *Rhizoctonia* sp. and *F. verticillioides* have an optimum pH of 4.0 and 5.0, respectively (Marlida et al. 2010). The optimum pH for bacterial phytases is 6.0–8.0 (Kerovuo et al. 1998; Kim et al. 1998a). Acidic phytases have an optimum pH range from 4.5 to 6.0 (Konietzny and Greiner 2002), and pH 8.0 is the optimum for alkaline phytases in legume seeds (Scott 1991), lily pollen (Baldi et al. 1988), and cattail (Kara et al. 1985; Scott 1991).

14.3.9.3 Effect of Temperature on Phytase Action

Temperature is the most indispensable factor of enzyme action, influencing both enzyme generation and degradation rates by microorganism. The ideal temperature of phytate-degrading enzyme fluctuates from 35 to 77 °C. Predominantly plant phytases have the greatest action at lower temperature compared to microbial phytases (Konietzny and Greiner 2002). The ideal temperature for plant phytases ranges from 45 to 60 °C (Johnson et al. 2010). In general, metabolic rate of enzyme producing life forms increases with temperature over the range 5–40 °C. In this way, temperature supposes a more vital job in the rate of extracellular enzyme activity when contrasted with enzyme kinetics itself.

14.3.9.4 Effect of Soil Type on Phytase Action

The action of phytase in soil is additionally influenced by physicochemical properties of the soil, which incorporates soil compose, organic matter content, nitrogen content, C/N proportion, and aggregate P content (Djordjevic et al. 2003). The soil performance of phytase fluctuates with soil compose, and the movement of phytase lost expeditiously is dependent on three differentiating soil nature. The initial fate of phytase is confined by adsorption in the soil. The degradation and magnitude of phytase adsorbed continue as before for a wide range of soil arrangements. The highest adsorption was recorded at low pH, and it becomes nearly equivalent to zero when pH is adjusted to 7.5. The adsorption bestows defense to phytase degradation in the soil, but also limits loss of enzyme activity in the adsorbed state.

14.3.10 Mechanism of Phytase Activity

Microorganisms can enhance the capacity of a plant to acquire P through various mechanisms, and the important one is phytase like enzyme production (Richardson and Simpson 2011). The purified crystalline form of phytase has different catalytic properties with specific diverse mechanisms. The principal action of all portrayed phytases depends on the enzymatic hydrolysis of the bonds among inositol and phosphoric acid deposits. Enzymatic hydrolysis of bonds happens among inositol and phosphoric acid deposits whereupon the component of activity of all phytases is based. The results of this arrangement of responses are six-fold alcohol and phosphates (Mukhametzyanova et al. 2012). Microbial phytases decay fresh plant build-ups in the soil prompting the release of phosphorus from organic compounds. There are various arrangements alongside differing rates of responses by which the phosphoric acid deposits are discharged through microbial hydrolysis of phytate

(Mukhametzyanova et al. 2012). The histidine acidic phytases catalyze the release of phosphates in neighboring free hydroxyl group, after the dephosphorylation of a first phosphate group. For the most part, plant phytases display a difference in transitional myo-inositol pentaphosphate development among the first phase of the response. In the course of the first venture of hydrolysis, microbial 6-phytases frame a different set of intermediates. The acid phosphatases with phytate hydrolyzing properties hydrolyze glucose-1-phosphate in *Enterobacteriaceae* (Greiner and Sajidan 2008). Alkaline phosphatases in lily pollen, *B. subtilis*, and reed mace formed myo-inositol triphosphates as end products (Greiner et al. 2007; Greiner and Sajidan 2008; Mukhametzyanova et al. 2012).

14.3.11 Importance of Microbes for Phosphorous Mobility with Phytase

Soil microorganisms, particularly the higher plant rhizosphere, are exceptionally powerful in discharging P from natural pools of aggregate soil P by mineralization and inorganic complexes through solubilization (Hayat et al. 2010).

Mineralization results from the transformation of organic P, for example, phytate to plant-accessible inorganic P, by microorganisms through their expressed enzyme phytase (Ariza et al. 2013). Phytases have been recognized in roots and root exudates in plants (Li et al. 1997; Hayes et al. 2000; Richardson et al. 2000). Despite the fact that it is accounted for the enzymatic action in root exudates, it is not sufficient for efficient use of natural phosphorous (Brinch-Pedersen et al. 2002; Richardson et al. 2000). The addition of exogenous phytase into the media resulted in phytate availability for plant growth (Hayes et al. 2000; Idriss et al. 2002; Unno et al. 2005). The addition of exogenous phytase (Idriss et al. 2002; Richardson et al. 2001b; Singh and Satyanarayana 2010; Hayes et al. 2001a; Li et al. 2007a, b, 2009) resulted in growth of plant with phytate as solitary source of phosphate. The current research is targeted on the genetic expression of phytase genes in the plant for organic P utilization from the soil. The graphic demonstration of the function of microorganisms in phosphate solubilization is described in Fig. 14.7.

The action of plant phytases comprises just a little extent of the aggregate phosphatase reaction and is viewed as insufficient for guaranteeing adequate phosphate securing (Richardson et al. 2000; Findenegg and Nelemans 1993; Hayes et al. 2000). Bacterial phytases are effective for growth and yield of the plant. The limitation of plants to extort P from soil phytate could be overcome by treatment with phytate-degrading bacteria, like biofertilizer. Microbial phytase plays a very important role for the availability and mobility of phosphorous in soil because of its agronomic and ecological value for the growth of the plant as suggested by the recent scientific research. The long-term phosphorous deprivation in plants could be met by phytase from microorganisms; hence, the use of microbial phytase on an industrial scale is very appealing nowadays (Jorquera et al. 2008). The fungal extracellular phytase-treated seeds support the plant phosphorus nutrition in high phytate

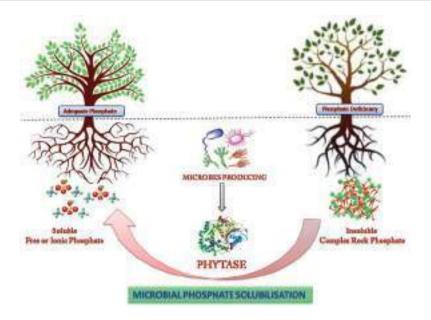


Fig. 14.7 Role of phytase from microorganisms in phosphate solubilization

content soil (Tarafdar 1995). The enrichment of soil with phytase from bacteria like *B. amyloliquefaciens* and *Bacillus mucilaginosus* advances the development of corn and tobacco, respectively (Li et al. 2007a, b; Idriss et al. 2002). Phytases from bacteria also release the vital soil micronutrients by phytate chelation and make it available to the plant. The purified microbial phytase or phytase-producing microbial strains could be functional as an effective and eco-friendly way to increase bioavailable soil phosphorus and limit the wide utilization of inorganic phosphate fertilizers.

14.3.11.1 Transgenic Plants for Phytase

Gene for phytase from a microorganism is integrated into plants like tobacco with a phyA gene from *A. niger* constituting phytase as soluble proteins in tobacco seeds. Genetically modified plants produce extracellular phytase from roots, which showed significant improvement in P nutrition in the soil, with higher phytate content or artificially modified for phytate (George et al. 2004, 2005). Thus the phytase from a microorganism is the critical element, and their existence in the rhizosphere helps the plant to recover from its inability to use the unavailable phytate.

Phytases have developed to be a valuable key to supportable agribusiness. It gives an approach to stop the revenue costs that turn out to be superfluously high because of the expansion of phosphorus manures. Broad research on phytase utilizing biotechnological applications will unquestionably give efficient arrangements towards practical agribusiness and ecological insurance in the coming years.

14.4 Use of Bacterivorous Microbes from Soil

14.4.1 Bacterivorous Protozoan

It was an accepted truth that soil microbes provide essential functions supporting soil fruitfulness and plant well-being. Recent evolution in molecular techniques like molecular sequencing resulted in a boom in studies of various microflora like an insect, animal gut, lakes, ponds, and terrestrial flora. However, all these studies cover bacteria and fungi only and neglect other trophic levels. But most attempts to use these bacteria and fungi as bioinputs in natural soil have been reported unsuccessful.

For the past 50 years the terms "biofertilizer" and "PGPR bacteria" only represent nitrogen fixer and phosphate and growth hormone producer. However, the truth is there is still no confirmation that these added bioinputs sustain soil fertility. The accepted truth is that these fungal and bacterial bioinputs have significant selective pressures of predation and not resource availability. These predators are bacterivorous and fungivorous protist. Protists massively consume bacteria as well as other soil microbes like fungi and yeast, and unicellular algae and release various micronutrients, growth-promoting substances, and different assimilable nitrogenous compounds and mineral (Ekelund and Rønn 1994).

Although various soil protozoans and nematodes are reported for their bacterivorous role, very few reports exist discussing the function of protozoans in the development of crop plant or soil richness (Bonkowski and Brandt 2002; Bonkowski 2004). The size of most soil protozoan ranges from 10 to 100 µm in diameter, but their weight is negligible. It was assumed that the biomass of total protozoan in soil is equal to the biomass of all other clusters of soil animals together except earthworm (Schaefer and Schauermann 1990; Schröteret al. 2003). In the biological energy coordination, the soil organic cycle plays an important role, which involves anabolic and catabolic steps of energy investment and energy escape or lost. Protozoans are major engineers which motion this organic energy cycle in the soil. Protozoa drive this cycle continuously where there is sufficient water available like moisture-containing intersoil capillaries, pore spaces, and fissures. Besides these, protozoans account for significant respiration of soil. It was noted that they contribute to 15-70% of the entire soil respiration. These indicate that protozoans are a vital component of the soil. The soil protozoans majorly include ciliates, flagellates, and naked and testate amoebae (Fig. 14.3). Although these protozoans have an extensive array of food assimilation and enzyme syntheses like a higher animal, they are not capable of synthesizing some vitamins and cofactors, and hence they depend on some microbial population for it.

Ciliates are one of the group including protozoan, which are identified for its extraordinary bacterivorous capacity (Sherr et al. 1987); owing to their large size. Algae, fungi, and small animals are foods for these ciliates (Bernard and Rassoulzadegan 1990; First et al. 2012). They have various habitats like freely swimming in the water, crawling on surfaces, and physically attached to surfaces by very flexible spring-like stalk, e.g., *Paramecium, Euplotes*, and *Vorticella* (James

and Hall 1995). There are some ciliates, which have special cilia for swimming and hairs for predation known as membranelle, which help for catching massive bacteria or prey in food vacuoles. Ciliate feeding rates are very high; it was recorded that single ciliates can digest 1254 bacteria h^{-1} (Iriberri et al. 1995).

Flagellates are another member of protozoans bearing one or more flagella having a different size from 2 to 20 μ m. They are versatile in nature like swimming freely or attaching to solid surfaces by trailing flagellum or stalks. Flagellates using these flagella either create feeding current or exploit it to put the water and prey in the oral furrow and at the base of the flagellum where the pseudopodia ingest the prey. Flagellates show selective grazing as per their size. They prefer smaller-size organisms as significant prey. It was reported that bacteria are more susceptible to flagellate grazing than other microbes having size >2.4 μ m. Chrzanowski and Šimek (1990) reported that flagellate bacterial grazing rate varies from 2 to 300 bacteria h⁻¹ (Davis and Sieburth 1984; Eccleston-Parry and Leadbeater 1994a).

Amoebas are widely occurring protozoans and are very normal in water, soil, and other habitats. They are abundant in the soil, i.e., $103-107 \text{ g}^{-1}$ of dry soil, with varying size <10 µm. Amoebas play a very important function in the cycling of various minerals and minute supplements such as nitrogen and phosphorus, particularly in shallow levels of nutrient environments (Goldman et al. 1985; Eccleston-Parry and Leadbeater 1994b). Amoebae, ciliates, and flagellates together selectively nurture on bacteria and control bacterial soil population (Table 14.1). They act as an essential constituent of the "microbial loop" (Azam et al. 1983). They are well recognized as Rhizopoda amoebae because they use their cytoplasmic protrusions, i.e., pseudopodia, for locomotion and nourishment. Amoebae are of two types, naked amoebae and shelled amoebae (testate amoebae).

Naked amoebae have no perfect shape but show three major morphological forms, i.e., floating, active form with extended lobose; fan-shaped, slug-like pseudopodial form trophozoites; and smaller and dormant form called cyst, an unusual rounded form (Page 1988; Griffiths 1970). Typical examples of naked amoeba are *Amoeba*, *Acanthamoeba*, *Vannella*, and *Vampyrella*.

Testate amoebae secrete the siliceous shell around the body. These testate are species-specific architectures. The testate shell amoebae designate the nutritional category of the living environment. The aperture is at one side of a shell, which is used for feeding or catching of different preys (Jassey et al. 2012). The dominant victims of amoebae are bacteria; the intake rate of the amoebic cell was reported to be 0.2-1465 bacteria h⁻¹ (Heaton et al. 2001; Huws et al. 2005).

14.4.2 Role of Protozoans as New Bioinputs

Various studies indicated that protozoans majorly preyed upon bacteria. Bacteria, unicellular fungi, yeast, algae, and cyanobacteria were assumed as a nutritional capsule. In addition to nitrogen and carbon sources, these nutritional capsules are enriched with micro- and macronutrients in addition to various growth factors (Table 14.2). It was formerly confirmed that the nitrogen and carbon content of a

Types	Example	Bacterivorous capacity (bacterial cell h ⁻¹)	References	
Amoeba				
Naked	Saccamoeba	0.2–1465	Heaton et al. (2001) and Huws et al. (2005)	
	Acanthamoeba			
	Euglypha cristata			
	Hartmannella			
	Cf. Mayorella			
	Cf. Polychaos			
Shelled	Vannella			
	Vampyrella			
	Arcellinid testate			
	Euglypha cristata			
	Arcella gibbosa			
	Difflugia			
	Foraminifera			
	Nebela			
Flagellates	Giardia intestinalis	2	Davis and Sieburth (1984 and Eccleston-Parry and Leadbeater (1994a)	
	Peltomonas hanelisp. nov.			
	Apusomonas australiensis sp.			
	Cetcomonar crassicauda			
Ciliates	Paramecium	20-1254	Iriberri et al. (1995)	
	Vorticella			
	Balantidium coli			
	Oxytricha trifallax			
	Stentor roeselii			

 Table 14.1
 Bacterivorous capacity of various protozoans

 Table 14.2
 Elemental composition of bacteria and fungi

Element	Bacteria (% dry weight)	Fungi (% dry weight)
Carbon	50–53	40-63
Hydrogen	7	_
Nitrogen	12–15	7–10
Phosphorus	2.0–3.0	0.4–4.5
Sulphur	0.2–1.0	0.1–0.5
Potassium	1.0-4.5	0.2–2.5
Sodium	0.5–1.0	0.02–0.5
Calcium	0.01-1.1	0.1–1.4
Magnesium	0.1–0.5	0.1–0.5
Chloride	0.5	_
Iron	0.02–0.2	0.1–0.2
References	Luria (1960)	Lilly (1965)
	Aiba et al. (1973)	Aiba et al. (1973)
	Herbert (1956)	

fungal and bacterial cell are 10-15% and 50-63% by dry weight of fungi and bacteria, respectively. Similarly, bacterial and fungal mass sufficiently contain valuable micronutrients such as phosphate, potassium, sulphur, calcium, and iron (Luria 1960; Herbert 1956; Aiba et al. 1973). All protozoans are well characterized for their enormous feeding habits on other microbes such as bacteria and other microbes. Different soil bacterial flora assimilated the atmospheric nitrogen with organic and inorganic matters from the soil and locked in their cells, which are not freely accessible for the plants. The enormous grazing activity remobilized this immobilized nitrogen and released ammonia, which is ultimately utilized by the plant (Goldman and Caron 1985). Griffith and Bardget (1997) proved that the nitrogen requirement of protozoans is comparatively less, and they make about 60% of ingested nitrogen available to plants in the form of ammonia. Hence after the ingestion of bacteria by a protozoan, nitrogen is not only released but also various nutrients like 50-63%carbon, 2.0-4.5% phosphorus, and 0.02-0.5% iron (Table 14.3). Bonkowski (2004) reported the essential function of protozoa in sustaining soil productiveness and plant health.

Protozoa provide all essential nutrients by mineralizing complex material in bacteria during feeding. They also control the structure and activity of bacterial loops of soil and root-associated communities (Sieburth and Davis 1982; Bonkowski and Brandt 2002). Krome et al. (2010) reported that selective predation of bacteria promotes the production of various plant growth hormones. Besides offering different mineralized nutrients, it was proved that protozoans also increased the nutrient assimilation rate by altering the root morphology. Bonkowski and Brandt (2002) reported that when the *Acanthamoeba castellanii* was inoculated in the rhizosphere, it induces the extensive fibrous and fine root, suggesting that protozoans play an important role like plant growth hormones (Krome et al. 2010). Jousset et al. (2010) also proved that protozoans not only stimulate growth but also play a noteworthy function in pathogen suppressions by encouraging other bacterial soil flora for antibiotics like chemicals. Similarly, it induces iron chelating organic molecule production, which makes iron unavailable for plant pathogen growth and multiplication (Levrat 1989; Mazzola et al. 2009; Müller et al. 2013; Mellano et al. 1970).

Nielsen et al. (2002) proved that bacteria such as *Pseudomonas* and *Bacillus* produce various antipathogenic compounds such as phenazines, DAPG (diacetyl phloroglucinol), and cyclic lipopeptides like tensin, amphisin, and viscosinamide, but Mazzola et al. (2009), Jousset and Bonkowski (2010), and Weidner et al. (2017) revealed that protozoan grazing pressure induced the making of such antipathogenic

Sr. no.	Bacterivorous organism	Phosphatase (IU/h)	ACC deaminase activity (μ M of α -ketoglutarate/mg/h)	Tryptophan (µg/h)
1	Acanthamoeba sp.	16.20	0.161	15
2	Paramecium sp.	18.40	0.093	17
3	Amoeba sp.	11.20	0.218	11
4	<i>Tetrahymena</i> sp.	14.00	0.187	07

Table 14.3 Performance of protozoans for phosphatases, ACCD, and tryptophan

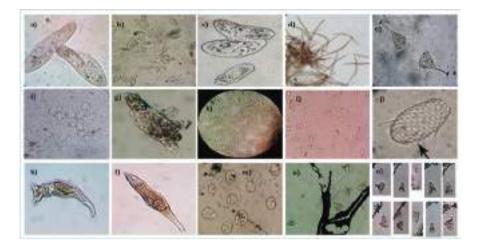


Fig. 14.8 Bacterivorous animals of soil cultured at School of Life Sciences, KBC NMU laboratory (**a**-**c**) *Paramecium* sp., (**d**) *Spirostomum* sp., (**e**) *Suctoria* sp., (**f**, **g**) *Acanthamoeba* sp., (**h**, **i**) cyst of amoebae, (**j**) testate amoebae, (**k**, **l**) Rotifer, (**m**) *Actinosphaerium* sp., (**n**, **o**) *Vorticella* sp.

fungal and bacterial compound. Recently in our laboratory studies at KBC North Maharashtra University (KBC NMU), Jalgaon, we have isolated and cultured various important agricultural bacterivorous animals, viz., *Paramecium, Amoeba*, Rotifer, and Vorticella (Fig. 14.8). It was revealed that *Acanthamoeba castellanii*, *Paramecium caudatum, Spirostomum*, and *Amoeba* spp. have the potential to produce various enzymes like phytase, phosphatase, and ACC deaminase. All these enzymes previously assumed the essential character of plant growth–promoting bacteria (Zahir et al. 2004). In laboratory-grown culture studies, it was discovered that *Paramecium* and *Acanthamoeba* efficiently utilized ACC and phytate and phosphate. Similarly, *Suctoria* sp. and *Spirostomum* were also investigated to use phosphate, phytic acid, and ACC like substrate at low concentrations (Table 14.3). *Amoeba* sp., *Acanthamoeba*, and *Paramecium* sp. were also found to be the producer of metabolic products such as amino acids like tryptophan, which was previously reported for a vital role in the stimulation of auxin production (Krome et al. 2010).

Sayre (1973) reported the potential of *Amoebae* as a future potent nematicidal agent. At KBC NMU laboratory, the cultured *Amoebae* sp. was also established to have an extraordinary potential of controlling invasive plant nematodes. Nematodes are the root-knot disease-causing agents of tomato and brinjal, i.e., *Meloidogyne incognita* and *Meloidogyne javanica*. It was observed that amoeba had 50–65 egg ingestion rate per amoeba per 24 h of both *Meloidogyne incognita* and *Meloidogyne javanica* and the 10–20 juvenile and 6–7 adult nematode ingestion per amoeba in 24 h.

14.5 Conclusion

Currently, nitrogen fixers, phosphate solubilizers, mycorrhiza, and biocontrolling agents like *Trichoderma* sp. are the most popular bioinputs throughout the world, even though it is necessary to recommend the utilization of other microbial bioinputs like ACCD, phytase producing microorganisms, Zn, K, S mobilizers. Besides that, latest studies proved the extraordinary potential of protozoa as the real new age bioinput, which proved their beneficial power for plant growth development, soil fertility augmentation, and biocontrol of soilborne pathogen. Recent advances in protozoans as bioinput will open a new avenue for plant-microorganism interaction research to solve current agricultural problems. The microbes present in the soil employ different strategies, and these beneficial belowground microbial interventions influence the plant beneficially. The character of these new age agricultural bioinputs is noteworthy for soil and plant well-being through nutrient fixation, solubilization, mineralization, and mobilization that are eventually accountable in the agroecological perspective. Such modern biological inputs in agriculture will help to achieve the future food demand of a growing world population and address the global problem of food security and malnutrition. So there is much more to do with nature's gift microorganisms which have tremendous metabolic flexibility and potential functionality.

Acknowledgment The corresponding author, SVP, is kindly acknowledging the Department of Biotechnology, New Delhi, for the Indo-US Foldscope Major Research Project grant (Grant No. BT/IN/Indo-US/Foldscope/39/2015).

References

- Adhya TK, Kumar N, Reddy G et al (2015) Microbial mobilization of soil phosphorus and sustainable P management in agricultural soils. Curr Sci 108(7):1280–1287
- Aiba S, Humphrey AE, Millis NF (1973) Scale-up. In: Biochemical engineering, 2nd edn. Academic, New York, pp 195–217
- Angel R, Tamim NM, Applegate TJ, Dhandu AS, Ellestad LE (2002) Phytic acid chemistry: influence on phytin-phosphorus availability and phytase efficacy. J Appl Poult Res 11:471–480
- Ariza A, Moroz OV, Blagova EV et al (2013) Degradation of phytate by the 6-phytase from *Hafnia alvei*: a combined structural and solution study. PLoS One 8(5):e65062
- Azam F, Fenchel T, Field JG (1983) The ecological role of water-column microbes in the sea. Mar Ecol Prog Ser 20:257–263
- Bakthavatchalu S, Thiam B, Lokanath CK (2013) Partial purification and characterization of phytases from newly isolated *Pseudomonas aeruginosa*. Asiat J Biotechnol Resour 4:7–12
- Baldi BG, Scott JJ, Everard JD et al (1988) Localization of constitutive phytases in lily pollen and properties of the pH 8 form. Plant Sci 56:137–147
- Belimov AA, Dodd IC, Safronova VI (2007) Pseudomonas brassicacearum strain Am3 containing 1-aminocyclopropane-1-carboxylate deaminase can show both pathogenic and growthpromoting properties in its interaction with tomato. J Exp Bot 24:1–11
- Bernard C, Rassoulzadegan F (1990) Bacteria or microflagellates as a major food source for marine ciliates: possible implications for the microzooplankton. Mar Ecol Prog Ser 64(1):147–155

- Bohn L, Meyer AS, Rasmussen SK (2008) Phytate: impact on environment and human nutrition, a challenge for molecular breeding. J Zhejiang Univ Sci B 9:165–191
- Bonkowski M (2004) Protozoa and plant growth: the microbial loop in soil revisited. New Phytol 162(3):617–631
- Bonkowski M, Brandt F (2002) Do soil protozoa enhance plant growth by hormonal effects? Soil Biol Biochem 34(11):1709–1715
- Brinch-Pedersen H, Sørensen LD, Holm PB (2002) Engineering crop plants: getting a handle on phosphate. Trends Plant Sci 7:118–125
- Bünemann EK (2008) Enzyme additions as a tool to assess the potential bioavailability of organically bound nutrients. Soil Biol Biochem 40:2116–2129
- Cao L, Wang L, Yang W et al (2007) Application of microbial phytase in fish feed. Enzyme Microb Technol 40:497–507
- Casey A, Walsh G (2004) Identification and characterization of a phytase of potential commercial interest. J Biotechnol 110:313–322
- Choi YM, Suh HJ, Kim JM (2001) Purification and properties of extracellular phytase from Bacillus sp. KHU-10. J Protein Chem 20:287–292
- Chrzanowski TH, Śimek K (1990) Prey-size selection by freshwater flagellated protozoa. Limnol Oceanogr 35(7):1429–136s
- D'Silva CG, Bae HD, Yanke LJ et al (2000) Localization of phytase in Selenomonas ruminantium and Mitsuokella multiacidus by transmission electron microscopy. Can J Microbiol 46:391–395
- Davis PG, Sieburth JM (1984) Estuarine and oceanic microflagellate predation of actively growing bacteria: estimation by frequency of dividing-divided bacteria. Mar Ecol Prog Ser 19(3):237–246
- De Angelis M, Gallo G, Corbo MR et al (2003) Phytase activity in sourdough lactic acid bacteria: purification and characterization of a phytase from *Lactobacillus sanfranciscensis* CB1. Int J Food Microbiol 87:259–270
- Djordjevic S, Djukic D, Govedarica M et al (2003) Effects of chemical and physical soil properties on activity phosphomonoesterase. Acta Agric Serbica 8:3–10
- Duan J, Müller KM, Charles TC (2009) 1-aminocyclopropane-1-carboxylate (ACC) deaminase genes in rhizobia from southern Saskatchewan. Microbial Ecol 57:423–436
- Eccleston-Parry JD, Leadbeater BS (1994a) A comparison of the growth kinetics of six marine heterotrophic nanoflagellates fed with one bacterial species. Mar Ecol Prog Ser 105:167–177
- Eccleston-Parry JD, Leadbeater BS (1994b) The effect of long-term low bacterial density on the growth kinetics of three marine heterotrophic nanoflagellates. J Exp Mar Biol Ecol 177:219–233
- Ekelund F, Rønn R (1994) Notes on protozoa in agricultural soil with emphasis on heterotrophic flagellates and naked amoebae and their ecology. FEMS Microbiol Rev 15(4):321–353
- Escobin-Mopera L, Ohtani M, Sekiguchi S et al (2012) Purification and characterization of phytase from *Klebsiella pneumoniae* 9-3B. J Biosci Bioeng 113:562–567
- Ezawa T, Smith SE, Smith FA (2002) P metabolism and transport in AM fungi. Plant Soil 244(1-2):221-230
- Farhat A, Chouayekh H, Farhatben M et al (2008) Gene cloning and characterization of a thermostable phytase from *Bacillus subtilis* US417 and assessment of its potential as a feed additive in comparison with a commercial enzyme. Mol Biotechnol 64:1234–1245
- Farouk AE, Greiner R, Hussain ASM (2012) Purification and properties of a phytate-degrading enzyme produced by *Enterobacter sakazakii* ASUIA279. J Biotechnol Biodivers 3:1–9
- Findenegg GR, Nelemans JA (1993) The effect of phytase on the availability of P from myoinositol hexaphosphate (phytate) for maize roots. Plant Soil 154:189–196
- Finlayson SA, Foster KR, Reid DM (1991) Transport and metabolism of 1-aminocyclopropanecarboxylic acid in sunflower (*Helianthus annuus* L.) seedlings. Plant Physiol 96:1360–1367
- First MR, Park NY, Berrang ME (2012) Ciliate ingestion and digestion: flow cytometric measurements and regrowth of a digestion-resistant *Campylobacter jejuni*. J Eukaryot Microbiol 59:12–19

- Fitriatin BN, Joy B, Subroto T (2008) The influence of organic phosphorous substrate on phosphatase activity of soil microbes. In: Proceedings of international seminar on chemistry. 2008 Oct 30–31. Universitas Padjadjaran, Jatinangor
- Frias J, Doblado R, Antezana JR et al (2003) Inositol phosphate degradation by the action of phytase enzyme in legume seeds. Food Chem 81:233–239
- Fu S, Sun J, Qian L et al (2008) Bacillus phytases: present scenario and future perspectives. Appl Biochem Biotechnol 151:1–8
- George TS, Richardson AE, Hadobas PA et al (2004) Characterization of transgenic *Trifolium subterraneum* L. which expresses phyA and releases extracellular phytase: growth and P nutrition in laboratory media and soil. Plant Cell Environ 27:1351–1361
- George TS, Simpson RJ, Hadobas PA et al (2005) Expression of a fungal phytase gene in *Nicotiana tabacum* improves phosphorus nutrition of plants grown in amended soils. Plant Biotechnol J 3:129–140
- George TS, Simpson RJ, Gregory PJ et al (2007) Differential interaction of *Aspergillus niger* and *Peniophora lycii* phytases with soil particles affects the hydrolysis of inositol phosphates. Soil Biol Biochem 39:793–803
- Gibson DM (1987) Production of extracellular phytase from *Aspergillus ficuum* on starch media. Biotechnol Lett 9:305–310
- Glick BR (1995) The enhancement of plant growth by free-living bacteria. Can J Microbiol 41:109–117
- Glick BR, Cheng Z, Czarny J (2007) Promotion of plant growth by ACC deaminase-producing soil bacteria. Eur J Plant Pathol 119:329–339
- Goldman JC, Caron DA (1985) Experimental studies on an omnivorous microflagellate: implications for grazing and nutrient regeneration in the marine microbial food chain. Deep-Sea Res 32:899–915
- Goldman JC, Caron DA, Andersen OK (1985) Nutrient cycling in a microflagellate food chain. I. Nitrogen dynamics. Mar Ecol Prog Ser 24:231–242
- Greiner R, Sajidan I (2008) Production of D-myo-inositol (1, 2, 4, 5, 6) pentakisphosphate using alginate-entrapped recombinant *Pantoea agglomerans* glucose-1-phosphatase. Braz Arch Biol Technol 51:235–246
- Greiner R, Konietzny U, Jany KD (1993) Purification and characterization of two phytases from *Escherichia coli*. Arch Biochem Biophys 303:107–113
- Greiner R, Lim BL, Cheng C (2007) Pathway of phytate dephosphorylation by β-propeller phytases of different origins. Can J Microbiol 53:488–495
- Griffiths AJ (1970) Encystment in amoebae. Adv Microb Physiol 4:105-120
- Griffiths BS, Bardgett RD (1997) Interactions between microbe-feeding invertebrates and Soil Microorganisms. In: van Elsas JD, Trevors JT, Wellington EMH (eds) Modern soil microbiology. Marcel Dekker, New York, pp 165–182
- Guimarães LH, Terenzi HF, Jorge JA et al (2004) Characterization and properties of acid phosphatases with phytase activity produced by *Aspergillus caespitosus*. Biotech Appl Biochem 40:201–207
- Guinel FC (2015) Ethylene, a hormone at the center-stage of nodulation. Front Plant Sci 6:1121
- Gupta RK, Gangoliya SS, Singh NK (2015) Reduction of phytic acid and enhancement of bioavailable micronutrients in food grains. J Food Sci Technol 52:676–684
- Gyaneshwar P, Kumar GN, Parekh LJ et al (2002) Role of soil microorganisms in improving P nutrition of plants. Plant Soil 245:83–93
- Haefner S, Knietsch A, Scholten E et al (2005) Biotechnological production and applications of phytases. Appl Microbiol Biotechnol 68:588–597
- Hao X, Cho CM, Racz GJ et al (2002) Chemical retardation of phosphate diffusion in an acid soil as affected by liming. Nutr Cycle Agroecosyst 64:213–224
- Hayat R, Ali S, Amara U et al (2010) Soil beneficial bacteria and their role in plant growth promotion: a review. Ann Microbiol 60:579–598
- Hayes JE, Richardson AE, Simpson RJ (1999) Phytase and acid phosphatase activities in extracts from roots of temperate pasture grass and legume species. Aust J Plant Physiol 26:801–809

- Hayes J, Simpson R, Richardson A (2000) The growth and phosphorus utilisation of plants in sterile media when supplied with inositol hexaphosphate, glucose 1-phosphate or inorganic phosphate. Plant Soil 220:165–174
- Heaton K, Drinkall J, Minett A et al (2001) Amoeboid grazing on surface associated prey. In: Gilbert P, Allison DG, Brading M et al (eds) Biofilm community interactions: chance or necessity? Bioline Press, Cardiff, pp 293–301
- Hegeman CE, Grabau EA (2001) A novel phytase with sequence similarity to purple acid phosphatases is expressed in cotyledons of germinating soybean seedlings. Plant Physiol 126:1598–1608
- Herbert D (1956) Stoichiometric aspects of microbial growth. In: Evans C, Melling J (eds) Continuous culture 6: applications and new field, vol 6. Ellis Horword, Chichester, pp 1–30
- Honma M, Shimomura T (1978) Metabolism of 1- aminocyclopropane-1-carboxylic acid. Agric Biol Chem 42:1825–1831
- Howson S, Davis R (1983) Production of phytate hydrolyzing enzymes by some fungi. Enzym Microb Technol 5:377–382
- Hsiao A (2000) Effect of water deficit on morphological and physiological characterizes in rice (Oryza sativa). J Agric Res 3:93–97
- Huang H, Shi P, Wang Y (2009) Diversity of beta-propeller phytase genes in the intestinal contents of grass carp provides insight into the release of major phosphorus from phytate in nature. Appl Environ Microbiol 75:1508–1516
- Huws SA, McBain AJ, Gilbert P (2005) Protozoan grazing and its impact upon population dynamics in biofilm communities. J Appl Microbiol 98:238–244
- Idriss EE, Makarewicz O, Farouk A et al (2002) Extracellular phytase activity of *Bacillus amyloliquefaciens* FZB45 contributes to its plant growth-promoting effect. Microbiology 148:2097–2109
- Iriberri J, Ayo B, Santamaria E (1995) Influence of bacterial density and water temperature on the grazing activity of two freshwater ciliates. Freshw Biol 33:223–231
- Jacobson CB, Pasternak JJ, Glick BR (1994) Partial purification and characterization of ACC deaminase from the plant growth-promoting rhizobacterium *Pseudomonas putida* GR12-2. Can J Microbiol 40:1019–1025
- James MR, Hall JA (1995) Planktonic ciliated protozoa: their distribution and relationship to environmental variables in a marine coastal ecosystem. J Plankton Res 17:659–683
- Jassey VE, Shimano S, Dupuy C et al (2012) Characterizing the feeding habits of the testate amoebae *Hyalosphenia papilio* and *Nebela tincta* along a narrow "fen-bog" gradient using digestive vacuole content and 13C and 15N isotopic analyses. Prosit 163:451–464
- Jia YJ, Kakuta Y, Sugawara M (1999) Synthesis and degradation of 1-aminocyclopropane-1carboxylic acid by *Penicillium citrinum*. Biosci Biotech Biochem 63:542–549
- Johnson SC, Yang MP, Murthy PN (2010) Heterologous expression and functional characterization of a plant alkaline phytase in *Pichia pastoris*. Protein Express Purif 74:196–203
- Jorquera M, Martinez O, Maruyama F (2008) Current and future biotechnological applications of bacterial phytases and phytase-producing bacteria. Microbes Environ 23:182–191
- Jousset A, Bonkowski M (2010) The model predator Acanthamoeba castellanii induces the production of 2, 4, DAPG by the biocontrol strain *Pseudomonas fluorescens* Q2-87. Soil Biol Biochem 42:1647–1649
- Jousset A, Rochat L, Scheu S et al (2010) Predator-prey chemical warfare determines the expression of biocontrol genes by rhizosphere-associated *Pseudomonas fluorescens*. Appl Environ Microbiol 76:5263–5268
- Kara A, Ebina S, Kondo A et al (1985) A new type of phytase from pollen of *Typha latifolia* L. Agric Biol Chem 49:3539–3544
- Kerovuo J, Lauraeus M, Nurminen P et al (1998) Isolation, characterization, molecular gene cloning and sequencing of a novel phytase from *Bacillus subtilis*. Appl Environ Microbiol 64:2079–2085
- Kerovuo J, Rouvinen J, Hatzack F (2000) Analysis of myoinositol hexakisphosphate hydrolysis by Bacillus phytase, indication of a novel reaction mechanism. Biochem J 352:623–628

- Khan AA, Jilani G, Akhtar MS et al (2009) Phosphorus solubilizing bacteria: occurrence, mechanisms and their role in crop production. J Agric Biol Sci 1:48–58
- Kim Y-O, Lee J-K, Kim H-K et al (1998a) Cloning of thermostable phytase gene (phy) from Bacillus sp. DS11 and it's over expression in *Escherichia coli*. FEMS Microbiol Lett 162:185–191
- Kim YO, Kim HK, Bae KS et al (1998b) Purification and properties of thermostable phytase from Bacillus sp. DS11. Enzym Microbiol Technol 22:2–7
- Kim H-W, Kim Y-O, Lee J-H et al (2003) Isolation and characterization of a phytase with improved properties from *Citrobacter braakii*. Biotechnol Lett 25:1231–1234
- Klee HJ, Hayford MB, Kretzmer KA (1991) Control of ethylene synthesis by expression of a bacterial enzyme in transgenic tomato plants. Plant Cell 3:1187–1193
- Konietzny U, Greiner R (2002) Molecular and catalytic properties of phytase degrading enzymes (phytases). Int J Food Sci Technol 37:791–812
- Konietzny U, Greiner R (2004) Bacterial phytase: potential application, in vivo function and regulation of its synthesis. Braz J Microbiol 35:12–18
- Krome K, Rosenberg K, Dickler C (2010) Soil bacteria and protozoa affect root branching via effects on the auxin and cytokinin balance in plants. Plant Soil 328:191–201
- Lan GQ, Abdullah N, Jalaludin S et al (2002) Culture conditions influencing phytase production of *Mitsuokella jalaludinii*, a new bacterial species from the rumen of cattle. J Appl Microbiol 93:668–674
- Levrat P (1989) Actiond' Acanthamoeba castellarni (Protozoa: Amoebida) sur la production de siderophores par la bacterie Pseudomonas putida. C R Acad Sci Sér 3 Sci Vie 308:161–164
- Li M, Osaki M, Madhusudana Rao I et al (1997) Secretion of phytase from the roots of several plant species under phosphorus-deficient conditions. Plant Soil 195:161–169
- Li XG, Porres JM, Mullaney EJ et al (2007a) Phytase: source, structure and application. In: Industrial enzymes. Springer, Dordrecht, pp 505–529
- Li X, Wu Z, Li W et al (2007b) Growth promoting effect of a transgenic *Bacillus mucilaginosus* on tobacco planting. Appl Microbiol Biotechnol 74:1120–1125
- Li G, Yang S, Li M et al (2009) Functional analysis of an *Aspergillus ficuum* phytase gene in *Saccharomyces cerevisiae* and its root-specific, secretory expression in transgenic soybean plants. Biotechnol Lett 31:1297–1303
- Li R, Zhao J, Sun C et al (2010) Biochemical properties, molecular characterizations, functions, and application perspectives of phytases. Front Agric China 4:195–209
- Lilly VG (1965) The chemical environment for growth. 1. In: Ainsworth GC, Sussman AS (eds) The fungi, media, macro and micronutrients, vol 1. Academic, New York, pp 465–478
- Lott JN, Ockenden I, Raboy V et al (2000) Phytic acid and phosphorus in crop seeds and fruits: a global estimate. Seed Sci Res 10(1):11–33
- Luria SE (1960) The bacterial protoplasm: composition and organization. Bacteria 1:1–34
- Ma W, Charles TC, Glick BR (2004) Expression of an exogenous 1-aminocyclopropane-1carboxylate deaminase gene in *Sinorhizobium meliloti* increases its ability to nodulate alfalfa. Appl Environ Microbiol 70:5891–5897
- Marlida Y, Delfita R, Adnadi P et al (2010) Isolation, characterization and production of phytase from endophytic fungus its application for feed. Pak J Nutr 9:471–474
- Mazzola M, De Bruijn I, Cohen MF et al (2009) Protozoan-induced regulation of cyclic lipopeptide biosynthesis is an effective predation defense mechanism for *Pseudomonas fluorescens*. Appl Environ Microbiol 75:6804–6811
- Mellano HM, Munnecke DE, Endo RM (1970) Relationship of seedling age to development of Pythium ultimum on roots of Antirrhinum majus. Phytopathology 60:935–942
- Menezes-Blackburn D, Jorquera MA, Greiner R et al (2013) Phytases and phytase-labile organic phosphorus in manures and soils. Crit Rev Environ Sci Technol 43:916–954
- Minami R, Uchiyama K, Murakami T (1998) Properties, sequence and synthesis in *Escherichia coli* of 1-aminocyclopropane-1-carboxylate deaminase from *Hansenula saturnus*. J Biochem 123:1112–1118

- Minggang L, Mitsuru O, Idupulapati MR, Tadano T (1997) Secretion of phytase from the roots of several plant species under phosphorus-deficient conditions. Plant Soil 195:161–169
- Mittal V, Singh O, Nayyar H et al (2008) Stimulatory effect of phosphate solubilizing fungal strains (Aspergillus awamori and Penicillium citrinum) on the yield of chickpea (Cicer arietinum L. cv.GPF2). Soil Biol Biochem 40:718–727
- Morgan PW, Drew MC (1997) Ethylene and plant response to stress. Physiol Plant 100:620-630
- Mukhametzyanova AD, Akhmetova AI, Sharipova MR (2012) Microorganisms as phytase producers. Microbiology 81:267–275
- Mullaney EJ, Ullah AHJ (2003) Phytases: attributes, catalytic mechanisms and applications. Biochem Biophys Res Commun 312:179–184
- Mullaney EJ, Daly CB, Ullah AH (2000) Advances in phytase research. Adv Appl Microbiol 47:157–199
- Müller MS, Scheu S, Jousset A (2013) Protozoa drive the dynamics of culturable biocontrol bacterial communities. PLoS One 8:e66200
- Nielsen TH, Sorensen D, Tobiasen C et al (2002) Antibiotic and biosurfactant properties of cyclic lipopeptides produced by fluorescent *Pseudomonas* spp. from the sugar beet rhizosphere. Appl Environ Microbiol 68:3416–3423
- Page FC (1988) A new key to freshwater and soil Gymnamoebae: with instructions for culture. Freshwater Biological Association, Ambleside
- Pandey A, Szakacs G, Soccol CR et al (2001) Production, purification and properties of microbial phytases. Bioresour Technol 77:203–214
- Pasamontes L, Haiker M, Wyss M (1997) Gene cloning, purification, and characterization of a heatstable phytase from the fungus Aspergillus fumigatus. Appl Environ Microbiol 63:1696–1700
- Patel KJ, Singha AK, Nareshkumarb G (2010) Organic-acid-producing, phytate-mineralizing rhizobacteria and their effect on growth of pigeon pea (*Cajanus cajan*). Appl Soil Ecol 44:252–261
- Penmetsa RV, Cook DR (1997) A legume ethylene-insensitive mutant hyperinfected by its rhizobial symbiont. Science 275:527–530
- Penrose DM, Glick BR (2001) Levels of ACC and related compounds in exudate and extracts of canola seeds treated with ACC deaminase-containing plant growth-promoting bacteria. Can J Microbiol 47:368–372
- Penrose DM, Glick BR (2003) Methods for isolating and characterizing ACC deaminase containing plant growth-promoting rhizobacteria. Physiol Plant 118:10–15
- Powar VK, Jagannathan V (1982) Purification and properties of phytate-specific phosphatase from Bacillus subtilis. J Bacteriol 151:1102–1108
- Quan C-S, Tian W-J, Fan S-D et al (2004) Purification and properties of a low-molecular weight phytase from Cladosporium sp. FP-1. J Biosci Bioeng 97:260–266
- Quiquampoix H, Burns RG (2007) Interactions between proteins and soil mineral surfaces: environmental and health consequences. Elements 3:401–406
- Raboy V, Dickinson DB (1987) The timing and rate of phytic acid accumulation in developing soybean seeds. Plant Physiol 85:841–844
- Rahdari P, Hosseini SM, Tavakoli S (2012) The studying effect of drought stress on germination, proline, sugar, lipid, protein and chlorophyll content in purslane (*Portulaca oleracea* L.) leaves. J Med Plant Res 6:1539–1547
- Ramaekers L, Remans R, Rao IM (2010) Strategies for improving phosphorus acquisition efficiency of crop plants. Field Crop Res 117:169–176
- Reddy MS, Kumar S, Babita K (2002) Biosolubilization of poorly soluble rock phosphates by *Aspergillus tubingensis* and *Aspergillus niger*. Bioresour Technol 84:187–189
- Reddy CS, Kim SC, Kaul T (2017) Genetically modified phytase crops role in sustainable plant and animal nutrition and ecological development: a review. 3 Biotech 7:195
- Richardson AE, Hadobas PA (1997) Soil isolates of Pseudomonas spp. that utilize inositol phosphates. Can J Microbiol 43:509–516
- Richardson AE, Simpson RJ (2011) Soil microorganisms mediating phosphorus availability. Plant Physiol 156:989–996

- Richardson A, Hadobas P, Hayes J (2000) Acid phosphomonoesterase and phytase activities of wheat (*Triticum aestivum* L.). roots and utilization of organic phosphorus substrates by seedlings grown in sterile culture. Plant Cell Environ 23:397–405
- Richardson AE, Hadobas PA, Hayes JE (2001a) Extracellular secretion of Aspergillus phytase from Arabidopsis roots enables plants to obtain phosphorus from phytate. Plant J 25:641–649
- Richardson AE, Hadobas PA, Hayes JE (2001b) Utilization of phosphorus by pasture plants supplied with myo-inositol hexaphosphate is enhanced by the presence of soil micro-organisms. Plant Soil 229:47–56
- Richardson AE, Barea J-M, McNeill AM (2009) Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by microorganisms. Plant Soil 321:305–339
- Rodríguez H, Fraga R (1999) Phosphate solubilizing bacteria and their role in plant growth promotion. Biotechnol Adv 17:319–339
- Rossolini GM, Schippa S, Riccio ML et al (1998) Bacterial nonspecific acid phosphohydrolases: physiology, evolution and use as tools in microbial biotechnology. Cell Mol Life Sci 54:833–850
- Sajidan A, Farouk A, Greiner R (2004) Molecular and physiological characterisation of a 3-phytase from soil bacterium Klebsiella sp. ASR1. Appl Microbiol Biotechnol 65:110–118
- Sarapatka B (2002) Phosphatase activity of Eutric cambisols (Uppland, Sweden) in relation to soil properties and farming systems. Acta Agric Bohem 33:18–24
- Sayre RM (1973) *Theratromyxa weberi*, an amoeba predatory on plant-parasitic nematodes. J Nematol 5:258
- Schaefer M, Schauermann J (1990) The soil fauna of beech forests: comparison between a mull and a modern soil. Pedobiologia 34:299–314
- Scholz RW, Hellums DT, Roy AA (2015) Global sustainable phosphorus management: a transdisciplinary venture. Curr Sci 108:3–12
- Schröter D, Wolters V, De Ruiter PC (2003) C and N mineralisation in the decomposer food webs of a European forest transect. Oikos 102:294–308
- Scott JJ (1991) Alkaline phytase activity in nonionic detergent extracts of legume seeds. Plant Physiol 95:1298–1301
- Selvakumar G, Reetha S, Thamizhiniyan P (2012) Response of biofertilizers on growth, yield attributes and associated protein profiling changes of blackgram (*Vigna mungo* L. Hepper). WASJ 16:1368–1374
- Sgherri C, Stevanovic B, Navari-Izzo F (2000) Role of phenolic acids during dehydration and rehydration of *Ramonda serbica*. Physiol Plant 122:478–485
- Sharma A, Rawat US, Yadav BK (2012) Influence of phosphorus levels and phosphorus solubilizing fungi on yield and nutrient uptake by wheat under sub-humid region of Rajasthan, India. ISRN Agron 15:2012
- Sherr BF, Sherr EB, Fallon RD (1987) Use of monodispersed, fluorescently labeled bacteria to estimate in situ protozoan bacterivory. Appl Environ Microbiol 53:958–965
- Shimizu M (1992) Purification and characterization of a phytase from *Bacillus subtilis* (natto) N-77. Biosci Biotechnol Biochem 56:1266–1269
- Sieburth JM, Davis PG (1982) The role of heterotrophic nanoplankton in the grazing and nurturing of planktonic bacteria in the Sargasso and Caribbean Seas. Ann Inst Oceanogr 58(S):285–296
- Singh B, Satyanarayana T (2010) Plant growth promotion by an extracellular HAP-phytase of a thermophilic mold *Sporotrichum thermophile*. Appl Biochem Biotechnol 160:1267–1276
- Singh B, Satyanarayana T (2015) Fungal phytases: characteristics and amelioration of nutritional quality and growth of non-ruminants. J Anim Physiol Anim Nutr 99:646–660
- Singh B, Kunze G, Satyanarayana T (2011) Developments in biochemical aspects and biotechnological applications of microbial phytases. Biotechnol Mol Biol Rev 6:69–87
- Tamimi SM, Timko MP (2003) Effects of ethylene and inhibitors of ethylene synthesis and action on nodulation in common bean (*Phaseolus vulgaris* L.). Plant Soil 257:125–131
- Tanaka Y, Sano T, Tamaoki M (2005) Ethylene inhibits abscisic acid-induced stomatal closure in Arabidopsis. Plant Physiol 138:2337–2343

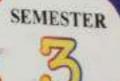
- Tarafdar JC (1995) Dual inoculation with Aspergillus fumigatus and Glomus mosseae enhances biomass production and nutrient uptake in wheat (*Triticum aestivum* L.) supplied with organic phosphorus as Na-Phytate. Plant Soil 173:97–102
- Tarafdar JC, Yadav RS, Meena SC (2001) Comparative efficiency of acid phosphatase originated from plant and fungal sources. J Plant Nutr Soil Sci 164:279–282
- Tittabutr P, Piromyou P, Longtonglang A (2013) Alleviation of the effect of environmental stresses using co-inoculation of mungbean by *Bradyrhizobium* and *Rhizobacteria* containing stressinduced ACC deaminase enzyme. Soil Sci Plant Nutr 59:559–557
- Tran HT, Hurley BA, Plaxton WC (2010) Feeding hungry plants: the role of purple acid phosphatases in phosphate nutrition. Plant Sci 179:14–27
- Turk M, Sandberg AS, Carlsson N et al (2000) Inositol hexaphosphate hydrolysis by baker's yeast. Capacity, kinetics and degradation products. J Agric Food Chem 48:100–104
- Turner BL, Papházy MJ, Haygarth PM et al (2002) Inositol phosphates in the environment. Philos Trans R Soc Lond B Biol Sci 357:449–469
- Tye AJ, Siu FKY, Leung TYC et al (2002) Molecular cloning and the bio-chemical characterization of two novel phytases from *Bacillus subtilis* 168 and *Bacillus licheniformis*. Appl Microbiol Biotechnol 59:190–197
- Unno Y, Okubo K, Wasaki J et al (2005) Plant growth promotion abilities and microscale bacterial dynamics in the rhizosphere of Lupin analysed by phytate utilization ability. Environ Microbiol 7:396–404
- Vats P, Banerjee UC (2004) Production studies and catalytic properties of phytases (myo-inositol hexakisphosphate phosphohydrolases): an overview. Enzym Microb Technol 35:3–14
- Vats P, Bhattacharyya MS, Banerjee UC (2005) Use of phytases (myo-inositolhexakis phosphate phosphohydrolases) for combating environmental pollution: a biological approach. Crit Rev Environ Sci Technol 35:469–486
- Wallenstein MD, Burns RG (2011) Ecology of extracellular enzyme activities and organic matter degradation in soil: a complex community-driven process. In: Dick RP (ed) Methods of soil enzymology. Soil Sci Soc Am, Madison, pp 35–55
- Wallenstein MD, McMahon SK, Schimel JP (2009) Seasonal variation in enzyme activities and temperature sensitivities in Arctic tundra soils. Glob Chang Biol 15:1631–1639
- Weidner S, Latz E, Agaras B (2017) Protozoa stimulate the plant beneficial activity of rhizospheric pseudomonads. Plant Soil 410:509–515
- Wyss M, Brugger R, Kronenberger A et al (1999) Biochemical characterization of fungal phytases (myo-inositol hexakisphosphate phosphohydrolases): catalytic properties. Appl Environ Microbiol 65:367–373
- Xiao C, Chi R, Li X et al (2011) Biosolubilization of rock phosphate by three stress-tolerant fungal strains. Appl Biochem Biotechnol 165:719–727
- Xuguang N, Lichao S, Yinong X et al (2018) Drought-tolerant plant growth-promoting Rhizobacteria associated with foxtail millet in a semi-arid agroecosystem and their potential in alleviating drought stress. Front Microbiol 8:2580
- Yang SF, Hoffman NE (1984) Ethylene biosynthesis and its regulation in higher plants. Ann Rev Plant Physiol 35:155–189
- Yoon SJ, Choi YJ, Min HK et al (1996) Isolation and identification of phytase-producing bacterium, *Enterobacter* sp. 4, and enzymatic properties of phytase enzyme. Enzym Microb Technol 18:449–454
- Zahir ZA, Arshad M, Frankenberger WT (2004) Plant growth promoting rhizobacteria: applications and perspectives in agriculture. Adv Agron 81:98–169
- Zamudio M, González A, Medina JA (2001) *Lactobacillus plantarum* phytase activity is due to nonspecific acid phosphatase. Lett Appl Microbiol 32:181–184
- Zamudio M, González A, Bastarrachea F (2002) Regulation of *Raoultella terrigena* comb.nov. phytase expression. Can J Microbiol 48:71–81

AS PER THE CBCS SYLLABUS OF WARDHA UNIVERSITIES

A TEXT BOOK OF ELEMENTS OF CHEMISTRY

- TEKADE
- MOHABANSI
- DADURE
- BANSINGE
- BARWAT
- BORKAR

A Pragati Edition



OTHER USEFUL BOOKS

Inorganic Chemistry I, II & III. Organic Chemistry 1, II & III Physical Chemistry I, II & III अकार्वनिक रसायन भाग 1, 2, 3 कार्वनिक, रसायन भाग 1, 2, 3 भौतिक रसायन भाग 1, 2, 3 B.Sc. Chemistry 1, 2, 3 (Unified) बी.एस.सी. रसायन भाग 1, 2, 3 (Unified) वी.एस.सी. रसायन भाग 1, 2 (M.P.) वी.एस.सी. रसायन प्रथम जर्ष (Durg) Help Books -Inorganic Chemistry I, II, III Organic Chemistry I, II, III . Physical Chemistry I, II, III अकाबीनेक रसायन भाग 1, 2, 3 काबीनेक रसायन भाग 1, 2, 3 भौतिक रसायम भाग-1, 2, 3 Practical Books

Text Books

Analytical Chemistry I Analytical Chemistry II Analytical Chemistry III -H.C. Khera -Jagdamba Singh, L.D.S. Yaday -J.N. Gurtu -एच.सी. खेरा, रविळान्त -एच.सी. खेरा, रविळान्त -जे.एन. गुर्टू, एच.सी. खेरा -Gurtu, Singh, Khera -जे.एन. गुर्टू, एच.सी. खेरा, रविळान्त -पिम्पलापुरे, सिद्दिकी, चितले, जेन तिवारी, गुप्ता, गुप्ता, सिंह

> - Gurtu, Khera - Gurtu, Khera - ज़िंदू व खेरा - ज़ुर्दू व खेरा - ज़ुर्दू व खेरा - जुर्दू व खेरा - जुर्दू व खेरा - जुर्दू व खेरा - ज़ुर्दू व खेरा - ड.K. Agarwal, Keemti Lal

- S.K. Agarwal, Keemti Lal

PRAGATI PRAKASHAN, MEERUT

♣ Pragati Bhawan, 240, W.K. Road, Meerut.
 © 0121-2640642, 2643636, 4007643
 ➡ support@pragatiprakashan.in
 ⊕ www.pragatiprakashan.in



C Author-A Text Book of Elements of Chemistry-III Sem

PRAGATI PRAKASHAN

Head Office : Educational Publishers PRAGATI BHAWAN 240, W. K. Roso, Meerut-250 001 SMS/Ph.: (0121) 6544642, 6451644 Tele/Fax: (0121) 2640642, 2643636 www.pragatiprakashan.in e-mail : pragatiprakashan@gmail.com Repd. Office : New Market, Begum Bridge, Meerut-250 001

First Edition 2019

ISBN-978-93-89403-57-2

Price : ₹ 325/-

Published by : A.K. Mittal for Pragati Prakashan, Meerut-250 001, Laser Typesetting : Pragati Prakashan, Printed at Annah Electric Press, Meerut.

Pragati's.

BAJAJ COLLEGE OF SCIENCE, WARDHA

A TEXT BOOK OF ELEMENTS OF CHEMISTRY

(For B.Sc. II Semester III, Bajaj College of Science, Wardha)

(CT)

Dr. P. V. Tekade M. Sc. B. Ed. SET, NET, GATE Ph. D.

> Dr. K. M. Dadure M. Sc. MPhil Ph. D.

Mr. N. A. Barwat

Dr. Mrs. N. P. Mohabansi M. Sc. B. Ed. SET Ph. D.

Mr. M. D. Bansinge M. Sc. NET

Dr. P. G. Borkar M. Sc. SET, NET, GATE, Ph. D. Post Doc.

Department of Chemistry Bajaj College of Science, WARDHA



PRAGATI PRAKASHAN

Contents at a Glance

UNIT-I

- A. Molecular Orbital Theory UNIT-II
- A. Chemistry of Elements of First Transition Series
- B. Chemistry of Elements of Second and Third Transitions series
- C. Non-Aqueous Solvents

UNIT-III

- A. Alkyl Halides
- B. Aryl Halides
- C. Organometallic Compounds UNIT-IV
- A. Alcohols
- B. Phenols
- C. Amines

UNIT-V

A. Chemical Kinetics

UNIT-VI

- A. Ionic Equilibrium
- B. Colligative Properties

UNIT-VII

PRACTICALS

Inorganic Chemistry Experiments Organic Chemistry Experiments Physical Chemistry Experiments Structure Elucidation of Organic Compounds by Spectroscopic Techniques

Dr. Pradip V. Tekade

About the Author



Dr. Pradip V. Tekade

M.Sc. (Chem.), Ph.D., NET, SET, GATE, Associate Professor in Chemistry Jankidevi Bajaj College of Science Wardha, Maharashtra (Autonomous)

His specialization is organic chemistry and has more than 15 years of teaching experience. He has published over 44 research papers in Journals and nine books. He has three UGC sponsored research projects to his credit and also recognized research supervisor for Rashtrasant Tukdoji Maharaj, Nagpur University.



Publisher & Distributors

A-22/1, Akash Vihar (Baprola), Street No. 1, Shaheed Chander Shekhar Marg, Near - Bakkarwala Mode, Nangloi - Najafgarh Road, New Delhi - 110043, E-Mail: ssbook@rediffmail.com / selectivebook@gmail.com E-mail: info@ssbook.in // website: www.ssbook.in Moh. 09810766022 / 09810766722 Website: www.foreasicbookstore.com



Thermoluminescence glow curve analysis of RE doped LiMgBO₃ phosphor using GCCD function

Cite as: AIP Conference Proceedings **2104**, 030043 (2019); https://doi.org/10.1063/1.5100470 Published Online: 07 May 2019

M. M. Yerpude, and S. J. Dhoble





AIP Conference Proceedings **2104**, 030043 (2019); https://doi.org/10.1063/1.5100470 © 2019 Author(s).

Get 30% off all

print proceedings!

AP Conference Proceedings

2104, 030043

Enter Promotion Code [PDF30] at checkout

Thermoluminescence Glow Curve Analysis of RE Doped LiMgBO₃ Phosphor Using GCCD Function

M.M. Yerpude^{1,a)}, S.J. Dhoble^{2,b)}

¹Department of Physics, J. B. College of Science, Wardha, India. ²Department of Physics, RTM Nagpur University, Nagpur, India.

^{a)}Corresponding author: mangesh.yerpude@gmail.com, ^{b)}sjdhoble@gmail.com

Abstract. The sol-gel synthesized Eu^{3+} , Dy^{3+} and Tb^{3+} doped LiMgBO₃ was studied for its TL response. The phase purity of the sample is checked using powder XRD pattern. The TL studies were performed by irradiating samples with γ -ray from ⁶⁰Co irradiation source in the dose range 10Gy to 1kGy. The samples have shown a linear dose response over the said range. The complex glow curves were analyzed using GCCD function given by Kitis taking general order kinetic into consideration. The activation energy and order of kinetics were calculated using the curve fitting method and frequency factors using Chen's formula.

INTRODUCTION

Boron-based materials are very important for radiation dosimetric applications due to their tissue equivalent absorption coefficient, low cost, thermal stability and neutron sensitivity[1–4]. Boron-based materials show interesting results of thermoluminescence when exposed to ionizing radiations. Improvement in the TL characteristics of borates was reported for recently developed rare earth doped mixed lithium calcium borates (LCBs)[5,6]. Previously luminescence properties of Lithium borate and Magnesium borate in both microcrystalline and nanocrystalline form has been studied. Recently researchers studied lithium magnesium borate phosphor and was found to have the lowest Z_{eff} value and suitable for a dosimetric application. Earlier very few reports were found for LiMgBO3 [7]. Recently the thermoluminescence properties of rare earth (RE= Tb, Gd, Dy, Pr, Mn, Ce, Eu) doped Lithium magnesium borate, synthesized by simple solid state diffusion technique, were studied [1]. The optical properties of Lithium magnesium borate glasses doped with Dy^{3+} and Sm^{3+} were reported by Alajerami for their potential application as laser material [8]. The intense red emitting Eu³⁺ doped LiMgBO₃ and the effect of codoping by Bi³⁺ on photoluminescence properties were reported by Liang *et. al.*[9]. The present study reports the TL response and glow curve analysis of sol-gel synthesized rare earth (RE) doped LiMgBO₃ phosphor.

EXPERIMENTAL

The starting materials were LiNO₃, Mg(NO₃)₂, H₃BO₃, citric acid, ethylene glycol and RE₂O₃ (RE=Eu, Dy, Tb) all of AR grade procured from Loba Chemie. RE(NO₃)₃ salts were freshly prepared by a reaction of RE₂O₃ with dilute nitric acid. Lithium, magnesium, europium, dysprosium, and terbium citrates were prepared from appropriate mixtures of nitrates with citric acid and ethylene glycol in aqueous media. After being evaporated for several hours in an 80 °C water-bath, the solution becomes a pale yellowish polymeric gel. The gel was dried at 150 °C for 12h, and then the precursor was calcined on a high-temperature muffle furnace at 600 °C temperatures for 2 h.TL glow curves were recorded using a Harshaw TLD reader (Model 3500) fitted with a 931B photomultiplier tube (PMT). The heating rate for TL measurement is kept at 5 °C/s.

International Conference on "Multidimensional Role of Basic Science in Advanced Technology" ICMBAT 2018 AIP Conf. Proc. 2104, 030043-1–030043-5; https://doi.org/10.1063/1.5100470 Published by AIP Publishing, 978-0-7354-1836-3/\$30.00

RESULTS & DISCUSSION

XRD

To check the phase purity of the doped and undoped materials, XRD pattern was recorded on Rigaku diffractometer with Cu-K α radiation (λ =1.5406 Å) at 40 kV tube voltage and 75 mA tube current with the step size 0.02° of 20.The XRD patterns of as-prepared LiMgBO₃, LiMgBO₃:Eu³⁺, LiMgBO₃:Dy³⁺, LiMgBO₃:Tb³⁺ phosphorswere matched well with the standard JCPDS card no. 79–1996 (LiMgBO₃). LiMgBO₃ crystallized in the hexagonal crystal structure with space group C2/c (15). Also, it has been observed that the addition of small amounts of RE³⁺ (activator) did not alter the crystal structure of the host lattice, which suggests that the activator ion was fully incorporated in the host lattice. The calculated lattice parameters for pure LiMgBO₃ were approximated to be a = 5.163 Å, b = 8.885 Å, c = 9.914 Å, β = 91.22° and V = 454.787 Å³.

Thermoluminescence Studies in Eu, Dy, and Tb Doped LiMgBO₃

For taking thermoluminescence (TL) measurements 5 mg of each sample is exposed to 10 Gy of γ -ray dose from ⁶⁰Co irradiation source at room temperature. The heating rate of 5 °C/s is chosen for TL glow curve measurements. The TL glow curves for LiMgBO₃: Eu^{3+} for various concentrations of Eu^{3+} ions is shown in Fig. 1(a). The glow curve shows two well-separated peaks at 146 and 381 °C. With the increasing concentration of Eu³⁺ ions the intensity of both the peaks increases, the peak at 381 °C saturates early and shows decrease in intensity above 0.2 mol%, whereas the intensity of glow peak at 146 °C continues to increase till 0.5 mole% and further increase in concentration it saturates (Inset of Fig. 1(a)). The TL glow curves for Dy³⁺ doped samples (Fig. 1(b)) shows the main glow peak at 140 °C and a shoulder near 112 °C. The intensity of TL signal increases up to 0.5 mol% and a further increase of Dv^{3+} ions decreases the TL intensity (inset of Fig. 1(b)). The TL glow curves for Tb^{3+} doped LiMgBO₃ (Fig. 1(c)) shows a clear complex nature. For the lower concentration of Tb³⁺ions, the glow curve shows approximately four peaks which on increasing concentration decreases and mainly two overlapped glow peaks are clearly seen. The TL intensity increases with increasing concentration of Tb³⁺ ions and reaches a maximum at 0.5 mol% (inset Fig. 1(c)). Further addition of more Tb³⁺ ions decreases the TL intensity and two well-resolved peaks can be observed. The glow peak with maximum intensity (0.5 mol%) has the main peak of 191 °C and a shoulder near 161 °C. From the analysis of the glowcurve, it can be seen that the Dy^{3+} doped samples are having the highest intensity and Eu^{3+} doped samples show the least intensity. Thus $LiMgBO_3:Dy^{3+}$ is proved to be more sensitive than the other two. For further investigation of signal fading, dose-response and kinetic parameters, samples with the highest TL intensity are chosen.

The stability of traps can be examined by keeping irradiated samples for longer duration and taking TL measurements over certain intervals also known as TL fading. TL fading is taken over a period of 15 days. The TL signal of Dy^{3+} doped material fades about 40%, whereas the Eu^{3+} doped material fades about 35%. The Tb^{3+} doped material shows least fading with 5%. Thus it can be concluded that the traps in Dy^{3+} and Eu^{3+} doped materials are quite unstable and traps in Tb^{3+} doped materials are stable. Thus irradiation memory of Tb^{3+} doped materials can be sustained for a longer time than the other two. For dose-responsestudies, the phosphor materials are exposed to γ -ray dose of 10 Gy to 1 kGy. All the materials show linear TL response over the said range and the glow curve structure remains invariant under changing the dose. This is the key feature for any tissue equivalent low Z dosimetric material.

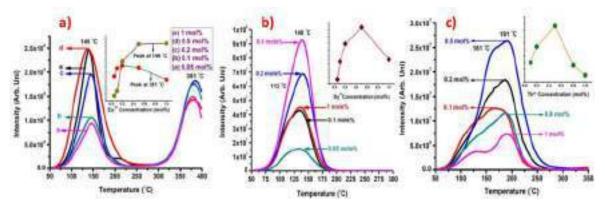


FIGURE 1.TL Glow curve for a) LiMgBO₃:xEu³⁺, b) LiMgBO₃:xDy³⁺ and c) LiMgBO₃:xTb³⁺ (x= 0.05, 0.1, 0.2, 0.5, 1 mol%) exposed to 10 Gy γ -ray from ⁶⁰Co.

TL Glow Curves Analysis and Determination of Kinetic parameters

For the analysis and determination of kinetic parameters of TL glow curves, many authors have suggested several methods [10–14]. Most of the methods require well resolved single glow curve. But for complex glow curve analysis, these methods can't be directly applied. The first step is to separate these individual glow peaks from the complex glow curve. Currently, no experimental methods are available to completely separate out individual glow curve. But combining some experimental and some theoretical processes could be useful also known as glow curve convolution de-convolution (GCCD)[13,15]. The process of GCCD requires first the approximation of maximum peak temperature (T_m) and maximum intensity (I_m) of a glow curve that has to be separated from the complex glow curve. Both can be obtained experimentally by using the process of thermal cleaning. The explanation of the thermal cleaning process can be found elsewhere [16]. We have used modified efforts for the thermal cleaning process. For this process, we have irradiated about 100 mg of all three samples with 50 Gy gamma-ray dose and taking 5 mg of each sample 20 batches are made each batch having a set of three irradiated materials. Each batch is kept for 10 s at a different temperature from 60 to 250 °C with a difference of 10 °C. We have stopped to only 250 °C as 2 out of three samples show the absence of any possible glow peak above 250 °C, only in the Eu³⁺ doped sample a glow peak at 381 °C is seen which is taken as an individual glow peak. The samples annealed at different temperature are taken for TL measurements immediately. All the glow curves for each sample are collected and carefully examined. From the analysis of the data, some rough estimation of T_m and I_m is made. This data is further used to generate a theoretical glow curve by using the GCCD function for general order kinetic (GOK) given by Kitis [13]. The GCCD function for GOK is fitted into an excel spreadsheet keeping activation energy (E) and order of kinetic (b) as variable parameters. The solver program in excel is used to minimize the figure of merit (FOM). The fitting method by GCCD function in excel spreadsheet with solver program is explained in detail by Afouxenidis et. al. [17]. The GCCD equation for GOK to fit the data is given in Eq. (1) and FOM can be calculated using Eq. (2). The frequency factor (s) is calculated using Eq. (3) given by Chen[14].

$$I(T) = I_m \times b^{b/b-1} \times \exp\left(\frac{E}{kT} \times \frac{T-T_m}{T_m}\right) \times \left[(b-1) \times \left(1 - \frac{2kT}{E}\right) \times \frac{T^2}{T_m} \times \exp\left(\frac{E}{kT} \times \frac{T-T_m}{T_m}\right) + Z_m\right]^{-b/b-1}$$
(1)

Where, $Z_m = 1 + (b-1)\frac{2kT_m}{E}$ and $k = 8.617 \times 10^{-5} eV/K$ =Boltzman's constant

$$FOM(\%) = 100 \times \frac{\Sigma |I_{exp} - I_{fit}|}{\Sigma I_{fit}}$$
(2)

I_{exp} and I_{fit} experimental and fitted TL intensity.

$$s = \frac{\beta E}{kT_m^2} \exp\left(\frac{E}{kT_m}\right) \left[1 + (b-1)\Delta_m\right]$$
(3)

Phosphor	Glow Peak Temperature	Activation Energy	Order of Kinetics	Frequency Factor
	111 °C	0.87 eV	1.2	8.83×10 ¹⁰ s ⁻¹
$LiM_{\alpha}DO \cdot En^{3+}$	140 °C	0.92 eV	2	4.90×10 ¹⁰ s ⁻¹
LiMgBO ₃ :Eu ³⁺	181 °C	1.02 eV	2	5.61×10 ¹⁰ s ⁻¹
	381 ^{oc}	1.6 eV	2	4.33×10 ¹¹ s ⁻¹
$LiM_{\alpha}DO : Du^{3+}$	116 °C	0.92 eV	1.2	2.89×10 ¹¹ s ⁻¹
LiMgBO ₃ :Dy ³⁺	141 °C	1.04 eV	1.4	1.57×10 ¹² s ⁻¹
-	111 °C	0.81 eV	2	1.26×10 ¹⁰ s ⁻¹
LiMgBO ₃ :Tb ³⁺	154 ^o C	0.87 eV	1.2	$5.05 \times 10^9 \text{ s}^{-1}$
	194 ⁰ C	1.11 eV	2	2.62×10 ¹¹ s ⁻¹

TABLE 1.Kinetic parameters for various deconvoluted glow peaks of a complex glow curve.

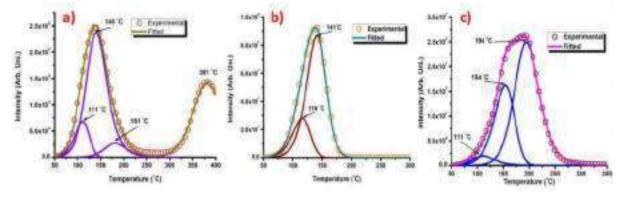


FIGURE 2. Experimental and theoretically fitted TL glow curves a) LiMgBO₃:0.5Eu³⁺, b) LiMgBO₃:0.5Dy³⁺ and c) LiMgBO₃:0.2Tb³⁺ exposed to 10 Gy γ-ray from ⁶⁰Co using GCCD function.

The obtained results are shown in Fig. 2 (a), (b) and (c) and the values of kinetic parameters are tabulated in Table.1. Form the fitting data it can be seen that the Eu^{3+} doped sample has four glow peaks at 111, 140, 181 and 381 °C, Dy^{3+} sample have two glow peaks at 116 and 141°C and Tb³⁺ samples have three glow peaks at 111, 154 and 194 °C. The FOM for Eu^{3+} , Dy^{3+} , and Tb³⁺ doped samples are 3.81%, 3.25%, and 3.05% respectively, which are well below 5% and thus can be considered as the good fit.

CONCLUSIONS

 Eu^{3+} , Dy^{3+} , and Tb^{3+} doped LiMgBO₃wassuccessfully prepared using sol-gel synthesis method. The XRD studies show the pure phase of LiMgBO₃ with a small amount of MgO phase. The addition of RE impurity in the host lattice didn't affect the actual crystal structure apart from alteration in lattice parameters. TL studies show that the TL glow curve of Eu^{3+} doped LiMgBO₃ has total four glow peaks, three of them are in shallow trap region and one in the deep trap region. The Dy^{3+} doped LiMgBO₃ has two glow peaks, both in shallow trap region. The Tb^{3+} doped samples show three glow with two in shallow trap region and one in deep trap region.

ACKNOWLEDGEMENT

One of the authors MMY is thankful to UGC for providing financial assistance through UGC-NET-JRF scheme in science, social science and humanities sanctioned vide file no. F.17-86/2008(SA-I).

REFERENCES

- 1. S.R. Anishia, M.T. Jose, O. Annalakshmi, V. Ramasamy, J. Lumin. 131, 2492–2498 (2011).
- 2. M. Prokic, Radiat. Meas. 33, 393–396 (2001).
- 3. Z. Fu, S. Pan, F. Yang, S. Gu, X. Lei, Y. Heng, G. Ren, M. Qi, Radiat. Meas. 72, 39–43 (2015).
- 4. L.H. Jiang, Y.L. Zhang, C.Y. Li, J.Q. Hao, Q. Su, Mater. Lett. 61, 5107–5109 (2007).
- 5. İ. Pekgözlü, E. Erdoğmuş, S. Çubuk, A. Sadi Başak, J. Lumin. 132, 1394–1399 (2012).
- 6. S.R. Anishia, M.T. Jose, O. Annalakshmi, V. Ponnusamy, V. Ramasamy, J. Lumin. 130, 1834–1840 (2010).
- 7. R. Norrestam, Zeitschrift Für Krist. 187, 103–110 (1989).
- 8. Y.S.M. Alajerami, S. Hashim, W.M.S. Wan Hassan, A. Termizi Ramli, A. Kasim, Phys. B Condens. Matter 407, 2398–2403 (2012).
- 9. Z. Liang, F. Mo, X. Zhang, L. Zhou, J. Lumin. 151, 47–51 (2014).
- 10. M.S. Rasheedy, J. Fluoresc. 15, 485–491 (2005).
- 11. R. Chen, S.A.A. Winer, J. Appl. Phys. 41, 5227-5232 (1970).
- 12. G. Kitis, V. Pagonis, Nucl. Instrum. Methods Phys. Res. B 262, 313-322 (2007).
- 13. G. Kitis, J.M. Gomez-Ros, J.W.N. Tuyn, J. Phys. D. Appl. Phys. 31, 2636–2641 (1998).
- 14. S.W.S. McKeever, R. Chen, Radiat. Meas. 27, 625-661 (1997).
- 15. A.M. Sadek, H.M. Eissa, A.M. Basha, G. Kitis, Nucl. Instrum. Methods Phys. Res. B 330, 103-107 (2014).
- 16. V. Pagonis, G. Kitis, C. Furetta, Numerical and Practical Exercises in Thermoluminescence, (Springer, New York, 2011) pp. 8-20.
- 17. D. Afouxenidis, G.S. Polymeris, N.C. Tsirliganis, G. Kitis, Radiat. Prot. Dosimetry. 149, 363-370 (2012).

DIGITAL ELECTRONICS FUNDAMENTALS

V.V. Shinde S.J. Dhoble

DIGITAL ELECTRONICS FUNDAMENTALS

Engineering

This book on Digital Electronics, gives full coverage of basic electronics with simple and clear diagrams, numerous in chapter solved examples and review questions at the end of chapters for understanding the concept and assimilating the theory comprehensively.

Although many books are available in the areas of digital electronics, this book is developed specifically to meet the needs of a self-paced course in which students are expected to study the material on their own.

The book is ideal for students who are studying digital electronics for the first time at any level including degree, diploma and post-graduation in electronic science and engineering, computer science and physics departments.

V.V. Shinde is serving as Associate Professor and Head. Department of Electronics. Jankidevi Bajaj College of Science, Wardha, Maharashtra, India.

S.J. Dhoble is serving as Associate Professor d, Department of Physics, RTM Nagpur University, Nagpur, Maharashtra, India.



S TUDERA P RESS 1586/113, FF, Tri Nagar Delhi- 110035, India Ph: 011 27383728 Email: info@studerapress.com Web: www.studerapress.com





INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S

RESEARCH JOURNEY

International E-Research Journal

PEER REFREED & INDEXED JOURNALFebruary-2019Special Issue – 110 (H)

Electronics

Guest Editor :

Dr. F. C. Raghuwanshi

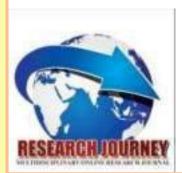
Principal,

- Vidya Bharati Mahavidyalya, Amarawati
- & Dean, Faculty of Science & Technology,
- Sant Gadge Baba Amravati University, Amravati

Executive Editors of the Issue :

- Dr.D. S. Dhote
- Professor & Head, Department of Electronics, Brijalal Biyani College, Amaravati
- Dr. R. A. Mishra
- Pircipal, Amolakchand Mahavidyala, Yeotmal
- Dr. Y. B. Gandule
- Principal, Adharsh Mahavidyala, Dhamangaon Rly.
- Dr. Bhimarao Ladgaonkar
- Professor & Head, Department of Electronics, Mohite Mahavidyala,Akluj Dist.Solapur Dr. P. B. Dahikar
 - Professor & Head, Department of Electronics ,Kamala Nehru Mahavidyala,Nagpur

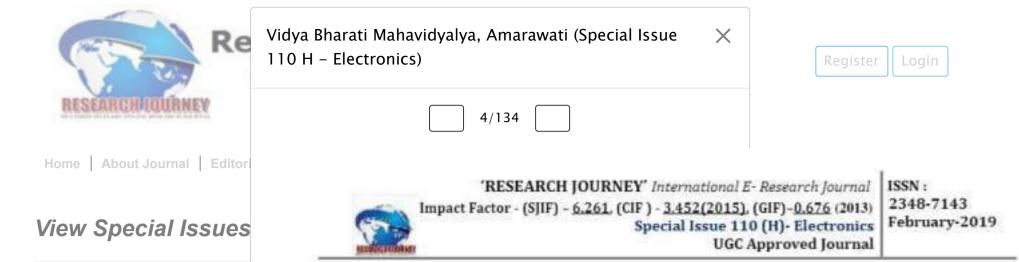
Chief Editor : Dr. Dhanraj Dhangar (Yeola)



This Journal is indexed in :

- University Grants Commission (UGC)
- Scientific Journal Impact Factor (SJIF)
- Cosmoc Impact Factor (CIF)
- Global Impact Factor (GIF)
- International Impact Factor Services (IIFS)





INDEX

No.	Title of the Paper Author's Name	Page No.
1	Virtual Instrumentation of Some Characterized Biomaterials Arun V. Padole, & Y.B. Gandole	05
2	A High Performance Cmos 1 Bit full Adder Ayushi S. Patankar	14
3	Efficient Smart Waste Management System With Multifunction Embedded Controller Pradeep B Dahikar	18
4	Characterists of Electronic Transducer in Biomedical Instruments Dr.Narendra B.Raut & Dr.Rammanohar A.Mishra	24
5	Use of AINN Filters for Reduction of Noise from ECG Signals R.J.Gajbe & Dr. Y.B.Gandole	30
6	Study of Open Source Tools and Technologies for Data Mining and Data Visualization Dr. Sumita U. Sharma,	37
7	Design and Development of Microcontroller Based Low Cost System for CO2 Trapping In Greenhouse P. A. Saudagar, D. S. Dhote, & G. A. Raut	41
8	Alternative Miracle Vehicle Gaurav.K.Dewale & Abhiram.M.Lomte	44
9	Advanced Sit and Reach Flexibility Measurement Test Box Using Arduino Sachin Winchurkar & Sudhakar Sarkate	47
10	Selection of Transfer Function for The Hidden Layer and Output Layer of MLP Network Y.M. Pharkade & Dr. G.D. Agrahari	51
11	U-Slotted Reconfigurable Multiband Micro Strip Antenna for Wireless Networks and SDR Sharanagouda N Patil , P.V Hunagund & R M Vani	56
12	Integrated High-Tech Intelligent Security System C.R.Chaudhari , S.V.Dudul & D.R.Solanke	61
13	Methods of Non-Invasive Blood Glucose Monitoring Using Nir Spectroscopy: A Review Nilima Jajoo, Dr. Deepak Dhote, & Dr. Gopal Agrahari	67
14	Microcontroller Based Techeye System for Obstacle Detection & Ranging To Assist Blind Person Yash Vidyasagar & Shri R.G.Chavan	72
15	Controlling Home Appliances Using Advanced Microcontroller: A Novel Approach Rameshwar S. Devhade, D.R. Solanke and S D Pachpande	74
16	Arduino Uno Based Accident Avoiding System IN Mountainous Area ACROSS U-Turn R. D. Chaudha & Dattaraj Vidyasagar	79
17	Green Electricity Response of Silver and Magnesium Electrode Pair Mr. G. S. Wajire & Dr. Y. B. Gandole	81
18	Design and Implementation of Fuzzy Logic Technique for Aircraft Control System K.Y. Rokde, P.B.Dahikar, S.S.Shende, & S.M.Ghatole	85
19	An Intelligent Controller for Greenhouse Temperature Control Using Fuzzy Logic P. A. Saudagar, D. S. Dhote, & G. V. Lakhotiya	92
20	Enhancement of Bandwidth and Reduction of Mutual Coupling in Microstrip Antenna Array K. Prahlada Rao, Vani R.M. & P.V. Hunagund	97
21	Evaluation of Mixed Multicast Architecture for Internet of Things Environment Using Adaptive Fountain Code Miss.Reshma Siddique & Dr.V.M.Thakare	107
	Cloud Computing With Big Data: Challenges & Issues	114

List of Article :

Select Year :

2019

2019	
Sr.	Date
51	4 February, 2019
52	4 February, 2019
53	4 February, 2019
54	4 February, 2019
55	4 February, 2019
56	4 February, 2019
57	4 February, 2019
58	5 February, 2019
59	5 February, 2019
60	5 February, 2019



An Intelligent Controller for Greenhouse Temperature Control Using Fuzzy Logic

P. A. Saudagar1, D. S. Dhote2, G. V. Lakhotiya3

1Jankidevi Bajaj College of Science, Wardha, India, saudagar.pa@gmail.com 2Brijlal Biyani Science College, Amravati, India, dsdhote@rediffmail.com 3Jankidevi Bajaj College of Science, Wardha, India, lakhotiya.govinda@gmail.com

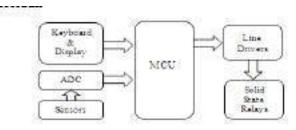
Abstract –

The present paper deals with the design of an intelligent controller for greenhouse to control temperature inside the greenhouse. A fuzzy logic is used to control the temperature. This controller is designed to handle two inputs, two outputs and 27 fuzzy rules; also inside and outside temperature can be monitored using LCD. The temperature inside the greenhouse can be set by the user as per the need of the crop in different seasons during the lifecycle of the plant. The PWM outputs are generated to control temperature according to the set point value.

Keywords - Intelligent Controller, FLC, Fuzzy Inference, Fuzzy Logic, Greenhouse, PWM

Introduction:

The greenhouse is a structure that is covered with a material that is transparent to the visible portion of the electromagnetic spectrum, which is utilized in the growth of plant. The performance of the greenhouse is best when temperature is not too hot and not too cold. It is necessary to maintain suitable temperature at growth stage of several plants. So, with the controlled environment in the greenhouse it is possible to increase the quality and quantity of crop produce per unit land in minimum possible time [1]. So, with the controlled environment in the greenhouse it is possible to increase the quality of crop produce per unit land in minimum possible time [1]. So, with the controlled environment in the greenhouse is very important for successful management of the greenhouse crops [2][3]. Fuzzy control has been widely applied in industrial controls and domestic electrical equipment [4]. The automatic learning of fuzzy rules is a key technique in fuzzy control. In the present work a fuzzy logic based temperature controller is designed which will sense the inside and outside temperature of greenhouse, displays it on the screen, allows user to set inside temperature as per the requirement and activate the relays accordingly so as to maintain temperature.



System Block Diagram2.1 Hardware

Atmel's 89C52 microcontroller was used for system design, which after initialization, reads the sensors, displays the inside and outside temperature values of on LCD and act accordingly as per the algorithm. National Semiconductor's LM 35 ICs were used as temperature



Vidya Bharati Shaikshanik Mandal's

VIDYA BHARATI MAHAVIDYALAYA, AMRAVATI

Re-accredited with Grade 'A' by the NAAC (CGPA 3.26 - Second Cycle) College with Potential for Excellence

In Collaboration with

S.S.S.K.R. INNANI MAHAVIDYALAYA, KARANJA (LAD) DIST. WASHIM Re-accredited by NAAC with Grade 'A' (CGPA 3.24 - Second Cycle) College with Potential for Excellence

UGC Sponsored National Conference on

Emerging Trends in Science

1st & 2nd February, 2019

CERTIFICATE

This is to certify that Prof./Dr./Mr./Ms. P. A. Saudagar

of Jankidevi Bajaj College of Science, Wardha

has actively participated in the Conference held on 1" & 2nd February, 2019 and has presented a research paper

oral / poster in the subject _____Electronics

entitled Design and Development of Microcontroller Based Low Cost System for CO2 Trapping in Greenhouse

during technical session of the conference.

Dr. R. V. Joat Organizing Secretary

Dr. P. R. Rajput Principal & Co-Convenor S.S.S.K.R.I. Mv., Karanja Lad

Dr. F. C. Raghuwanshi Principal & Convenor VBMV, Amravati

INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S

RESEARCH JOURNEY

International E-Research Journal

PEER REFREED & INDEXED JOURNALFebruary-2019Special Issue – 110 (H)

Electronics

Guest Editor :

Dr. F. C. Raghuwanshi

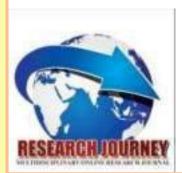
Principal,

- Vidya Bharati Mahavidyalya, Amarawati
- & Dean, Faculty of Science & Technology,
- Sant Gadge Baba Amravati University, Amravati

Executive Editors of the Issue :

- Dr.D. S. Dhote
- Professor & Head, Department of Electronics, Brijalal Biyani College, Amaravati
- Dr. R. A. Mishra
- Pircipal, Amolakchand Mahavidyala, Yeotmal
- Dr. Y. B. Gandule
- Principal, Adharsh Mahavidyala, Dhamangaon Rly.
- Dr. Bhimarao Ladgaonkar
- Professor & Head, Department of Electronics, Mohite Mahavidyala,Akluj Dist.Solapur Dr. P. B. Dahikar
 - Professor & Head, Department of Electronics ,Kamala Nehru Mahavidyala,Nagpur

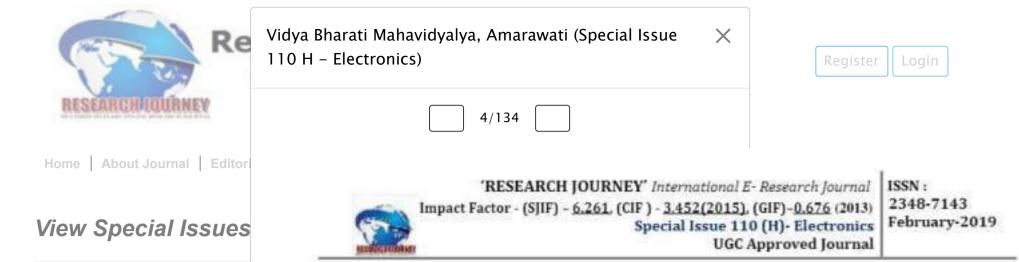
Chief Editor : Dr. Dhanraj Dhangar (Yeola)



This Journal is indexed in :

- University Grants Commission (UGC)
- Scientific Journal Impact Factor (SJIF)
- Cosmoc Impact Factor (CIF)
- Global Impact Factor (GIF)
- International Impact Factor Services (IIFS)





INDEX

No.	Title of the Paper Author's Name	Page No.
1	Virtual Instrumentation of Some Characterized Biomaterials Arun V. Padole, & Y.B. Gandole	05
2	A High Performance Cmos 1 Bit full Adder Ayushi S. Patankar	14
3	Efficient Smart Waste Management System With Multifunction Embedded Controller Pradeep B Dahikar	18
4	Characterists of Electronic Transducer in Biomedical Instruments Dr.Narendra B.Raut & Dr.Rammanohar A.Mishra	24
5	Use of AINN Filters for Reduction of Noise from ECG Signals R.J.Gajbe & Dr. Y.B.Gandole	30
6	Study of Open Source Tools and Technologies for Data Mining and Data Visualization Dr. Sumita U. Sharma,	37
7	Design and Development of Microcontroller Based Low Cost System for CO2 Trapping In Greenhouse P. A. Saudagar, D. S. Dhote, & G. A. Raut	41
8	Alternative Miracle Vehicle Gaurav.K.Dewale & Abhiram.M.Lomte	44
9	Advanced Sit and Reach Flexibility Measurement Test Box Using Arduino Sachin Winchurkar & Sudhakar Sarkate	47
10	Selection of Transfer Function for The Hidden Layer and Output Layer of MLP Network Y.M. Pharkade & Dr. G.D. Agrahari	51
11	U-Slotted Reconfigurable Multiband Micro Strip Antenna for Wireless Networks and SDR Sharanagouda N Patil , P.V Hunagund & R M Vani	56
12	Integrated High-Tech Intelligent Security System C.R.Chaudhari , S.V.Dudul & D.R.Solanke	61
13	Methods of Non-Invasive Blood Glucose Monitoring Using Nir Spectroscopy: A Review Nilima Jajoo, Dr. Deepak Dhote, & Dr. Gopal Agrahari	67
14	Microcontroller Based Techeye System for Obstacle Detection & Ranging To Assist Blind Person Yash Vidyasagar & Shri R.G.Chavan	72
15	Controlling Home Appliances Using Advanced Microcontroller: A Novel Approach Rameshwar S. Devhade, D.R. Solanke and S D Pachpande	74
16	Arduino Uno Based Accident Avoiding System IN Mountainous Area ACROSS U-Turn R. D. Chaudha & Dattaraj Vidyasagar	79
17	Green Electricity Response of Silver and Magnesium Electrode Pair Mr. G. S. Wajire & Dr. Y. B. Gandole	81
18	Design and Implementation of Fuzzy Logic Technique for Aircraft Control System K.Y. Rokde, P.B.Dahikar, S.S.Shende, & S.M.Ghatole	85
19	An Intelligent Controller for Greenhouse Temperature Control Using Fuzzy Logic P. A. Saudagar, D. S. Dhote, & G. V. Lakhotiya	92
20	Enhancement of Bandwidth and Reduction of Mutual Coupling in Microstrip Antenna Array K. Prahlada Rao, Vani R.M. & P.V. Hunagund	97
21	Evaluation of Mixed Multicast Architecture for Internet of Things Environment Using Adaptive Fountain Code Miss.Reshma Siddique & Dr.V.M.Thakare	107
	Cloud Computing With Big Data: Challenges & Issues	114

List of Article :

Select Year :

2019

2019	
Sr.	Date
51	4 February, 2019
52	4 February, 2019
53	4 February, 2019
54	4 February, 2019
55	4 February, 2019
56	4 February, 2019
57	4 February, 2019
58	5 February, 2019
59	5 February, 2019
60	5 February, 2019



Design and Development of Microcontroller Based Low Cost System for CO₂ Trapping In Greenhouse

P. A. Saudagar, Jankidevi Bajaj College of Science, Wardha, India, saudagar.pa@gmail.com D. S. Dhote, Brijlal Biyani Science College, Amravati, India, dsdhote@rediffmail.com G. A. Raut Brijlal Biyani Science College, Amravati, India, rautgajanan10@gmail.com

Abstract –

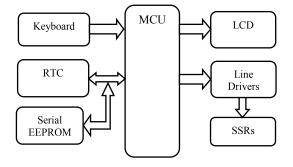
The scope of this paper includes the design and development of a low cost system for CO_2 trapping in greenhouse using microcontroller. The hardware of the system consists of interactive user interface, which allows the user to enable or disable the trapping system. The system also supports digital clock designed using IC PCF8583 clock/calendar catering the need of a clock for the system. The software was developed for the system for user interface which includes the keyboard, display and processing the settings made by the gardener and generating the output accordingly at different port lines.

Keywords: - Greenhouse, Microcontroller, RTC, SSR.

Introduction:

The classical definition of greenhouse is a structure that is covered with a material that is transparent to the visible portion of the electromagnetic spectrum, which is utilized in the growth of plant Automation in greenhouse is very important for successful management of the greenhouse crops [1]-[3]. Carbon Dioxide (CO₂) is essential for photosynthesis. The CO₂ released by the plants and soil during night could be trapped and hence CO₂ level could be increased by 6-7 times and photosynthetic activity by 3-4 times [4]. Normally air contains about 300ppm of CO₂. Studies have shown that levels of 1000 to 1500 ppm are very beneficial for good yields [5]. Therefore, the economical method to achieve this is designed to close all openings of the greenhouse in the late afternoon and open them in the morning automatically.

1. System Block Diagram



2. Hardware 3.1. The MCU

Atmel's 89C52 IC was used to design the system hardware to which 12MHz crystal was connected for generation of system clock. The port 2 pins were connected to the line driver IC ULN2803, which was used to drive the Solid State Relays (SSRs) to which mechanical system may be connected to drive doors and vents of greenhouse. The port1 pins P1.0, P1.1 and P1.2 were used for the keyboard design. Audible alerts were generated using buzzer connected to port 1 pin P1.3. Port pins P1.4 and P1.5 were used to interface the serial devices. P1.4 was defined as the serial Data line and P1.5 was used as the clock signal to the serial devices.

3.2. Keyboard and Display

A simple keyboard consisting of three keys was designed for the system which could be used to set clock and to start or stop trapping mechanism. The limited number of keys reduces the complexity in the operation. A 20x4 LCD display module was used to display the current time and also to show interactive messages for the user during the settings. This module has four rows of twenty characters in each row.

3.3. Real Time Clock and EEPROM

The system uses I^2C bus for integrated clock/calendar IC PCF 8583P by Philips semiconductor and serial EEPROM. The clock/calendar IC was very helpful in the current system which was used to control the mechanical system at proper time for trapping CO₂. The minutes and hours were used in the system, which may be modified as per the local time. The EEPROM IC24C02 was used in the design to store the settings made by the user. This facility avoids the need of making frequent settings in case of power failure and system resumes its functioning as per the previous settings when power is turned on. Both, RTC and EEPROM are serial devices which reduces the hardware and ultimately the power consumption [6].

3.4. Line drivers and SSRs

To control the mechanical system used for doors and vents of greenhouse as per the software, The single phase solid state relays (SSR) 006 JDA 330705 by ERI having rms on state current of 7 Amperes were used. The output port lines were first connected to the line driver IC ULN2803, which is designed to be compatible with standard TTL families, which drives the solid state relays (SSRs).

4. Software

Three main software modules were developed for the system. The modular programming approach was used so that individual modules could be upgraded as and when needed [6]. These modules are:

- i. The initialization Module,
- ii. The Keyboard and Display Module, and
- iii. CO₂ Trapping Module

The initialization module, prepares the system for the normal operation. This module first defines the variables and initiates them to their default values. Reads the settings from the serial EEPROM and stores them to corresponding location. It also initiates the LCD display by sending the commands and displays the welcome message on it then displays the current time. The keyboard and display module scans the keyboard and detects the key pressed by the user and the display the related message on the LCD. If user wishes to change the current time, it could be done with the help of proper keys on the keyboard. Intelligence was developed in the system

such that if user does not press any key, the system remains unaffected and the current information from the RTC was used. But, in case the user modifies the minutes or the hour values, it will be immediately loaded to the corresponding locations in the RTC, which proceeds with the new values. The CO_2 module, if activated, generates the signal at 5 p.m., which may be used to drive a mechanical system which close all the doors and vents of the greenhouse so that CO_2 can be trapped and accumulated and in the morning, the system deactivates the signal at 8 a.m. so that the mechanical system can now open all the doors and vents of the greenhouse. This will enhance the CO_2 level. The signal generated by the controller remains activated between 5 p.m. to 8 a.m. and during this period complete vents of greenhouse will be closed. Here, the signal was kept continuously alive for the required duration so that in case of power failure, if trapping mechanism was activated, then system could take the action when the power resumes. The Control signals generated by microcontroller were used to drive the solid state relays through line drives. The mechanical system, to open and close greenhouse vents, needs to be designed and would be driven by these solid state relays.

Result And Discussion:

Respiration by the plant at night causes the Carbon dioxide levels to increase to around 500 ppm by early morning. The plant uses this high level in the first two or three hours after sunrise causing the levels to drop to around 150 ppm. At this stage the doors and vents could be opened to allow outside fresh air and the levels then rise back to around 300 ppm [4]. The actual test of this system was conducted by simulation, where the system clock time was changed and system actions were verified for the signal generation for the mechanical system. The controller generated the signals only when the CO_2 trapping was started. If user stops the CO_2 trapping, the system immediately deactivates the signal generated for the mechanical system. The functioning of this mechanism was tested by having different setting pertaining to clock and CO_2 trapping mode.

References

- **1.** A. Sriraman and R. V. Mayorga, A Fuzzy Inference System Approach for Greenhouse Climate Control, Environmental Informatics Achieves, Vol. 2(2004),pp699-710.
- **2.** ZHOU Xiaobo, WANG Chengduan and LAN Hong, The Research and PLC Application of Fuzzy Control in Greenhouse Environment, 2009 Sixth International Conference on Fuzzy Systems and Knowledge Discovery, pp340-344.
- **3.** R. Caponetto, L. Fortuna, G. Nunnari and L. Occhipinti, A Fuzzy Approach to Greenhouse Climate Control, Proceedings of the American Control Conference Philadelphia, Pennsylvania, June 1998, pp1866-1870.
- **4.** Shinde, P. P., and D. G. Hapse. Greenhouse Technology. Treatise on Agrophysics and Agrielectronics, Vasantdada Sugar Institute, Manjari (BK), Pune, Manjari (BK), Pune: Vasantdada Sugar Institute, 1993, 27.1-27.10.
- **5.** Roberts, W. J. "Horticulture Engineering Center for Controlled Enironmental Agriculture." RUTGERS School of Environmental and Biological Sciences.2005. aesop.rutgers.edu/~horteng/FactSheets/EnvContrlofGHs.pdf (accessed 8 7, 2012).
- **6.** Saudagar, P. A., Fuzzy and Integrated Fuzzy Logic Controller for Greenhouse Climate Control (Unpublished doctoral thesis). Sant Gadge Baba Amravati University, Amravati, India (2013).

I N T E R N INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S RESEARCH JOURNEY	43
R INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S	
A RESEARCH INTRNEV	
T RESEARCH JOURNET	
International E-Research Journal	
0 PEER REFREED & INDEXED JOURNAL	
N February-2019 Special Issue – 110 (E)	
Α	
ΒΟΤΟΝΥ	
R	
Ε	
S	
 E Guest Editor: A Dr. F. C. Raghuwanshi 	
Principal,	
 Vidya Bhrati Mahavidyalaya, Amaravati Executive Editor of the issue: 	
H Dr. P. G. Bansod	
Dr. M. U. Ghurde Dr. P. V. Pulate	
E Ms. Lubna Khalid	
L Chief Editor:	
L Dr. Dhanraj Dhangar (Yeola)	
0	
W	
S	
Α	
S	
S	
O C	
I This Journal is indexed in : - University Grants Commission (UGC)	
A Scientific Journal Impact Factor (SJIF)	
T - Cosmoc Impact Factor (CIF) - Global Impact Factor (GIF)	
I International Impact Factor Services (IIFS)	
O N For Details Visit To : www.researchjourney.net Swatidhan Juli	LICATIONS



 'RESEARCH JOURNEY' International E- Research Journal
 IS

 Impact Factor - (SJIF) - 6.261, (CIF) - 3.452(2015), (GIF) - 0.676 (2013)
 23

 Special Issue 110 (E) - Botony
 Feedback



UGC Approved Journal

ISSN : 2348-7143 February-2019

45	Campus Flora of Art and Science College Pulgaon- Nachangaon District-Wardha (Maharashtra) Ajay B.Jadhao, & Aboli A. kshirsagar & Dipti B.Kadu	264
46	Antifungal Activity of Two Medicinal Plants Against Some Selected Fungi Monali Ughde & Shital Tripathi & Pratibha Dhabarde	270
47	Study of Aeromycoflora in Mandev Garden Yavatmal Dr. Ninad Dharkar	274
48	Diversity of Aeromycoflora from Indoor Environment of Hostel Kalyani Wasurkar, Swati kalode,Dr.Lalchand Dalal	276
49	Review of Traditional and Phytochemical Investigations of Essential oil Yielding Plant Pelargonium Graveolens Prof. Mrs. Vaishali N. Badgujar	280
50	Study of Different Mitotic Abnormalities Induce by Ems in Dianthus Caryophyllus Var. Chabaud Deshmukh P.D. And S.N. Malode	284
51	Taxonomic Study on Plants of Malvaceae Reeta Satone, Pratibha Dhabarde,Swati kalode	295
52	In Vitro Antioxidant Activity of Clerodendrum Phlomidis Linn. Verbanaceae Sonali D. Suple1 and Varsha D. Hutke1	299
53	Seed Surface Characteristics and Preliminary Phytochemical Analysis of Pimpinella Anisum Linn. Seeds of Apiaceae (Umbelliferae) Ulhe P.P.	304
54	Phylogenetic Relationship Between Bambusoideae-Pooideae Complex Based on Plastid and Nuclear Genome Markers Ashiq Khanday & Prashant Gawande & Irfan Badroo & , Wagay N.A.	316
55	Effect of Drinking Water on Potential Kidney Stone from Sangrampur Region of Buldana District, Maharashtra Dhammapal L. Bhade & R.E.Khadsan	323
56	Usef A Prototype Biomass Fired Gasifier Stove: Key To Reduce Pollution Er. Mangesh D. Ghungrud & Er. Sushant Bakal & Er. H. Y. Shrirame	327
57	Water Quality Index Assessment of Khekara Nullah Dam, Nagpur B. S. Tapase and J.L.Tarar	333
58	Eco-Friendly Disposal of Pesticide Remenants From Utensiles Kamalakar K. Wavhal and S. B. Borul	339
59	Pollution Ecology Its Affects and Solution Ku. Sima Hari Kothalkar and Mr. Yogesh Bhaskarrao Hage	343
60	Review of Wastewater Treatment and its Reuse Kirti Kalbande and Jayashree Dhote	348
61	Land Use/Land Cover Mapping of Amravati Taluka (Maharashtra) Using Gis and Remote Sensing Techniques Sonone K. H., Ingole S.P., Kakde A. U.	352
62	Potential Health Impact of Hard Water - A Case Study of Bhatkuli Taluka, District Amravati Saleha Ahmad, Kakde A U , Ingole S P	356
63	Effect of Chloride in Pond Water Sample in Nerul, Navi Mumbai Yashodhara Varale	361
64	Review- Role of EIA in Maintenance of Railway Station Badase A. S., Ingole S P	364
65	A Study on Performance of Shgs of Organic Farming And Dairy owners P. G. Chaudhari, S. S. Thakare,	368
66	Economic Analysis of Production of Gerbera (Cut Flower) Under Protected Condition in Amravati District Shelake P.N., D.H. Ulemale ,Bochare K.V and Nagre K.G	374
67	Temporal Changes in Input-Output Prices And Cost of Cultivation of Soybean in Vidarbha S. A. Borde, S. S. Thakare, P. G. Awagan and Ku. V. A. Deshmukh	380



Diversity of Aeromycoflora from Indoor Environment of Hostel

Kalyani Wasurkar, Swati kalode,Dr.Lalchand Dalal Department of Botany, J.B.College of Science, Wardha

Abstract:

An aeromycological study on indoor environment of hostel area was conducted during the month of dec-jan2018-19 which shows the variation in numbers as well as percentage of fungal species in these area. The study shows highest percentage contribution of Cladosporium spp(53%)on PDA and Czapeck-dox medium. The rest of the fungi like Aspergillus, Fusarium and Helminthosporium spp shows minimum percentage contribution.

Keywords: Aeromycoflora, PDA, CZA, Hostel.

Introduction :

Fungus is a member of the group of eukaryotic organisms that includes micro – organism such as yeast and molds as well as mushrooms. Fungi is a achlorophyllous organisms and they do not take part in photosynthesis hence it is called heterotroph. Fungi play and important role to decomposition of organic matter. Some fungi is mycotoxin and some are used for the production of antibiotic.

Aerobiology is deals with the studies of microrganisms present in the air. Meir (1930) was the first aerobiologists who used the term aerobiology for the studies of airborne fungal spores and their – organisms. The fungal species such as Aspergillus, Fusarium and Cladosporium is abundantly found in air .Fungi are common in indoor and outdoor environment, and nearly 10% of people worldwide have fungal allergy(Burge H.A2001). Epidemiological studies showed that to high concentration of micro – organism in the air can be allergic however even very low concentration of some particular micro – organism can cause serious disease. Singh and his coworkers are actively engaged in the field of aerobiology using various technique with reference to airspora inside hostel kitchen, cowshed etc.Climate play an important role in growth of fungi. The aim of the study is totally based on to isolate the fungal spores from indoor environment of hostel area, their prevalence and their impact on human being.

Material and methods:-

The present study deals with the Aeromycoflora isolated from the indoor environment of hostel area from J. B. College of science, Wardha i.e. ground floar, first floar and terrace. The fungi isolated on two different types of media i.e PDA and CZA media. 2 Petriplates of freshly prepared potato dextrose agar media and czapeck-dox media were exposed at each site for 10 min in the morning and evening. Then temperature is recorded. After exposure, the plates were incubated at $27 \circ C$ + upto 3-4 days in incubator. The slides were prepared with the help of Lactophenol cotton blue stain and identified using certain literatures. (Bernett and hunter, 1972; cooke1963; Tilak 1989; Kalbende etal2012)

Result and Discussion:

The present work based on collection and identification of different fungi from three sites of hostel i.e. ground floar, first floar and Terrace.



I N T E R N INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S RESEARCH JOURNEY	43
R INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S	
A RESEARCH INTRNEV	
T RESEARCH JOURNET	
International E-Research Journal	
0 PEER REFREED & INDEXED JOURNAL	
N February-2019 Special Issue – 110 (E)	
Α	
ΒΟΤΟΝΥ	
R	
Ε	
S	
 E Guest Editor: A Dr. F. C. Raghuwanshi 	
Principal,	
 Vidya Bhrati Mahavidyalaya, Amaravati Executive Editor of the issue: 	
H Dr. P. G. Bansod	
Dr. M. U. Ghurde Dr. P. V. Pulate	
E Ms. Lubna Khalid	
L Chief Editor:	
L Dr. Dhanraj Dhangar (Yeola)	
0	
W	
S	
Α	
S	
S	
O C	
I This Journal is indexed in : - University Grants Commission (UGC)	
A Scientific Journal Impact Factor (SJIF)	
T - Cosmoc Impact Factor (CIF) - Global Impact Factor (GIF)	
I International Impact Factor Services (IIFS)	
O N For Details Visit To : www.researchjourney.net Swatidhan Juli	LICATIONS



'RESEARCH JOURNEY' International E- Research Journal **ISSN**: Impact Factor - (SJIF) – <u>6.261</u>, (CIF) - <u>3.452(2015)</u>, (GIF)–<u>0.676</u> (2013) 2348-



LIF) - <u>3.452(2015)</u>, (GIF)-<u>0.676</u> (2013) Special Issue 110 (E) - Botony

2348-7143 February-2019

UGC Approved Journal

Estimation of Input Demand and output Supply of Sorghum 22 125 S. S. Thakare N. V. Shende And S. N. Ingle Diversity of Members of Family Asteraceae in Melghat from Amravati District 23 132 (M.S.), India ManjushaWath* ,MayuriKathalkar and PoojaMahalle Anacardiospermum Deccanensis Gen.Et.Sp.Nov. A Report of New Fossil Seed from Deccan Intertrappen Beds of Mohgaonkalan, M.P., India. 24 138 Dighe S. W.1 & Kokate P. S.2 Phytoplanktondiversity of adan Reservoir of Washim District 25 142 Ghude, R.S.; Halwe, D.R. Gas Chromatography and Mass Spectroscopy Study of oil Extracted from Some 26 146 Poaceae Family Plants Sambhaji S. Gawali and Shrusti S. Khandare Effect of Ph on Growth of Insect Lac Fungi 27 Mayuri Bhowate & D.U. Gawai 152 Outdoor Aerospora Study from Play Ground of Jbcs College, Wardha 28 156 Swati Kalode & Dr. Lalchand Dalal Preliminary Phytochemical observations of Tinosporacordifolia (Willd) Miers. 29 160 Ashwini Sirsat1, Rupali Shirsat2, Pratiksha Kokate1 and Deepak Koche1 Phenotypic Variation and The Relationships Among 9 Genotypes of Brassica 30 166 Campestris L. and Their Application for Dustesting N. S. Hinge and S. N. Malode Estimation of Phenolic Compounds By Spectrophotometric Method from Fruits of 31 173 Cordia Dichotoma Forst Poonam R. Gulhane and K. D. Jadhao. Improvement of Groundnut (Arachis Hypogaea L.) Through Chemical Mutagen 32 177 (Ems). Suradkar S. W. Medico-Ethno Botany of Some Medicinally Important Plants from Melghat Tiger 33 183 Reserve Dist. Amravati. (Ms) India Mangesh Baliramji Bobade Specimen Browser System - an Image Based Tool for Accessing Digitized Botanical 34 186 Collections Ranjan B. Kalbande Biosorption of Nickel by The Aquatic Plant Ipomea Aquatica 35 194 N.S.Gopkar & U.S.Patil Conservation of Wild Edible Plants in India To Combat Future Challenges 36 204 Savita Borse & Nikhila Bhagwat Herbal Medicine for The Snake Bite Treatment By The Korku Tribals of Melghat 37 213 Region (Ms) India Nitin A. Khandare, Pornima D. Malviya. Observations on Important Pharmacognostic Characters of An Ethno-Medicinal 38 215 Plant Spilanthes Calva Dc Malode U. G & Belsare S.D. Priliminary Phyotochemical Screening of Asystasia Gangetica(L.)Anders. 39 223 Kothale K. V., Thakur S.B., Wankhade M.R. and Atram P.W. Cytotoxic Properties of Curcuma Inodora Leaf Against (Miapaca-2) Human **40** 228 Pancreatic Carcinoma Cell Line M.U.Ghurde and S.N.Malode Investigations on Morphological Variations and Mitotic Index in Lilium L. Cultivars 41 233 Deshmukh S. K. and Nathar V. N. Natural Pollinators and Their Effect on Yield of Sesamum Indicum L. 42 242 P. J. Kale & J.A. Tidke and S. S. Rokade Seasonal Water Quality Assessment of Shahanoor Dam, Anjangaon Surji, District 43 Amravati (M.S.) India By Using Multivariate Analysis and Water Quality Index 249 S.R.Bansod & N.S.Gopkar & U.S.Patil (Wqi) Effect of Cyanobacteria and Mycorrhizal Biofertilizer for Sustainable Crop 44 259 Production in Cicer Arietinum Dr. Pradhnya Khapekar

Outdoor Aerospora Study from Play Ground of Jbcs College, Wardha

Swati Kalode, Dr. Lalchand Dalal Department of Botany, J.B.College of Science, Wardha

Abstract:

Air does not act as a natural environment for the growth and multiplication of Aero mycoflora, but it act as a very good medium for their dispersal from one place to another. Aeromycoflora of playground constitute fungal spores abundance and frequency in the environment. Aeromycoflora simply refers to the airborne fungal contributors of the environment. In air, all types of microorganisms are present like bacteria and fungi, spores are disperse in air from different sources and causes many diseases to human being and animals. Environmental factors like temp, and humidity play and important role for distribution of fungal spores. Air sampling was done at monthly intervals by using petriplate method for isolation of airborne fungi. Aspergillus niger, A. flavus, A. fumigatus & Cladosporium cladosporoides were more dominant fungi than other rest form of fungi. The present investigation shows that 27 fungal spores were recorded during the study period. Cladosporium cladosporoides was the highest percentage (24.16%) followed by Aspergillus and Fusarium spp.

Keywords: Aeromycoflora, Playground, PDA, Seasonal variations, Fungal spores.

Introduction:

Air borne fungi which are found in all types of environment. They are found largely in Indoor and outdoor environment. Earth atmosphere contain propagules of diverse group of microbes and other particle of biological origin (Rajendra et al 2017). The abundance of fungal spores are dependent upon many biotic and abiotic factors including temperature, humidity and sudden environmental changes, thus the airborne microorganism of any environment is specific in nature. Airborne microbe is a component of our environment and it is a potential economic and health implications (Gregory, 1961; Hashimoto, 1986). The aim of this study was to determine the Aeromycoflora and their identification, seasonal distribution of the airborne cultivable fungi in the air of a playground associated environments at outdoor in order to evaluate.

Materials And Methods :

For isolation of Aeromycoflora, Potato Dextrose Agar culture medium was used to isolate the variety of fungal spores from the environment. Aeromycoflora were studied from the play ground of JBCS Wardha. Culture plate exposure method containing PDA media were used for isolation of mycoflora present in the outdoor environment. This method also used by Tiwari P. (2008) for survey of aeromycoflora. The 10 cm diameter size petriplate were exposed twice in a day . The exposed petriplate were brought into the laboratory and incubated at 27 ± 1 °C for 4-5 days. At the end of incubation period the fungal colonies were counted and fungal spores identification were made by using lacto-phenol and cotton blue stain. Identification were done on the basis of colourization of colonies and their morphological characterization and also using available literature (Barnett, 1969; Nigmani et al. 2006). Temperature and Humidity were recorded in the college campus during the sampling period using a Hygrometer.

Result And Discussion:

The study deals with the exploration of Aeromycoflora from Playground of JBCS college which are situated at centre of the college. During this study a no of fungal propagules are encountered through a PDA media using gravity plate exposure methods. It shows that the variation in number of fungal species in this area. The Identified fungal spores belonging to



Vidya Bharati Shaikshanik Mandal's VIDYA BHARATI MAHAVIDYALAYA, AMRAVATI Re-accredited with Grade 'A' by the NAAC (COTH CH. AMRAVAT) In Collaboration with S.S.S.K.R. INNANI MAHAMOLOGICAL COMPANY AND College with Potential for Excellence UGC Sponsored National Conference on

Emerging Trends in Science 1st & 2nd February, 2019

CERTIFICATE

This is to certify that Prof./Dr./Mr./Ms. Pratibha Dhabarde

of J, B, College of Science, Wardha

has actively participated in the Conference held on 1" & 2" February, 2019 and has presented a research paper

oral / poster in the subject Botany

entitled Antifungal Activity of Two Medicinal Plants against Some Selected Fungi.

during technical session of the conference.

Dr. R. V. Joat Organizing

Dr. P. R. Rajput Principal & Co-Convenor S.S.S.K.R.I. My, Karania Lad

Dr. F. C. Raghuwanshi Principal & Convenor VBMV, Amravati

I N T E	Impact Factor – 6.261 ISSN – 2348-7143
R N	INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S
A T	RESEARCH JOURNEY
I	International E-Research Journal
0	PEER REFREED & INDEXED JOURNAL
Ν	February-2019 Special Issue – 110 (E)
A	
L	BOTONY
R	
E	
S E	
E A	Guest Editor: Dr. F. C. Raghuwanshi
R	Principal, Vidya Bhrati Mahavidyalaya, Amaravati
C	Executive Editor of the issue:
H	Dr. P. G. Bansod
F	Dr. M. U. Ghurde Dr. P. V. Pulate
E	Ms. Lubna Khalid
L	Chief Editor:
L	Dr. Dhanraj Dhangar (Yeola)
0	
W	
S	
Α	
S	
S O	
0 C	
I	This Journal is indexed in : - University Grants Commission (UGC)
A	- Scientific Journal Impact Factor (SJIF)
Т	 Cosmoc Impact Factor (CIF) Global Impact Factor (GIF)
I	- International Impact Factor Services (IIFS)
0 N	For Details Visit To : <u>www.researchjourney.net</u>



 'RESEARCH JOURNEY' International E- Research Journal
 IS

 Impact Factor - (SJIF) - 6.261, (CIF) - 3.452(2015), (GIF) - 0.676 (2013)
 23

 Special Issue 110 (E) - Botony
 Feedback



UGC Approved Journal

ISSN : 2348-7143 February-2019

45	Campus Flora of Art and Science College Pulgaon- Nachangaon District-Wardha (Maharashtra) Ajay B.Jadhao, & Aboli A. kshirsagar & Dipti B.Kadu	264	
46	Antifungal Activity of Two Medicinal Plants Against Some Selected Fungi Monali Ughde & Shital Tripathi & Pratibha Dhabarde		
47	Study of Aeromycoflora in Mandev Garden Yavatmal Dr. Ninad Dharkar		
48	Diversity of Aeromycoflora from Indoor Environment of Hostel Kalyani Wasurkar, Swati kalode,Dr.Lalchand Dalal		
49	Review of Traditional and Phytochemical Investigations of Essential oil Yielding Plant Pelargonium GraveolensProf. Mrs. Vaishali N. Badgujar280		
50	Study of Different Mitotic Abnormalities Induce by Ems in Dianthus Caryophyllus Var. Chabaud Deshmukh P.D. And S.N. Malode		
51	Taxonomic Study on Plants of Malvaceae Reeta Satone, Pratibha Dhabarde,Swati kalode		
52	In Vitro Antioxidant Activity of Clerodendrum Phlomidis Linn. Verbanaceae Sonali D. Suple1 and Varsha D. Hutke1	299	
53	Seed Surface Characteristics and Preliminary Phytochemical Analysis of Pimpinella Anisum Linn. Seeds of Apiaceae (Umbelliferae) Ulhe P.P.	304	
54	Anisum Linn. Seeds of Apiaceae (Umbelliferae) Uline P.P. Phylogenetic Relationship Between Bambusoideae-Pooideae Complex Based on Plastid and Nuclear Genome Markers 316 Ashiq Khanday & Prashant Gawande & Irfan Badroo & , Wagay N.A.		
55	Effect of Drinking Water on Potential Kidney Stone from Sangrampur Region of Buldana District, Maharashtra Dhammapal L. Bhade & R.E.Khadsan 323		
56	Usef A Prototype Biomass Fired Gasifier Stove: Key To Reduce Pollution Er. Mangesh D. Ghungrud & Er. Sushant Bakal & Er. H. Y. Shrirame		
57	Water Quality Index Assessment of Khekara Nullah Dam, Nagpur 333 B. S. Tapase and J.L.Tarar		
58	Eco-Friendly Disposal of Pesticide Remenants From Utensiles Kamalakar K. Wavhal and S. B. Borul		
59	Pollution Ecology Its Affects and Solution Ku. Sima Hari Kothalkar and Mr. Yogesh Bhaskarrao Hage		
60	Review of Wastewater Treatment and its Reuse Kirti Kalbande and Jayashree Dhote	348	
61	Land Use/Land Cover Mapping of Amravati Taluka (Maharashtra) Using Gis and Remote Sensing Techniques Sonone K. H., Ingole S.P., Kakde A. U.	352	
62	Potential Health Impact of Hard Water - A Case Study of Bhatkuli Taluka, District Amravati Saleha Ahmad, Kakde A U , Ingole S P	356	
63	Effect of Chloride in Pond Water Sample in Nerul, Navi Mumbai Yashodhara Varale	361	
64	Review- Role of EIA in Maintenance of Railway Station Badase A. S., Ingole S P 364		
65	A Study on Performance of Shgs of Organic Farming And Dairy owners P. G. Chaudhari, S. S. Thakare, 368		
66	Economic Analysis of Production of Gerbera (Cut Flower) Under Protected 374 Condition in Amravati District 374 Shelake P.N., D.H. Ulemale ,Bochare K.V and Nagre K.G 374		
67	Temporal Changes in Input-Output Prices And Cost of Cultivation of Soybean in Vidarbha S. A. Borde, S. S. Thakare, P. G. Awagan and Ku. V. A. Deshmukh	380	

Antifungal Activity of Two Medicinal Plants Against Some Selected Fungi

Monali Ughde, Shital Tripathi , Pratibha Dhabarde Post- Graduate Department of Botany, Bajaj College Of Science, Wardha Email id-pratibhadhabarde@gmail.com

Absract:

The aim of the study was totally based on evaluation of antifungal activity using plant extract against some selected fungi. The plants were selected on the basis of their reported ethnobotanical uses. Ethanolic extraction were done with the help of Soxhlet appratus by using dried leaves powdered of two medicinally important plants i.e Vitex negundo and Holarrhena antidysentrica .Well diffusion method were used to inhibit the growth of fungi on PDA media. Two plants extract shows the minimum inhibitory growth of fungi on PDA media.

Keywords: Antifungal, Fungi, Medicinal Plants, Solvent extract.

Introduction:

Pathogenic fungi are the main allergic agents in plants causing alterations during developmental stage including post-harvest to mankind. It will not be an exaggeration to say that use for herbal drug for human healthcare is probably as ancient as mankind. Medicinal plant represent rich source of antimicrobial agents (Mahesh and Satish, 2008). Plant extracts have been developed and proposed for use as antimicrobial substance (Del et al., 2000) Vitex negundo :(family- Verbenaceae). The genus consist of 250 species of which about 14 species are found in India and have some commercial and medicinal importance . Vitex negundo Linn; commonly known as five-leaved choste tree are mank's pepper is used as medicine fairly throughout the greater part of the India (Chopra et al., 1956). The Vitex negundo plant is a large aromatic shrub or sometimes a smaller slender tree and quadrangular, densely whitish branchlets up to 4.5 -5.5m in height. Plants is bitter, antiseptic, asthma and enlargement of spleen. Leaves contain an alkaloids nishindine and other constituents like vitamin C, carotene, benzoic acid (Husain, 1992).

Holarrhena antidysentrica: (family-Apocynaceae) commonly known as bitter oleander and locally as "inderjotulkh" (Baquar, 1989). Seeds are 1-2cm long, linear or oblong concave with a coma shaped. The plant is use traditionally for a health disorders including Diarrhea ,dysentery (Jain, 1991). Alkaloids have been reported in the leaf of plant.

Material and Method:

Plant Collection: The plant were collected from the Dhaga forest, Wardha . Sterilization of plant material: The disease free and fresh plants were selected. The plant parts were washed with distilled water for three times. Then surface sterilized with 0.1% Mercuric chloride for 20 second. Again the leaves were washed thoroughly with distilled water (three time).

Preparation of plant extract: The collected plant material was air dried at room temperature for 10 to 15 days. The dried plants material was crushed by mortar and pestle without adding any solvent into it . the powder material was kept in air tight glass bottles. This stock powder was added in 150 ml solvent (Ethanol) by using a soxhlet extract for minimum 8-9





Vidya Bharati Shaikshanik Mandal's

VIDYA BHARATI MAHAVIDYALAYA, AMRAVATI

Re-accredited with Grade 'A' by the NAAC (CGPA 3.26 - Second Cycle) College with Potential for Excellence

In Collaboration with

S.S.S.K.R. INNANI MAHAVIDYALAYA, KARANJA (LAD) DIST. WASHIM Re-accredited by NAAC with Grade 'A' (CGPA 3.24 - Second Cycle) College with Potential for Excellence

UGC Sponsored National Conference on

Emerging Trends in Science

Botany

1st & 2nd February, 2019

CERTIFICATE

This is to certify that Prof./Dr./Mr./Ms. Reeta Satone

of _____J. B. College of Science, Wardha

has actively participated in the Conference held on 1" & 2" February, 2019 and has presented a research paper

oral / poster in the subject

entitled Taxonomic Study On Plants Of Malvaceae

during technical session of the conference.

Dr. R. V. Joat Organizing Secretary

Dr. P. R. Rajput Principal & Co-Convenor S.S.S.K.R.I. Mv., Karanja Lad

Dr. F. C. Raghuwanshi Principal & Convenor VBMV, Amravati

I N T E	Impact Factor – 6.261 ISSN – 2348-7143
R N	INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S
A T	RESEARCH JOURNEY
I	International E-Research Journal
0	PEER REFREED & INDEXED JOURNAL
Ν	February-2019 Special Issue – 110 (E)
A	
L	BOTONY
R	
E	
S E	
E A	Guest Editor: Dr. F. C. Raghuwanshi
R	Principal, Vidya Bhrati Mahavidyalaya, Amaravati
C	Executive Editor of the issue:
H	Dr. P. G. Bansod
F	Dr. M. U. Ghurde Dr. P. V. Pulate
E	Ms. Lubna Khalid
L	Chief Editor:
L	Dr. Dhanraj Dhangar (Yeola)
0	
W	
S	
Α	
S	
S O	
0 C	
I	This Journal is indexed in : - University Grants Commission (UGC)
A	- Scientific Journal Impact Factor (SJIF)
Т	 Cosmoc Impact Factor (CIF) Global Impact Factor (GIF)
I	- International Impact Factor Services (IIFS)
0 N	For Details Visit To : <u>www.researchjourney.net</u>



 'RESEARCH JOURNEY' International E- Research Journal
 IS

 Impact Factor - (SJIF) - 6.261, (CIF) - 3.452(2015), (GIF) - 0.676 (2013)
 23

 Special Issue 110 (E) - Botony
 Feedback



UGC Approved Journal

ISSN : 2348-7143 February-2019

45	Campus Flora of Art and Science College Pulgaon- Nachangaon District-Wardha (Maharashtra) Ajay B.Jadhao, & Aboli A. kshirsagar & Dipti B.Kadu	264	
46	Antifungal Activity of Two Medicinal Plants Against Some Selected Fungi Monali Ughde & Shital Tripathi & Pratibha Dhabarde		
47	Study of Aeromycoflora in Mandev Garden Yavatmal Dr. Ninad Dharkar		
48	Diversity of Aeromycoflora from Indoor Environment of Hostel Kalyani Wasurkar, Swati kalode,Dr.Lalchand Dalal		
49	Review of Traditional and Phytochemical Investigations of Essential oil Yielding Plant Pelargonium GraveolensProf. Mrs. Vaishali N. Badgujar280		
50	Study of Different Mitotic Abnormalities Induce by Ems in Dianthus Caryophyllus Var. Chabaud Deshmukh P.D. And S.N. Malode		
51	Taxonomic Study on Plants of Malvaceae Reeta Satone, Pratibha Dhabarde,Swati kalode		
52	In Vitro Antioxidant Activity of Clerodendrum Phlomidis Linn. Verbanaceae Sonali D. Suple1 and Varsha D. Hutke1	299	
53	Seed Surface Characteristics and Preliminary Phytochemical Analysis of Pimpinella Anisum Linn. Seeds of Apiaceae (Umbelliferae) Ulhe P.P.	304	
54	Anisum Linn. Seeds of Apiaceae (Umbelliferae) Uline P.P. Phylogenetic Relationship Between Bambusoideae-Pooideae Complex Based on Plastid and Nuclear Genome Markers 316 Ashiq Khanday & Prashant Gawande & Irfan Badroo & , Wagay N.A.		
55	Effect of Drinking Water on Potential Kidney Stone from Sangrampur Region of Buldana District, Maharashtra Dhammapal L. Bhade & R.E.Khadsan 323		
56	Usef A Prototype Biomass Fired Gasifier Stove: Key To Reduce Pollution Er. Mangesh D. Ghungrud & Er. Sushant Bakal & Er. H. Y. Shrirame		
57	Water Quality Index Assessment of Khekara Nullah Dam, Nagpur 333 B. S. Tapase and J.L.Tarar		
58	Eco-Friendly Disposal of Pesticide Remenants From Utensiles Kamalakar K. Wavhal and S. B. Borul		
59	Pollution Ecology Its Affects and Solution Ku. Sima Hari Kothalkar and Mr. Yogesh Bhaskarrao Hage		
60	Review of Wastewater Treatment and its Reuse Kirti Kalbande and Jayashree Dhote	348	
61	Land Use/Land Cover Mapping of Amravati Taluka (Maharashtra) Using Gis and Remote Sensing Techniques Sonone K. H., Ingole S.P., Kakde A. U.	352	
62	Potential Health Impact of Hard Water - A Case Study of Bhatkuli Taluka, District Amravati Saleha Ahmad, Kakde A U , Ingole S P	356	
63	Effect of Chloride in Pond Water Sample in Nerul, Navi Mumbai Yashodhara Varale	361	
64	Review- Role of EIA in Maintenance of Railway Station Badase A. S., Ingole S P 364		
65	A Study on Performance of Shgs of Organic Farming And Dairy owners P. G. Chaudhari, S. S. Thakare, 368		
66	Economic Analysis of Production of Gerbera (Cut Flower) Under Protected 374 Condition in Amravati District 374 Shelake P.N., D.H. Ulemale ,Bochare K.V and Nagre K.G 374		
67	Temporal Changes in Input-Output Prices And Cost of Cultivation of Soybean in Vidarbha S. A. Borde, S. S. Thakare, P. G. Awagan and Ku. V. A. Deshmukh	380	



Taxonomic Study on Plants of Malvaceae

Reeta Satone, Pratibha Dhabarde,Swati kalode Department of Botany,Bajaj College of Science,Wardha

Abstract:

The present work deals with the identification of some weed plants belonging to family Malvaceae. In this study we select Dhaga forest area situated near Wardha city. The study is totally based on the identification of herb and shrub plants belonging to Malvaceae family. The study were carried on the month of September to January within 1 to 3 km area around the dhaga temple or forest. Near about 11 plants were identified on the basis of their morphological characterization. A total about 11 species belonging to 6genera of family Malvaceae.

Introduction :

The present study based on Angiospermic plants belonging to family Malvaceae also known as mallow family. The Malvaceae family comprises 88 genera and 2300species distributed across the world (Heywood 1978; Burkil 1997) also mentioned by Pradeepkumar 1996. The main characters of these family shows a large showy variously coloured flowers. One of the distinguishing character's include presence of various types of trichomes. In the Malvaceae family there are variations with herb, shrub and tree with mucilaginous sap in the all parts. Different taxonomist classified the family into tribe from time to time(bentham and Hooker in 1862). Plants of Malvaceae shows the diverse life forms. Habit and habitat of these malvaceae plants varies from plant to plant. It includes tufted or stellate hairs mucilage is found in root, stem , leaves and flowers of some species which gives it a slimy texture . The leaves are simple , alternate ,lobed free lateral stipule present in flowers, hermaphrodite, polypetalous, actinomorphic, Solitary inflorescence, sepal 5, basely connate calyx, stamens numerous monothecous , carpels as many as sepals long slender style, carpels5; axile placentation .

Study area:

Dhaga forest is situated near Wardha city. They are about 40 km away from Wardha. It is a very thick forest having the variations with Herb Shrub and Tree plant. We are selected about 1-3 km area around the temple of Shiva which are situated in Dhaga forest. It is much frequented tourist place in Wardha district .Best season to visit on September to January.

Climate:

Wardha city experienced humid and dry climate. Sometimes in the summer season the temperature rises above 46-48°C.and another time its stay average. Sufficient rain falls at rainy seasons from July to early October. In winter temperature falls up to 8-10 °C.

Soil is a fertile and vegetation of plants belonging to monocot and dicot plants including herb, shrub, tree and climbers.

Material and Methods:

The present study in based on the field of the area during the period of September 2018 to January 2019. Field visit and data collection were done once in every month. A total 11 species under 6 genera belonging to the family Malvaceae were collected and identified. The collected



Vidya Bharati Shaikshanik Mandal's

VIDYA BHARATI MAHAVIDYALAYA, AMRAVATI

Re-accredited with Grade 'A' by the NAAC (CGPA 3.26 - Second Cycle) College with Potential for Excellence

In Collaboration with

S.S.S.K.R. INNANI MAHAVIDYALAYA, KARANJA (LAD) DIST. WASHIM Re-accredited by NAAC with Grade 'A' (CGPA 3.24 - Second Cycle) College with Potential for Excellence

UGC Sponsored National Conference on

Emerging Trends in Science

1st & 2nd February, 2019

CERTIFICATE

This is to certify that Prof./Dr./Mr./Ms. P. V. Tekade

of J. B. Science College of Science, Wardha

has actively participated in the Conference held on 1" & 2" February, 2019 and has presented a research paper

oral / poster in the subject Chemistry

entitled In-vitro binding interaction of 2-(4-Hydrophenyl)-1H-benzimidazole withBSA: Equilibrium Dialysis

during technical session of the conference.

Dr. R. V. Joat Organizing Secretary

Dr. P. R. Rajput Principal & Co-Convenor S.S.S.K.R.I. Mv., Karanja Lad

Dr. F. C. Raghuwanshi Principal & Convenor VBMV, Amravati



Vidya Bharati Shaikshanik Mandal's

VIDYA BHARATI MAHAVIDYALAYA, AMRAVATI

Re-accredited with Grade 'A' by the NAAC (CGPA 3.26 - Second Cycle) College with Potential for Excellence

In Collaboration with

S.S.S.K.R. INNANI MAHAVIDYALAYA, KARANJA (LAD) DIST. WASHIM Re-accredited by NAAC with Grade 'A' (CGPA 3.24 - Second Cycle) College with Potential for Excellence

UGC Sponsored National Conference on

Emerging Trends in Science

1" & 2" February, 2019

CERTIFICATE

This is to certify that Prof./Dr./Mr./Ms. N. A. Barwat

J. B. Science College of Science, Wardha

has actively participated in the Conference held on 1" & 2" February, 2019 and has presented a research paper oral / poster in the subject Chemistry

entitled In-vitro binding interaction of 2-(4-Hydrophenyl)-1H-benzimidazole with BSA:

during technical session of the conference.

Dr. R. V. Joat Organizing

Secretary

Dr. P. R. Rajput Principal & Co-Convenor S.S.S.K.F.I. Mv. Karanja Leo

Dr. F. C. Raghuwanshi Principal & Convenor VBMV, Amravati

INTERNATIONAL RESEARCH FELLOWS ASSOCIATION'S

RESEARCH JOURNEY

International

E-Research

Journal

PEER REFREED & INDEXED JOURNAL February-2019 Special Issue – 110 (B)

CHEMISTRY

Guest Editor : Dr. F. C. Raghuwanshi Principal, Vidya Bharati Mahavidyalya, Amarawati

Executive Editors of the Issue :

Dr. P. R. Rajput, Principal, S. S. S. K. R. Innani Mahaviadyalaya, Karnja Lad

Dr. V. V. Parhate, HoD, Department of Chemistry,

Vidya Bhrati Mahavidyalaya, Amaravati.

Dr. L. K. Vays, HoD, Department of Cosmetics,

VidyaBhratiMahavidyalaya, Amaravati.

Dr. A. Y. Deshmukh, HoD, S. S. S. K. R. Innani Mahaviadyalaya, Karnja Lad.

- Dr. M. M. Rathore, Associate Professor, Department of Chemistry,
- Vidya Bhrati Mahavidyalaya, Amaravati.
- Dr. V. H. Masand, Assistant Professor, Department of Chemistry,
- Vidya Bhrati Mahavidyalaya, Amaravati.
- Dr. P. S. Bolkhe, Assistant Professor, Department of Chemistry,
- Vidya Bhrati Mahavidyalaya, Amaravati.

Mr. Harsh Agrawal, Assistant Professor, Department of Chemistry,

Vidya Bhrati Mahavidyalaya, Amaravati.

Chief Editor : Dr. Dhanraj Dhangar (Yeola)



This Journal is indexed in :

- University Grants Commission (UGC)
- Scientific Journal Impact Factor (SJIF)
- Cosmoc Impact Factor (CIF)
- Global Impact Factor (GIF)
- International Impact Factor Services (IIFS)



'RESEARCH JOURNEY' International E- Research Journal ISSN : Impact Factor - (SJIF) - <u>6.261</u>, (CIF) - <u>3.452(2015)</u>, (GIF) - <u>0.676</u> (2013) Experimental States of the second s

ISSN : 2348-7143 February -2019



- (SJIF) – <u>6.261</u>, (CIF) - <u>3.452(2015)</u>, (GIF)–<u>0.676</u> (2013) Special Issue 110 (B) : Chemistry UGC Approved Journal

	Synthesis and Antibacterial Studies of Some Novel Quinoxalines and			
42	Azirinoquinoxalines	222		
	Mohammad Idrees & Naqui Jahan Siddiqui & Sachin Bhongade			
43	Synthesis and Characterization of M-Nitroaniline and Their Complexes and Antimicrobial Activity Shaikh M.H			
	Synthesis, Physicochemical Characterisation, Solid State Conductance and Biological			
	Study of VO (IV), Moo2 (VI) and UO2 (VI) Coordination Compounds With 2, 4-			
44	Dihydoxy-5-Acetyl Acetophenone And 1, 4-Diaminobutane Schiff Base Tetra	233		
	Dentate Ligand.			
	Muralidhar K. Rahangdale & Vandana B. Badwaik & Anand S. Aswar			
45	Synthesis, Characterisation and Antimicrobial Activity of Thiazolidinone	238		
	R.P. Pawar & Nadeem A. Sheikh			
16	Utility of Ninhydrin For Spectrophotometric Determination of Kanamycin in Pure	2.40		
46	Form and in Pharmaceutical Formulation	248		
	Naqui Jahan Siddiqui & Mohammad Idrees & Parinita Adkine			
47	Pharmacophore Modeling for Anti-Parasitic Activity of 2-Nitroimidazopyrazin- one/- Es Nilesh S. Kadu	252		
	Removal of Methyl Red Dye from Aqueous Solution By Employing Nife2o4 Nanoparticle			
48	Mr. Santosh M. Chavan & J. S. Jadhaoa & C. D. Ghugareb & P. J. Sakharea,	257		
40	&N.V. Rathodc & M. S. Morec, & R. R. Wankhaded, & A. B.Patila & S. M.	231		
	Chavana*			
10	Microwave Assisted Synthesis and Characterization of Carbazole Derivative and	• < 1		
49	Study of Their Antibacterial Activity Pravin A.Sonune & Pankaj S Chaudhari	261		
50	Synthesis, Characterization and Antibacterial Study of Chlorosubstituted Thiazoles	265		
50	Kolhe P.A. & Rajput P. R.	265		
51	Microbial Evaluation of Nanoparticles of Some New Lactosyl 5-Aryl 2,4	269		
51	Dithiobiurets Poonam T. Agrawal	268		
52	Microbial Evaluation of Ginger oil Against Dandruff	271		
34	Poonam Agrawal & Apurva Shah	2/1		
53	Non-Aqueous Potentiometric Analysis of Drug Furosemide in Bulk and Single	275		
	Component Pharmaceutical Tablets Pradip P. Deohate			
	In-Vitro Binding Interaction of 2-(4-Hydrophenyl)-1H-Benzimidazole With BSA:			
54	Equilibrium Dialysis and FT-IR Spectroscopic Study	283		
54	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez	283		
54	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar	283		
54 55	 Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In 	283 289		
55	Equilibrium Dialysis and FT-IR Spectroscopic StudyAjay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & ParvezSaudagarMicrowave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino RatsP. S. Nandurkar & R. Rajput & M. M. Rathore	289		
	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6 □ Hydroxypyridazinones as Σ1			
55 56	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi	289 292		
55	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6 – Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines	289		
55 56 57	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines P. S. Pande	289 292 295		
55 56	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6 – Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines	289 292		
55 56 57 58	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines P. S. Pande Synthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii)	289 292 295 302		
55 56 57	Equilibrium Dialysis and FT-IR Spectroscopic StudyAjay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & ParvezSaudagarMicrowave Synthesis and Wound Healing Effect of $\Delta 2$ – Pyrazoles ointment InAlbino RatsP. S. Nandurkar & R. Rajput & M. M. RathoreDevelopment of Consensus Pharmacophore For 6 \Box Hydroxypyridazinones as $\Sigma 1$ Receptor LigandsSampatrao B. SuryawanshiSynthesis and Study of Antioxidant Activity of N-Thiocarboxamido PyrazolinesP. S. PandeSynthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii)Complexes of Onno Donar Schiff BaseA. B. Sahare & R. B. MohodSynthesis, Charecteriation and Antibacterial Activity of Some ThiocarbamidesComplexesRahul B. Mohod	289 292 295		
55 56 57 58 59	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines P. S. Pande Synthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii) Complexes of Onno Donar Schiff Base A. B. Sahare & R. B. Mohod Synthesis, Charecteriation and Antibacterial Activity of Some Thiocarbamides Complexes Rahul B. Mohod Evaluation of water Quality by Physicochemical Parameter from Umarkhed Tehesil	289 292 295 302 306		
55 56 57 58	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines P. S. Pande Synthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii) Complexes of Onno Donar Schiff Base A. B. Sahare & R. B. Mohod Synthesis, Charecteriation and Antibacterial Activity of Some Thiocarbamides Complexes Rahul B. Mohod Evaluation of water Quality by Physicochemical Parameter from Umarkhed Tehesil District Yavatmal (M.S.) Ingole R.N. & Waghmare S.B. & Kanade D.B	289 292 295 302		
55 56 57 58 59 60	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines P. S. Pande Synthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii) Complexes of Onno Donar Schiff Base A. B. Sahare & R. B. Mohod Synthesis, Charecteriation and Antibacterial Activity of Some Thiocarbamides Complexes Rahul B. Mohod Evaluation of water Quality by Physicochemical Parameter from Umarkhed Tehesil District Yavatmal (M.S.) Ingole R.N. & Waghmare S.B. & Kanade D.B Predicted Biological Activities of Compounds Planned to be Synthesized, by the	289 292 295 302 306 310		
55 56 57 58 59	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines P. S. Pande Synthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii) Complexes of Onno Donar Schiff Base A. B. Sahare & R. B. Mohod Synthesis, Charecteriation and Antibacterial Activity of Some Thiocarbamides Complexes Rahul B. Mohod Evaluation of water Quality by Physicochemical Parameter from Umarkhed Tehesil District Yavatmal (M.S.) Ingole R.N. & Waghmare S.B. & Kanade D.B Predicted Biological Activities of Compounds Planned to be Synthesized, by the Application of PASS and Their Toxicity Risk Assessment By The Use of osiris	289 292 295 302 306		
55 56 57 58 59 60	Equilibrium Dialysis and FT-IR Spectroscopic StudyAjay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & ParvezSaudagarMicrowave Synthesis and Wound Healing Effect of $\Delta 2$ – Pyrazoles ointment InAlbino RatsP. S. Nandurkar & R. Rajput & M. M. RathoreDevelopment of Consensus Pharmacophore For 6 – Hydroxypyridazinones as $\Sigma 1$ Receptor LigandsSampatrao B. SuryawanshiSynthesis and Study of Antioxidant Activity of N-Thiocarboxamido PyrazolinesP. S. PandeSynthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii)Complexes of Onno Donar Schiff BaseA. B. Sahare & R. B. MohodSynthesis, Charecteriation and Antibacterial Activity of Some ThiocarbamidesComplexesRahul B. MohodEvaluation of water Quality by Physicochemical Parameter from Umarkhed TehesilDistrict Yavatmal (M.S.)Ingole R.N. & Waghmare S.B. & Kanade D.BPredicted Biological Activities of Compounds Planned to be Synthesized, by theApplication of PASS and Their Toxicity Risk Assessment By The Use of osirisProperty Explorer.Santosh G. Badnea and Gajanan W. Belsareb	289 292 295 302 306 310		
55 56 57 58 59 60 61	Equilibrium Dialysis and FT-IR Spectroscopic Study Ajay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & Parvez Saudagar Microwave Synthesis and Wound Healing Effect of Δ2 – Pyrazoles ointment In Albino Rats P. S. Nandurkar & R. Rajput & M. M. Rathore Development of Consensus Pharmacophore For 6□ Hydroxypyridazinones as Σ1 Receptor Ligands Sampatrao B. Suryawanshi Synthesis and Study of Antioxidant Activity of N-Thiocarboxamido Pyrazolines P. S. Pande Synthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii) Complexes of Onno Donar Schiff Base A. B. Sahare & R. B. Mohod Synthesis, Charecteriation and Antibacterial Activity of Some Thiocarbamides Complexes Rahul B. Mohod Evaluation of water Quality by Physicochemical Parameter from Umarkhed Tehesil District Yavatmal (M.S.) Ingole R.N. & Waghmare S.B. & Kanade D.B Predicted Biological Activities of Compounds Planned to be Synthesized, by the Application of PASS and Their Toxicity Risk Assessment By The Use of osiris Property Explorer. Santosh G. Badnea and Gajanan W. Belsareb Synthesis and Characterization of Cobalt Sulfide Nanoparticals by	289 292 295 302 306 310 314		
55 56 57 58 59 60	Equilibrium Dialysis and FT-IR Spectroscopic StudyAjay Pisudde & Pradip Tekade & Shrikant Thakare & Nakul Barwat & ParvezSaudagarMicrowave Synthesis and Wound Healing Effect of $\Delta 2$ – Pyrazoles ointment InAlbino RatsP. S. Nandurkar & R. Rajput & M. M. RathoreDevelopment of Consensus Pharmacophore For 6 – Hydroxypyridazinones as $\Sigma 1$ Receptor LigandsSampatrao B. SuryawanshiSynthesis and Study of Antioxidant Activity of N-Thiocarboxamido PyrazolinesP. S. PandeSynthetic, Spectral and Thermal Investigation of Mn(Ii), Co(Ii), Ni(Ii) and Cu(Ii)Complexes of Onno Donar Schiff BaseA. B. Sahare & R. B. MohodSynthesis, Charecteriation and Antibacterial Activity of Some ThiocarbamidesComplexesRahul B. MohodEvaluation of water Quality by Physicochemical Parameter from Umarkhed TehesilDistrict Yavatmal (M.S.)Ingole R.N. & Waghmare S.B. & Kanade D.BPredicted Biological Activities of Compounds Planned to be Synthesized, by theApplication of PASS and Their Toxicity Risk Assessment By The Use of osirisProperty Explorer.Santosh G. Badnea and Gajanan W. Belsareb	289 292 295 302 306 310		

In-Vitro Binding Interaction of 2-(4-Hydrophenyl)-1H-Benzimidazole With BSA: Equilibrium Dialysis and FT-IR Spectroscopic Study

Ajay Pisudde, Pradip Tekade*, Shrikant Thakare, Nakul Barwat, Parvez Saudagar Department of Chemistry, Jankidevi Bajaj College of Science, Jamnalal Bajaj marg, Civil lines, Wardha (India). Email: pisuddeajay@gmail.com *Corresponding author@: pradiptekade@gmail.com

Abstract:

This paper presented binding interaction of 2-(4-hydrophenyl)-1H-benzimidazole (4HPHBI) with bovine serum albumin (BSA) equilibrium dialysis and FT-IR study at physiological pH in various solvents. Findings were interpreted by scatchard plot in terms of association constants. This showed the increased in association constants with increase in concentrations of the 4HPHBI. It is seen that, the binding interaction supposed to be more in 1, 4-dioxane than DMSO and DMF. FT-IR study explained the binding interaction through shift in peak positions of amide I and II. FT-IR study concluded the changes in secondary structure of BSA on binding with 4HPHBI. In the present study, we have also reported the effect of foreign particles viz. arsenic and mercury on the binding interaction of 4HPHBI with BSA. Reactivity and association of 4HPHBI with BSA in presence of foreign moiety was determined and compared.

Keywords: BSA, Equilibrium dialysis, FT-IR study, Scatchard plot, association constant, foreign particles.

Introduction:

N-heterocyclic compounds are important class of compounds which are widely used in materials science, agro-chemistry, biomedical research and medicinal chemistry. Amongst the N-heterocyclic compounds, benzimidazole has a prominent place in organocatalysis, organometallic and material chemistry. 2-(4-Hydrophenyl)-1H-benzimidazole (4HPHBI) shows various biological properties especially, antimicrobial, antiviral, anticancer and antitumor¹⁻⁶. Serum albumins are the most abundant proteins in the circulatory system of wide variety of organisms. The structure of HSA explains numerous physiological phenomena and provides further insight in pharmacokinetics. Variation in the temperature is found to be a key factor in binding affinities of HSA⁷⁻⁸ as evident from various drugs viz. Ligustrazine, Ciprofloxacin⁹, methotrexate¹⁰ and cisplatin¹¹. It is difficult to obtain HSA for experimental purposes. HSA and BSA exhibit similar chemical properties due to high percentage of sequence identities. BSA in lieu of HSA is used in this study because of low cost and easy availability. Various techniques are available to monitor the binding interactions of ligands to protein like NMR¹², isothermal titration calorimetry¹³, U.V. visible absorbance¹⁴, fluorescence¹⁵, equilibrium & FT-IR¹⁶, fluorescence and CD spectroscopy¹⁷. Equilibrium dialysis and FT-IR study also focused on the effect of foreign particles such as arsenic on protein-drug binding¹⁸⁻¹⁹.

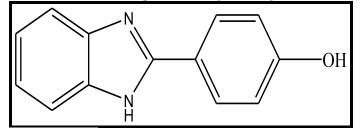


Figure 1: Structure of 2-(4-hydrophenyl)-1H-benzimidazole (4HPHBI)



This is to certify that, has participated / Presented research paper in oral / poster session / delivered invited talk / chaired session / worked as committee member in the National Seminar on Diversity of Environmental Allergens and Its Threat to Human Health.

Organizing Secretary

On

9th Feb. 2019

Certificate

Dr. Mrs. Sharayou Taywade

Dr. Ravindra Baheka

Dr. Chandrakant Bhaskar







Shri Sachhidanand Shikshan Sanstha's Taywade College, Mahadula-Koradi MAC Accredited with B Grade One Day National Seminar On DIVERSITY OF ENVIRONMENTAL ALLERGENS AND ITS THREAT TO HUMAN HEALTH Organized by - Department of Botany, Microbiology, Zoology & Chemistry

9th Feb. 2019

-:- Editorial Board for Abstract Proceeding -:-Dr. (Mrs.) Suvarna P. Patil, Dr. Chandrakant S. Bhaskar, Dr. Vijay N. Charde Dr. Ravindra S. Bahekar, Dr. Yogesh W. There

INDEX- Abstract

Sr. No.	Author	Title	Page No.
01	Parikshit T. Choudhary, Neha V. Nerkar, Subhash B. Kondawar	Removal of Synthetic Dyes from Industrial Waste Water using Nanocomposites	03
02	Shrirame, A. M. and Hiwale, S. R.	Survey on diversity of wild annual plant species at K.Z.S. Sci. College campus locality, Bramhani, Kalmeshwar	04
03	Ajay. K. Potbhare, RinaBagade, S. Zahra, S. Mondal, R. G. Chaudhary	Green Fabrication of TiO ₂ -Graphene Oxide Nanocomposites for the Photocatalytic applications	05
04	Deshmukh R.N., Mahakhode R.H. &Puranik S.D	Studies on Impact of ongoing Metro Railway Project on 'Green Campus'of Shivaji Science College, Nagpur.	06
05	Mohture V.M. and Kalkar S.A	Study of fungal diversity in the atmosphere of Nagpur (M.S.), India	07
06	Pranita A. Gulhane	Bioaerosols in ENT Hospitals: A Threat for Nosocomial Infection	08
07	S.S.Shende& P.S.Jogi	Qualitative Analysis of Kalinagar Village TalukaMulchera (MS)	09
08	MenghareVaibhav and Bhajbhuje M N	Aeromycoflora of Satpuda Botanical Garden, Nagpur	10
09	ChetnaUgaleand Jaykiran A. Tidke	Pollen physiological dynamics of <i>C. roseus</i> after exposure to heavy metals	11
10	S. C. Masram,K. T. Waghmare, D. B. Meshram and N. D. Gorghate	Truss Network Systemand DNA barcoding: Promising tools in identification conservation and management in fisheries	12
11	Anita D. Kushwaha , P.D. Dhethe	Structural and Thermal Decomposition Study of Resin-II Derived from p-Nitrophenol, Resorcinol and Formaldehyde	13
12	Shalini R.Singh and Sumer D. Thakur	Antimicrobial activity of heterocyclic compound containing nitrogen and sulphur	14
13	B.S.Tapase	Identification and Enumeration of Bacteria Present in Air Around the College Area, Nagpur.	15
14	S. S. Deshmukh	Individual Identification and Distribution of leeches in Melghat Region	16
15	Dilip M. Chafle	Impact of Fly Ash of Thermal Power Plant on Human Health	17
16	Chahande .P.P &Duragkar A. M.	Detection of Adulterants in Some Common Food Stuff	18
17	NusratBabar Shaikh	Discovery of fern Fossil in Sironcha Dist. Gadchiroli Maharashtra	19
18	K. P. Ghoshal and S. S. Chourasia	Aeroallergens and its Impact on Human Health	20
19	Gandhewar, S.S. and Zade, S.B.	Histopathological changes in the gill and liver of <i>Clariasbatrachus</i> (Linn.) exposed to copper	21
20	Girhe H.K., Meshram S. M. and Mohture V. M.	Ethnoveterinary Medicinal plants of Shirdon, DistRaigad (M.S.) India	22

Sr. No.	Author	Title	Page No.
21	Meshram, S.M.	Aeropalynological Studies in the Atmosphere of Khaperkheda, Dist. Nagpur (M. S.)	23
22	A. M. Duragkar and P. P. Chahande	Determination of Adulteration of Starch, Cane Sugar, Urea and Detergent in milk from Local Vendors	24
23	A.P. Sakharkar, S. S. Khandare, M. G. Ingale	Production, Extraction, Quantification of Biopigment "Prodigiosin" from Serratia Marcescensand Its Antibacterial activity.	25
24	Narkhedkar, V.R., Tidke, J.A.and Chikhale, N.J.	Variable Responses During Androgenesis in <i>Catharanthusroseus</i> (L.) G. Don. Under Different Nutrient Media	26
25	PunitaTiwari	Eco-Utilization of Waste Produced from Various Religious Shrines	27
26	Rahate US and Masram SC	Study of Preliminary Phytochemical Activity of Cassia Sophera (LINN) Seed Eextract	28
27	Shende B.G. and Masram, S.C.	Embrynonic Development of Green Marsh Hawk, Orthetrum Sabina Sabina (DRURY,1770)	29
28	S.V.Ghonmode	Application of the Silk SericinCoating on PolysterFabric from the Silkgland of Silkworm in Wound Healing	30
29	<u>S. Mondal</u> , R. G. Chaudhary, A. K. Potbhare, S. Zahra	Synthesis& Characterization of Magnetic Iron Oxide Loaded Activated Carbon (AC-Fe ₃ O ₄) for the Removal of Methylene BlueE	31
30	PravinMeshra, VaishaliMeshram, SaviShende	Phyco-remediation an Effective Technology for TDS Removal from Water & Wastewater	32
31	Sandip H. Shende	Phytochemical Screening of TagetesErectal L. Leaves	33
32	S. B. Page	Isolation and Characterization of probiotic Lactic Acid Bacteria from Intestine of fresh water fish Rohu (<i>Labeorohita</i>) From Chandrapur District	34
33	Meshram S. M., Mohture V.M., Purkam.V.P.	The Angiospermic Fossil Fruit from a New Locality Ghatparasia, M.P. India	35
34	Miss. Shweta S. Ingle	Diversity of Environmental Allergens and its Threat to Human Heath	36
35	Ayesha Hussain Khan	Studies on Aero-Spora of Dr. C. V. Raman Science College, SironchaPrimises Area. Tha. Sironcha, Dist. Gadchiroli, Maharashtra	37
36	JayshreeThaware	Monitoring Environmental Biopolution in Kamptee and its Relevance in Public Health	38
37	Ulka A.Malode and DivyaTanwar	Antimicrobial Activity of Chemically Synthesized Copper Oxide Nanoparticle	39
38	Junaid Sheikh and HarshaSonaye	A promise to scale up in pharamaceuticals : Pilot plant scale up technique.	40
39	K. S. Khandare, P.K.Kadu, J. M. Barabde	Synthesis and characterization of antibacterial Isoniazid Derivatives	41
40	MamtaMarar	Anacardiumoccidentale L Phytochemicals: A Great Remedy Against Skin Infection Causing Bacterial Pathogens	42





Production, Extraction, Quantification of Biopigment "Prodigiosin" from Serratia Marcescens and Its Antibacterial activity. A.P. Sakharkar¹, S. S. Khandare²*, M. G. Ingale³

¹ P.G. Student, Post graduate Dept. of Microbiology, J. B. College of Science, Wardha,
 ² Asso. Professor and HOD, Dept. of Microbiology, J. B. College of science, Wardha,
 ³ Asstt. Professor, Post graduate Dept. of Microbiology, J. B. College of Science, Wardha,

Prodigiosin is a biopigment and secondary metabolite produced by many strains of Serratia marcescens and other gram negative bacteria. Prodigiosin is a valuable molecule due to its reported antifungal, immunosuppressive and anti-proliferative activities. In the present study, total 17 natural samples ie. 10 soil samples, 3 water samples and 4 sewage effluent samples were collected from wardha region and screened for prodigiosin production, Total five isolate were found to produce the red pigment viz: AK1, AK2, AK3, AK4, AK5. Isolate AK5 was found to be most efficient prodigiosin producer and was selected for further study. AK5 was identified on the basis of cultural, morphological and biochemical characterization as Serratia of pigment carried *marcescens*.Production was out on nutrient broth. Pigmentextraction was carried out by using methanol and petroleum ether as solvent and subjected to spectrum scanning in range between 300-700 nm followed by presumptive test to confirm pigment as prodigiosin. Qualitative analysis of pigment was also carried out bypaper chromatographyand Rf value of 0.42 was obtained. Further, pigment was also quantified by using standard formula and prodigiosin was estimated to be 4020 unit/cell by methanol extraction method and 1580unit/cell by petroleum ether extraction method. Antibacterial activity of prodigiosin was also tested against E. coli, P.aeruginosa, S.aureus and S. typhi and zone of inhibitions were found to be 12 mm, 11 mm, 13 mm & 12 mm respectively. Antibiotic susceptibility pattern of isolate was also studied against 7 different antibiotics which are routinely used clinically. .

Key Words: Progidiosin, Serratiamarcescens, Antibacterial

37

PRODUCTION, EXTRACTION AND QUANTIFICATION OF PRODIGIOSIN, A BIOPIGMENT PRODUCED FROM SERRATIA MARCESCENS AND ITS ANTIBACTERIAL ACTIVITY AGAINST PATHOGENIC ORGANISMS

A.P. Sakharkar¹, S. S. Khandare²*, M. G. Ingale³

Post graduate Dept. of Microbiology, J. B. College of Science, Wardha. *Corresponding author e-mail: ksuhas21@gmail.com

Abstract

Prodigiosin is a biopigment and secondary metabolite produced by many strains of Serratia marcescens. Prodigiosin is a valuable molecule due to its immunosuppressive, antiproliferative and antifungal activities. In the present study, total 17 natural samples ie.10 soil samples, 3 water samples and 4 sewage samples were collected from wardha region and screened for prodigiosin production, Total five isolate were found to produce the red coloured pigment viz: AK1, AK2, AK3, AK4, AK5. Isolate AK5 was found to be most efficient prodigiosin producer and was selected for further study. AK5 was identified on the basis of cultural, morphology and biochemical characteristics and comparing with Bergeys Manual of Determinative Bacteriology (9th edition) as Serratia marcescens. Production of pigment was carried out on nutrient broth. Pigment extraction was done by using petroleum ether and methanol as solvent and subjected to spectrum scanning in range between 400-700 nm followed by presumptive test to confirm pigment as prodigiosin. Qualitative analysis of pigment was also carried out by paper chromatography and Rf value of 0.42 was obtained. Further, pigment was also quantified by using standard formula and prodigiosin was estimated to be 4020 unit/cell by methanol extraction method and 1580 unit/cell by petroleum ether extraction method. Prodigiosin was evaluated for its antibacterial activity against E. coli, P. aeruginosa, S. aureus, S. typhi and zone of inhibitions of 12 mm, 12 mm, 13 mm and 11 mm respectively were recorded. The isolate was found to be completely susceptible to Chloramphenicol, Ciprofloxacin, Cloxacillin, Erythromycin, Gentamicin and resistant towards Ampicillin and Streptomycin. Key Words: Prodigiosin, Serratia marcescens, Antibacterial activity

Introduction

The red pigment prodigiosin is having unique tripyrrole structure and belongs to the alkaloid class of secondary metabolites. This pigment can be obtained from a few members of *Pseudomonas, Streptomyces* and *Serratia* species [1]. It is a light sensitive pigment, insoluble in water and moderately soluble in ether and alcohol, and soluble in methanol, chloroform, acetonitrile and DMSO [2]. There are several organisms which can produce pigments and two major sources are plants (3) and microorganisms (4). There are various sources of pigments and plants occupies the main place among them. But problem with plant source is the attack of many pathogens on plants leads to the loss of the plants. [5]. There are variety of natural pigments



This is to certify that Prof./Dr/Mr./Mrs./Ms. **Dr. Pradip V. Tekade** has participated and delivered keynote address / invited lecture / oral presentation / poster presentation / chaired a technical session in the international conference on energy and environmental Challenges during 18-19th January 2019 in Nagpur, India

Prof. P.M. Padole Director

Prof. S.S.Umare Head, Dept. of Chemistry

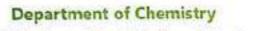


Dr. A.V. Wankhade Chair-CE.C-2019

International Conference on Energy and Environmental Challenges

1 1

18-19" January, 2019



Organized by

Visvesvaraya National Institute of Technology, Nagpur

In collaboration with

Tata Institute of Fundamental Research, Mumbai





Scanned by CamScanner

	Rahim. S. Sheikh, FarhanA. Khan	Synthesis And Characterization Oxoazetidin-N-Substituted
P-27		Applications of Carbon Based Nano-Materials
P-28	Satish Joshi, Manoj Pande,	and the billing of sumthaning days to start
	Gajanan Kotharu S.S. Deo, D. P. Gulwade, F.Inam,	Thermal stability of synthesized a-benzalidine - γ -phenyl-8,
P-29	H. P. Dahikar	p, journer i i i i i i i i i i i i i i i i i i i
	1 1 2 2 2 1 2 2 2 2 2 2 2 2 2 2 2 2 2 2	investigation
	Saurabh S Suranshe, Avnikumar P. Patil	Application of Nano-technology in Lithium Ion Battery
P-30 P-31	LI P Manwatkar, C.S. Bhassai,	Application of reality technology in Elanum ion Battery Amino acid and thiol modified graphene oxide: synthesis and characterization
P-31	M G. Dhonde, S. R. Inakare	Graphene: - A versatile Nano-material in Energy and
P-32	IIImani C Pandhurnekar,	Environmental Applications
	Dissel W Dechminkh, Kiran Kaul	Design and Analysis of On-grid Rooftop Solar Photovoltaic System
P-33	Jaya Goyal, S. S. Mantha, V. M. Phalle	Using MATLAB
	Jyoti N. Thakre, Sanjay R. Thakare ,	Removal of 2-Chlorophenol from Aqueous Solution By Zero-
P-34	P. T. Kosankar	
0.25	KasalaPrabhakar Reddy,	Mapping Valence Band and Interface Electronic Structure
P-35	Nitin B Mhamane, Manoj K. Ghosalya,	Changes during Oxidation of Mo to MoO ₃ via MoO ₂ and
	Chinnakonda S Gopinath	MoO ₃ Reduction to MoO ₂ : A NAPPES Study
P-36	Kiran Raut, H.C. Pandhurnekar	Different Solar Cell Technology and its Application
P-37	Komal Mahindrakar, Virendra Rathod	Utilization of Banana Peels for Removal of Strontium (II)
		from Water
P-38	Lakshman Gadekar, Pandit Khakre, Shama Lomate	Synthesis of azlactone by Cu-nanoparticle- stilbite catalyst
P-39	M. B. Kumbhare, V. S. Sapkal,	Preparation and Characterization of PVAc-MgO
1	A. K. Gade, S. S. Sen	Nanocomposite membrane
P-40	Mahendra Chaudhari, Deepak Kayande,	A highly efficient and sustainable synthesis of
	Murlidhar Shingare	dihydropyrano[2,3-c]pyrazoles using polystyrenesupported
		p-toluenesulfonic acid as reusable catalyst
P-41	ShaliniSahani, Yogesh Chandra Sharma	Biodiesel production using a novel heterogeneous basic catalyst: kinetic and thermodynamic investigations
P-42	Manoj Shanti, Kaveri Shanti,	E-liminating E-Waste: A Chemical Approach
	Manisha Raut	
P-43	Mr. Laxmikant N. Vairagade,	A Concept of Green Building
P-44	Neelam Kadam, Surekha Kalkar	Plants as Potential Renewable Energy Source for Biofuels
P-45	Nitesh D. Punyapreddiwar,	Saccharomyces cerevisiae catalyzed cyclocondensation
B 46	Umesh R. Pratap	reaction: synthesis of pyrazoline from chalcone
P-46	Nikhat Sheikh, Vijay Tangde,	Effect of dextrose and urea on the solution thermodynamics
	Niraj Khaty	of ornithine monohydrochloride in aqueous solution at T=
P-47	NEG. I	(288.15, 298.15 and 308.15) K
1-4/	Nitin Lavande, Rahul More,	Mg modified MnO ₄ -CeO _{2.8} catalyst for low temperature
	Pavan More	complete oxidation of simulated diesel engine exhaust and
P-48	P. P. Kalbende	VOCs
P-49		Synthesis of silver nanoparticles using plant latex extract
STERNING C	P. U. Meshram, Vaishali Meshram, Savi Shende	A Study of Occupational Health Effects on Flour Mill
P-50	P.R. Chaudhari,S.S.Darunkar,	Workers in Fastern Region of Nagnur City
	S. A. Acharya	Nanocrystalline NiFe2O4 thick film sensor for LPG sensing
P-51	Deepa Panhekar, Pooja Bure,	
19979792	Ashok Kalambe, Pravin Deharkar	Adsorption of Cu (II) ions from aqueous solution by
P-52	PradipTekade, Kailash Nemade,	A.ficoidea
	Gauri Bhoyar	Effect of Ultrasound on Particle Size and Gas Sensing
P-53	Pushpak Daryapurkar, Ganesh Wahile	Properties of Graphene/TiO ₂ Composite
P-54	RajdipUtane, Sujata Deo	Conversion of solid waste into electricity
	July chance, Sujata Deo	Low cost and Time Efficient Recycling and Separation
		Techniques of Plastic (HDPE and LDPE) by 12
	10	New Green Approach

preventional Conference on Energy and Environmental Challenges CE2C 2019

Effect of Ultrasound on Particle Size and Gas Sensing Properties of Graphene/TiO2Composite

Pradip Tekade⁴, Kailash Nemude⁴, Gauri Bhoyar⁴ "Department of Chemistry, Jankidevi Bajig College of Science, Wardba 442 001, India. "Department of Physics, India Mahovidyalaya, Kalamb 445 401, India. *E-mail: prostiptekade@gmoil.com*

Increased consciousness about the environmental issues, especially regarding health and safety, has focused on the need of air-quality monitoring.¹ Gas sensors have been playing crucial role in environmental monitoring, industrial safety, diagnosis of diseases, and traffic safety.²⁴This paper reported the effect of particle size on gas sensing properties. Graphene/TiO₂ composite system were used to analyze the effect of particle size. The particle size of TiO₂ nanoparticles reduced using probe sonication technique. Four different time dose of probe sonication given to TiO₂, in order to reduce the particle size. The particle size of TiO₂ nanoparticles after probe sonication soccessfully estimated using X-Ray Diffraction analysis. As-obtained TiO₂ nunoparticles of reduced particle size used for the preparation of graphene/TiO₂ composites. Ultraviolet-visible spectroscopy also shows that particle size can reduced by probe sonication and it is also useful for band gap engineering. The doctor blade technique was employed to fabrication of sensors. As-fabricated sensors shows good dependence on the particle size. The sensors were tested towards CO₂ and LPG. In both cases, sensing response increases with reduction in particle size.

References

- I. K. Wetchakun, T. Samerjai, N. Tamackong, C. Liewhitan, C. Siriwong, V. Kruefu, A. Wisitsonaat, A. Turntranom, S. Phanichphani, Sentors and Actuators B2011 [60, 589–59].
- Sintra, P.K. Guha, S.Z. Ali, P. Himlal, H.E. Unalan, J.A. Covington, G.A.J. Amaratunga, W.I. Milne, J.W. Garcher, F. Udrea, Sontory and Actuators 32910 146, 146559–565.
- 1. N. Sothada, G. Bronstrup, K. Funka, S. Christiansen, M. Leja, H. Haick, Nuso Letters 2014, 15,1288-1295.
- M. Hakim, Y.Y. Broza, O. Barash, N. Pelied, M. Phillips, A. Amann, H. Huick, Chemical Reviews 2012, 112, 5949–5956.

As per the Syllabus of Jankidevi Bajaj College of Science, Wardha (An Autonomous College)

A Text Book Concepts of Chemistry

B. Sc. - I Sem - II



Dr. P. V. Tekade Mr. M. D. Bansinge

Dr. Mrs. N. P. Mohabansi Mr. N.A. Barwat

Dr. P. G. Borkar





₹ 225/-

Meherbaba Publishers

Near Hasanbag Police Chowki, Hasanbag Road, New Nandanvan, Nagpur M.: 9923630315, 8087460470 E-mail : meherbabapublishers0616@gmail.com

Published by

MEHERBABA PUBLISHERS

Happy Home Boys Hostel, Near Hasanbag Police Chowki, Hasanbag Road, New Nandwan, NAGPUR Mob. : 9923630315, 8087460470 E-Mail : meherbabapublishers0616@gmail.com

B.Sc. - I (Semester - II)

Concepts of Chemistry

© PUBLISHERS-

The copyright of subject matter, illustrations, style and expression of all the editions reprints is strictly reserved. This book or any part thereof must not be reproduced or reprinted in any form what so ever, without the written permission of the publishers, expect for the purpose of references and review. Infringement of copyright is a criminal offence.

1" Edition - 2019

ISBN: 978-93-87558-30-8

Mr. M. D. Hansings

Dr. P. G. Borkar

₹ -225 /-

Dr. P. G. Borkar

As per the Syllabus of Jankidavi Bajaj College of Science, Wardha (An Autonomous College)

0

Concepts of Chemistry B.Sc. I - Sem - II

Dr. P. V. Tekade M. Sc. B. Ed. SET, NET, GATE Ph. D. Dr. Mrs. N. P. Mohabansi M. Sc. B. Ed. SET Ph. D.

Mr. M. D. Bansinge M. Sc. NET Mr. N. A. Barwat

Dr. P. G. Borkar M. Sc. SET, NET, GATE Ph. D. Post Doc.

CONTENTS

THEORY COURSE

UNIT I

s-block elements & Noble Gases

A. s-block elements	[-9
B. Chemistry of Noble Gases	18

UNIT- II

p-block elements

A. Comparative study of groups 13 to 1719-25	
8. Hydrides	ĺ

UNIT III

Aliphatic Hydrocarbons (I)

A. Alkanes	
B. Alkenes	
C. Alkynes	

UNIT IV

Aliphatic Hydrocarbons (II) and Aromatic hydrocarbons

A. Dienes	
B. Aromatic Compounds and Aromaticity	

UNIT V

Thermodynamics II

Second law of thermodynamics
Free energy functions

UNIT VI

Solid State and Phase Equilibra

A. Solid State	139-162
B. Phase Equilibrium	163-196

Practical Course	31
Inorganic Chemistry, Organic Chemistry, Physical Choemistry	
Appendix	37

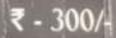
As per the Syllabus of Jankidevi Bajaj College of Science, Wardha (An Autonomous College)

A Text book of Fundamentals of Chemistry

B. Sc. - I Sem - I

Dr. P. V. Tekade Mr. M. D. Bansinge Dr. Mrs. N. P. Mohabansi Mr. N.A. Barwat

Dr. P. G. Borkar



Meherbaba Publishers

E

Near Hasanbag Police Chowki, Hasanbag Road, New Nandanvan, Nagpur M.: 9923630315, 8087460470 E-mail : meherbabapublishers0616@gmail.com

Published by MEHERBABA PUBLISHERS

Happy Home Boys Hostel, Near Hasanbag Police Chowki, Hasanbag Road, New Nandwan, NAGPUR Mob. : 9923630315, 8087460470 E-Mail : meherbabapublishers0616@gmail.com

B.Sc. - I (Semester - I)

Fundamentals of Chemistry

C PUBLISHERS-

The copyright of subject matter, illustrations, style and expression of all the editions reprints is strictly reserved. This book or any part thereof must not be reproduced or reprinted in any form what so ever, without the written permission of the publishers, expect for the purpose of references and review. Infringement of copyright is a criminal offence.

Or. P. G. Borkar

1" Edition - 2018

ISBN: 978-93-87558-20-5

Mr. R. A. Marryan

₹ - 300/-

Dr. Mahejabeen Hague

As per the Syllabus of Jankidavi Bajaj College of Science, Wardha (An Autonomous College)

A Text book of

Fundamentals of Chemistry B.Sc. I - Sem - I

Dr. P. V. Tekade M. Sc. B. Ed. SET, NET, GATE Ph. D. Dr. Mrs. N. P. Mohabansi

M. Sc. B. Ed. SET Ph. D.

Mr. M. D. Bansinge M. Sc. NET Mr. N. A. Barwat M. Sc. SET, NET

Dr. P. G. Borkar M. Sc. SET, NET, GATE Ph. D. Post Doc.

CONTENTS

Unit - I	
1. A. Atomic Structure	1-32
1. B. Periodic Properties	
Unit - II	
2. A. Chemical Bonding	
2. B. Ionic Bond	
Unit- III	
3. Basics of Organic Chemistry	
4. Stereochemistry	
5. A. Gaseous State	
5. B. Liquid State	
Unit - VI	
6. Thermodynamics	
7. Basics of Practical Chemistry	
8. Chemistry Lab: Practical Course	
A. Inorganic Chemistry	
B. Organic Chemistry	
C. Physical Chemistry	

Organic Chemistry Laboratory Course Book

(Preparations, Quantitative Analysis and Isolation of Organic Compounds from Natural Sources)

Dr. Pradip V. Tekade



OUR PUBLICATIONS



r



Former Science In Respond Journ. Technique







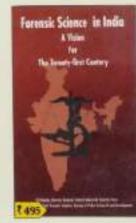












FORENSIC SEROLOGY R BLOOD EXAMINATION





Selective & Scientific Books Publisher & Distributors

A-22/1, Akash Vihar, Street No.1, Bakkarwala More, Nangloi-Najafgarh Road, New Delhi-110043 Mob.: 09810766022, 09810766722 * Website : www.ssbook.in E-mail : selectivebook@gmail.com / ssbook@rediffmail.com / info@ssbook.in Price: 395.00



Organic Chemistry Laboratory Course Book

(Preparations, Quantitative Analysis and Isolation of Organic Compounds from Natural Sources)

Dr. Pradip V. Tekade

M.Sc. (Chem.), Ph.D., NET, SET, GATE. Associate Professor in Chemistry Jankidevi Bajaj College of Science Wardha, Maharashtra (Autonomous)

Selective & Scientific Books

A-22/1, Akash Vihar, Street No.1, Bakkarwala More, Nangloi-Najafgarh Road, New Delhi-110043 Mob. 09810766022, 098107667221 E-mail: selectivebook@gmail.com/ssboks@rediffmail.com Website: www.foresicbookstore.com

Organic Chemistry Laboratory Course Book

(Preparations, Quantitative Analysis and Isolation of Organic Compounds from Natural Sources)

© 2017, All rights reserved, no part of this publication may be reproduced, stored in a retrival system or transmitted in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without the prior written persimmsion of the Publisher / Author,

First Edition		2018
(C)		Publishers
ISBN		978-81-89128-66-1
Price	: J	395/- cls Bhay Jush + 1 (1, 1, 1, 1, 1, 28/05/18 28/05/18

Selective & Scientific Books

A-22/1, Akash Vihar, Street No.1, Bakkarwala More, Nangloi-Najafgarh Road, New Delhi-110043 Mob. 09810766022, 098107667221 E-mail: selectivebook@gmail.com/ssboks@rediffmail.com Website: www.forensicbookstore.com

CONTENTS

PART-A

Elementary organic preparations with mechanisms

S.No.	Preparation/Synthesis	Page No.
V.	Dibenzalacetone from benzaldehyde (Aldol Condensation).	1
2	Benzhydrol from benzophenone (Reduction).	4
3.	Benzyl alcohol from benzaldehyde (Cannizaro Reaction)	6
4	Diethyl 1,4-dihydro-2,6-dimethyl-4-phenylpyridine-3,-5 dicarboxylate from ethyl acetoacetate (Hantzsch synthesis)	9
3.	Methyl orange from sulphanilic acid (Sandmeyer reaction: dye preparation)	12
6.	Fluorescein and eosin	15
N.	P-red from p-nitroaniline(Sandmeyer reaction: dye preparation)	18
8	Phenol- formaldehyde resin	20
9.	Anthranilic acid from phthalic anhydride (Phthalic anhydride→ phthalimide→ anthranilic acid)	22
J18,	4-Aryl-6-Methyl-3,4-dihydro-2(1H)-pyrimidinon ester from ethyl acetoacetate (Biginelli reaction)	26
11.	P-Chlorotoluene from p-toluidine (Sandmeyer reaction)	28
12.	β-benzoylpropionic acid from succinic anhydride (Friedal craft acylation reaction)	30
,18.	4-Benzylidine-2-phenyl-oxazole-5-one from glycin (Glycin→Benzoyl glycin→Benzylidine-2-phenyl-oxazole-5-one)	32
14.	Oxalic acid from sucrose	35
ys.	2,4,6-tribromoacetanilide from aniline (Aniline→2,4,6 tribromoaniline →2,4,6-tribromoacetanilide)	36
5 6.	M-nitroaniline from nitrobenzen (Nitrobenzene → m-nitroaniline)	39
NT.	p-nitroaniline from acetanilide (Acetanilide→ p-nitroacetanilide →p-nitroaniline)	42
18.	Adipic acid from cyclohexanol (Oxidation reaction).	45

44	ty asced multi-stage organic preparations with mechanism Page	No.
	Preparation/Synthesis	48
1.44	p-animumberuture dummente + p-animumberuene)	
£.	Diversit from bouron bouron + boury + books of	- 55
×	p-bremsiniles from uniter native -+ svetaniste -+ p-bremssertuniste -+ p-bromsämitus)	59
k	p-sisonnine from andine (antime → sortantiste → p-otroscettantilde → p-otroscettanti	47
£.	Henrik acii fun benzideliyê: (Beanskeliyê: + benzin → benzi) → benzik aciê)	68
ě.	p ashihenevia aod iron p-atronolazite (p-atrodolazite - p-atrobenzoic acid) (p-atrodolazite - p-atrobenzoic acid)	10
R.	2-hydroxy shalcone from peterol (p-ment) + p-mexylacement + 2-hydroxy-5-mentyl accordinate (p-ment) + p-mexylacement + 2-hydroxy-5-mentyl accordinate (p-ment) + p-mentyl + 2-hydroxy-5-mentyl accordinate (p-mentyl + p-mentyl) + 2-hydroxy-5-mentyl accordinate (p-mentyl + p-mentyl + p-men	2
	4,5-ditydrs-1,1.5-urglanyl-1H-pynaols ironthenroldditydr (Henzaldenyde → henzilidaneacetopherons → 6,5-ditydro-1,3,5- urghenyl-1H-pynaole)	
P	2-aminobearthlawshifteen militte (Aniline phenylthiocarbamide 1-aminobeaethiseole)	
h	2.4- dminophenylhydrazute from chlorobenzene 10 hlorobenzeneg2.4-drazochlorobinzene →2.4 dminophenylsydrazute	1
it.	>-placeytendole from anotoplammer (Accounter of the placey) hydrocome → 2-planylioidole)	
12	2.4.5-suplamy/oxamile from hyracon (Benzium + herazon herazonte + 2,4,5-tripheny) oxazole)	2
11	Benzopinacolone from benzopitenene (Benzophenene → benzopitacol → benzopitacolone)(Photochemicol preparation)	
×	Benzamlide from benzephenone Olenzophenone + benzephenotoxime + benzemlide)	
<u>)</u> 5:	Orange II from andline (Andline + auline hydrogen udplinte + udphanilie acid + Orange II)	
An.		1.2

620	pschloroberning: and from pistuals anny gran	1100
	(Philiatic ankydrale + philiation de + arthranilie acid + u-chlavofurucus)	acid)
150	Dighenic acid from plthaliz unbydridu	118
8.4	(Methalic, unbydride + philadiniide + miltranilic acid + diplomie acid)	

 4,4-dimme 3 metryl-pyrand 5 and from distant and a supervised of (Envyl action center + 3-metryl-pyrand 5-env +4,4-dimensio-3- methyl pyranol-5-env)

PART-C

Quantitative Estimation

S.N.	Formation	Fage No.
1000	Quantative Estimation of vitamin-C indometrically	115
	Quantanise Estimation of planat by KBrO, KBr	118
2 1	Quantitative Estimation of anine by monsine' bromide solution	301
A.	Quantitative Estimation of formuldyde by indemetry	734
5.	Quantitative Estimation of glacose by Hennikal's solution	126
4	Quantitative Estimation of carbonyl compound by healrances formation	128
A.	Committeeve Forimation of aldelayde by midation method	139
8	Quantitative Determination of percentage of number of hydroxy's arous in an organic compound by acets/ation method	1.44

PART-D

	ntitutive Analysis based on classical and instrumental	Page No.
S.No.	Quantizative analysis	134
1.	Quantitation analysis of Narogen by K glidail Method	137
2	Quantitative analysis of baloger by Suparary rethod	36
1	in the sector of ministry by Messenger's method	14
	Quantitative analysis of anins acids using multiod	
4	Quantitative analysis of proteins by Biant Method	1)#
5	Quantinance analysis of proceedings by untiling method	.19
0	Quantitative analysis of carbohydrates by anthrone method	14
1	Quantinative analysis of assurble anal	Ĥ
*	Quantitative analysis of aspein	

Solvent extraction of oil from oil seeds and determination of saponification value, iodine value 9.

PART-E

nic Compounds from Natural Source

	Isolation of Organic Composi-	Page No.
S.No.	Isolation	160
1.	Isolation of caffeine from tea leaves	162
2	Isolation of casein from milk.	163
3.	Isolation of lactose from milk	165
4.	Isolation of nicotine dipicrate from tobacco	167
5.	Isolation of piperine from black pepper	
6.	Isolation of lycopene from tomatoes	169
7.	Isolation of β-carotene from carrots	171
8.	Isolation of cysteine from hair	173
9.	Isolation of eugenol from cloves	175
10.	Isolation of (+) limonine from citrus rinds	177

154



MES'S INSTITUTE OF MANAGEMENT AND CAREER COURSES (IMCC) 131, MAYUR COLONY, KOTHRUD, PUNE 411038.

in collaboration with

SAVITRIBAI PHULE PUNE UNIVERSITY

CERTIFICATE OF APPRECIATION

AWARDED TO Dr. VILOS M. Ghadki FOR PUBLISHING/PRESENTING/PARTICIPATION PAPER ENTITLED Stochastic Comptimentation OF Analyzing The Relationship Between Two Set OF Nodes With Respect To Networking.

AT National Conference on Computer Technology , Management and it's Applications (NC2TMA-2018) March 27th - 28th, 2018

Dr. Santosh Deshpande Head of Department



Dr. Vikas Inamdar Director

05/12/2022, 17:59 STOCHASTIC COMPLEMENTATION OF ANALYZING THE RELATIONSHIP BETWEEN TWO SET OF NODES WITH RES...

editor@ijcea.com					About	Conference Support	Thesis Publications	Contact Us
Home	Authors	Call for Papers	Submission	Call for Editors	Editorial Board	Publications	Old Publication	S
				Interview Section				
					< Previous Next	>		

STOCHASTIC COMPLEMENTATION OF ANALYZING THE RELATIONSHIP BETWEEN TWO SET OF NODES WITH RESPECT TO NETWORKING

Chandrasekaran K Arun ¹, Dr. Vilas M Ghodki ², Dr. S.B. Kishor ³

¹Gondwana Research Scholar, Gadchiroli, Maharashtra, India

² Associate Professor, Dept. of Computer Science , J.B. College, Wardha, Maharashtra, India

³Head, Dept. of Computer Studies and Research, S.P College, Chandrapur, Maharsashtra, India

ABSTRACT:

A stochastic complementation concept is an idea which occurs naturally, although not always explicitly, in the theory and application of finite Markov chains. This paper brings stochastic complementation to the forefront with an explicit definition and a development of some of its properties and its applications with respect to networking. Furthermore, stochastic complementation are explored with respect to problems involving uncoupling procedures in the theory of Markov chains. Also, concept of stochastic complementation is used for analyzing the relationship between two set of nodes with respect to networking.

Keywords: Markov chains, stationary distributions, stochastic matrix, stochastic compelementation, nearly reducible systems, Simon-Ando theory



By admin | May 7th, 2018 | Conference Publications, NC2TMA-18 | 0 Comments

Share This Story, Choose Your Platform!

f 🎔 in 🍲 t 🦻 🕊 🖂

About the Author: admin





Save Tree! Save World?



STOCHASTIC COMPLEMENTATION OF ANALYZING THE RELATIONSHIP BETWEEN TWO SET OF NODES WITH RESPECT TO NETWORKING

Chandrasekaran K Arun¹, Dr. Vilas M Ghodki², Dr. S.B. Kishor³

¹Gondwana Research Scholar, Gadchiroli, Maharashtra, India

² Associate Professor, Dept. of Computer Science, J.B. College, Wardha, Maharashtra, India

³Head, Dept. of Computer Studies and Research, S.P College, Chandrapur, Maharsashtra, India

ABSTRACT:

A stochastic complementation concept is an idea which occurs naturally, although not always explicitly, in the theory and application of finite Markov chains. This paper brings stochastic complementation to the forefront with an explicit definition and a development of some of its properties and its applications with respect to networking. Furthermore, stochastic complementation are explored with respect to problems involving uncoupling procedures in the theory of Markov chains. Also, concept of stochastic complementation is used for analyzing the relationship between two set of nodes with respect to networking.

Keywords: Markov chains, stationary distributions, stochastic matrix, stochastic compelementation, nearly reducible systems, Simon–Ando theory

[1] INTRODUCTION

Traditional techniques like machine learning, statistical, pattern recognition, and data mining approaches (see, for example, [12]) usually assume a random sample of independent objects from a single relation. All these techniques have gone through the extraction of knowledge from data, almost always leading, in the end, to the classical double-entry tabular

STOCHASTIC COMPLEMENTATION OF ANALYZING THE RELATIONSHIP BETWEEN TWO SET OF NODES WITH RESPECT TO NETWORKING

format, containing features for a sample of the population. These features are therefore used in order to learn from the sample, provided that it is representative of the population as a whole. However, real-world data coming from many fields (such as World Wide Web, marketing, social networks, or biology; see [7]) are often multi relational and interrelated. The work recently performed in statistical relational learning [8], aiming at working with such data sets, incorporates research topics, such as link analysis [14], [19], web mining [1], [3], social network analysis [2], [20], or graph mining [5]. All these research fields intend to find and exploit links between objects (in addition to features—as is also the case in the field of spatial statistics [6], [17]), which could be of various types and involved in different kinds of relation-ships. The focus of the techniques has moved over from the analysis of the features describing each instance belonging to the population of interest (attribute value analysis) to the analysis of the links existing between these instances (relational analysis), in addition to the features. This paper is proposing to analyze the links exist between set of nodes in networking.

[2] SIMPLE CORRESPONDENCE ANALYSIS

As stated before, simple correspondence analysis (see, for instance, [9], [10], [12], [18]) aims to study the relation-ships between two random variables x_1 and x_2 (the features) having each mutually exclusive, categorical, outcomes, denoted as attributes. Suppose the variable x_1 has n_1 observed attributes and the variable x_2 has n_2 observed attributes, each attribute being a possible out- come value for the feature. An experimenter makes a series of measurements of the features x_1 ; x_2 on a sample of vg individuals and records the outcomes in a frequency (also called contingency) table, fij, containing the number of individuals having both attribute $x_1 = i$ and attribute $x_2 = j$. In our relational database, this corresponds to two tables, each table corresponding to one variable, and containing the set of observed attributes (outcomes) of the variable. The two tables are linked by a single relation (see Fig. 1 for a simple example). This situation can be modeled as a bipartite graph, where each node corresponds to an attribute and links are only defined between attributes of x_1 and attributes of x_2 . The weight associated to each link is set to $w_{ij} = f_{ij}$, quantifying the strength of the relationship between i and j. The

 $\mathbf{A} = \begin{bmatrix} \mathbf{Q}_{11} & \mathbf{Q}_{22}^{\mathrm{st}} \end{bmatrix}, \ \mathbf{P} = \begin{bmatrix} \mathbf{Q}_{11} & \mathbf{P}_{22}^{\mathrm{st}} \end{bmatrix},$

associated n * n adjacency matrix and the corresponding transition matrix can be factorized as where O is a matrix full of zeroes. Suppose we are interested in studying the relationships between the attributes of the first variable x1, which corresponds to the n1 first elements. By stochastic complementation (see (10)), $P_c=P_{12}P_{21}= \square_1^{-1} \wedge_{12} \square_2^{-1} \wedge_{21}$. Computing the diffusion map for t = 1aims to extract the subdominant right-hand eigenvectors of P_c , which exactly corresponds to correspondence analysis (see, for instance, [24], (4.3.5)). Moreover, it can easily be shown that P_c has only real nonnegative eigenvalues, and thus, ordering the eigenvalues by modulus is equivalent to ordering them by value. In correspondence analysis, eigenvalues reflect the relative importance of the dimensions: each eigenvalue is the amount of inertia a given dimension exhibits in the frequency table [12]. The basic diffusion map after stochastic complementation on this bipartite graph therefore leads to the same results as simple correspondence analysis. Relationships between simple correspondence analysis and link analysis techniques have already been highlighted. For instance, Zha et al. [24] showed the

equivalence of a normalized cut performed on a bipartite graph and simple correspondence analysis. On the other hand, Saerens et al. investigated the relationships between Kleinberg's HITS algorithm [25], and correspondence analysis [18] or princi-pal component analysis [16].

[3] MULTIPLE CORRESPONDENCE ANALYSIS

Multiple correspondence analysis assigns a numerical score to each attribute of a set of p > 2 categorical variables [23], [18]. Suppose the data are available in the form of a star-schema: the individuals are contained in a main table and the categorical features of these individuals, such as education level, gender, etc., are contained in p auxiliary, satellite, tables. The corresponding graph is built naturally by defining one node for each individual and for each attribute while a link between an individual and an attribute is defined when the individual possesses this attribute. This configuration is known as a star-schema [13] in the data warehouse or relational database fields (see Fig. 2 for a trivial example).Let us first renumber the nodes in such a way that the attribute nodes appear first and the individuals nodes last. Thus, the attributes-to-individuals matrix will be denoted by A_{12} , it contains a 1 on the (i, j) entry when the individual j has attribute i, and 0 otherwise. The individuals-to-attributes matrix, the transpose of the attributes-to-individuals matrix, is A_{21} . Thus, the adjacency matrix of the graph is



Now, the individuals-to-attributes matrix exactly corresponds to the data matrix $A_{21} = X$ containing, as rows, the individuals and, as columns, the attributes. Since the different features are coded as indicator (dummy) variables a row of the X matrix contains a 1 if the individual has the corresponding attribute and 0 otherwise. We thus have $A_{21} = X$ and $A_{12} = X^T$. Assuming binary weights, the matrix D_1 contains on its diagonal the frequencies of each attribute, that is, the number of individuals having this attribute. On the other hand, D_2 contains p on each element of its diagonal, since each individual has exactly one attribute for each of the p features (attributes corresponding to a feature are mutually exclusive). Thus,

$$D_2 = p I \text{ and } P_{12} = D_1^{-1} A_{12}, P_{21} = D_2^{-1} A_{21},$$

Suppose we are first interested in the relationships between attribute nodes, thereby hiding the individual nodes contained in the main table. By stochastic complementation (4), the corresponding attribute-attribute transition matrix is

$$\begin{split} \mathbf{P}_{c} &= \mathbf{D}_{1}^{-1} \mathbf{A}_{12} \mathbf{D}_{2}^{-1} \mathbf{A}_{21} = \frac{1}{p} \mathbf{D}_{1}^{-1} \mathbf{A}_{12} \mathbf{A}_{21} \\ &= \frac{1}{p} \mathbf{D}_{1}^{-1} \mathbf{X}^{\mathrm{T}} \mathbf{X} = \frac{1}{p} \mathbf{D}_{1}^{-1} \mathbf{F}, \end{split}$$

where the element f_{ij} of the frequency matrix $F = X^T X$, also called the Burt matrix, contains the number of co-occurrences of the two attributes i and j, that is, the number of individuals having both attribute i and attribute j. The largest nontrivial right eigenvector of the matrix P_c represents the scores of the attributes in a multiple correspondence analysis. Thus, computing the eigenvalues and eigenvectors of Pc and displaying the nodes with coordinates proportional to the eigenvectors, weighted by the corresponding eigenvalue, exactly corresponds to multiple

STOCHASTIC COMPLEMENTATION OF ANALYZING THE RELATIONSHIP BETWEEN TWO SET OF NODES WITH RESPECT TO NETWORKING

correspondence analysis. This is precisely what we obtain when computing the basic diffusion map on P_c with t = 1. Indeed, as for simple correspondence analysis, it can easily be shown that Pc has real nonnegative eigenvalues, and thus, ordering the eigenvalues by modulus is equivalent to ordering by value.

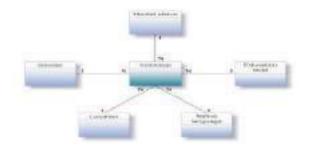


Fig. 1. Trivial example of a star-schema relation between a main variable, individual and auxiliary variables, Gender, Education level, etc. Each table contains outcomes of the corresponding random variable.

If we are interested in the relationships between elements of the main table (the individuals) instead of the attributes, we obtain,

$$\mathbf{P}_{\mathrm{t}} = \frac{1}{\nu} \mathbf{A}_{21} \mathbf{D}_{1}^{-1} \mathbf{A}_{12} = \frac{1}{\nu} \mathbf{X} \mathbf{D}_{1}^{-1} \mathbf{X}^{\mathrm{T}},$$

which, once again, is exactly the result obtained by multiple correspondence.

[4] ANALYZING RELATIONS BY STOCHASTIC COMPLEMENTATION

In the previous section, the concept of stochastic complementation is briefly reviewed and applied to the analysis of a graph through the random-walk-on-a-graph model. From the initial graph, a reduced graph containing only the nodes of interest, and which is much easier to analyze, is built.

4.1 COMPUTING A REDUCED MARKOV CHAIN BY STOCHASSTIC COMPLEMENTATION

Suppose we are interested in analyzing the relationship between two sets of nodes of interest. A reduced Markov chain can be computed from the original chain, in the following manner: First, the set of states is partitioned into two subsets, S_1 —corresponding to the nodes of interest to be analyzed—and S_2 —corresponding to the remaining nodes, to be hidden. We further denote by n1 and n2 (with n1 + n2 = n) the number of states in S_1 and S_2 , respectively; usually n2 > = n1. Thus, the transition matrix is repartitioned as

$$\mathbf{P} = \frac{\begin{array}{cc} S_{1} & S_{2} \\ S_{2} \begin{bmatrix} \mathbf{P}_{11} & \mathbf{P}_{12} \\ \mathbf{P}_{21} & \mathbf{P}_{22} \end{bmatrix}}{S_{2} \begin{bmatrix} \mathbf{P}_{11} & \mathbf{P}_{12} \\ \mathbf{P}_{21} & \mathbf{P}_{22} \end{bmatrix}}.$$

The idea is to censor the useless elements by masking them during the random walk. That is, during any random walk on the original chain, only the states belonging to S_1 are recorded; all the other reached states belonging to subset S_2 being censored, and therefore, not recorded. One can show that the resulting reduced Markov chain obtained by censoring the states S_2 is the stochastic complement of the original chain [15]. Thus, performing a stochastic complementation allows to focus the analysis on the tables and elements representing the factors/features of interest. The reduced chain inherits all the characteristics from the original chain; it simply censors the useless states. The stochastic complement Pc of the chain, partitioned as in (9), is defined as (see, for instance, [15])

$$\mathbf{P}_{c} = \mathbf{P}_{11} + \mathbf{P}_{12}(\mathbf{I} - \mathbf{P}_{22})^{-1}\mathbf{P}_{21}$$

It can be shown that the matrix P_c is stochastic, that is, the sum of the elements of each row is equal to 1 [15]; it therefore corresponds to a valid transition matrix between states of interest. We will assume that this resulting stochastic matrix is aperiodic and irreducible, that is, primitive [26]. Indeed, Meyer showed in [15] that if the initial chain is irreducible or aperiodic, so is the reduced chain. Moreover, even if the initial chain is periodic, the reduced chain frequently becomes aperiodic by stochastic complementation [15]. One way to ensure the aperiodicity of the reduced chain is to introduce a small positive quantity on the diagonal of the adjacency matrix A, which does not fundamentally change the model. Then, P has nonzero diagonal entries and the stochastic complement, P_c , is primitive. Let us show that the reduced chain also represents a random walk on a reduced graph G_c containing only the nodes of interest. We therefore partition the matrices

$$\mathbf{A} = \begin{bmatrix} \mathbf{A}_{11} & \mathbf{A}_{12} \\ \mathbf{A}_{21} & \mathbf{A}_{22} \end{bmatrix}; \ \mathbf{D} = \begin{bmatrix} \mathbf{D}_1 & \mathbf{O} \\ \mathbf{O} & \mathbf{D}_2 \end{bmatrix}; \ \mathbf{L} = \begin{bmatrix} \mathbf{L}_{11} & \mathbf{L}_{12} \\ \mathbf{L}_{21} & \mathbf{L}_{22} \end{bmatrix}$$

from which we easily find $P_c = D_1$. Notice that if A is symmetric (the graph G_c is undirected), Ac is symmetric as well. Since P_c is stochastic, we deduce that the diagonal matrix D_1 contains the row sums of Ac and that the entries of A_c are positive. The reduced chain thus corresponds to a random walk on the graph G_c whose adjacency matrix is A_c . Moreover, the corresponding Laplacian matrix of the graph G_c can be obtained by

$$\begin{split} \mathbf{L}_{c} &= \mathbf{D}_{1} - \mathbf{A}_{z} = (\mathbf{D}_{1} - \mathbf{A}_{11}) - \mathbf{A}_{12}(\mathbf{D}_{2} - \mathbf{A}_{22})^{-1}\mathbf{A}_{21} \\ &= \mathbf{L}_{11} - \mathbf{L}_{12}\mathbf{L}_{21}^{-1}\mathbf{L}_{21} \end{split}$$

since $L_{12} = A_{12}$ and $L_{21} = A_{21}$. If the adjacency matrix A is symmetric, L_{11} (L_{22}) is positive definite, since it is obtained from the positive semi definite matrix L by deleting the rows associated to S2 (S1) and the corresponding columns, thereby eliminating the linear relation-ship. Notice that L_c is simply the Schur complement of Thus, for an undirected graph G, instead of directly computing P_c , it is more interesting to compute L_c , which is symmetric positive definite, from which Pc can easily be deduced.

STOCHASTIC COMPLEMENTATION OF ANALYZING THE RELATIONSHIP BETWEEN TWO SET OF NODES WITH RESPECT TO NETWORKING

[4] EXPERIMENTS

This experimental section aims to answer four research question. Does the proposed stochastic complementation applied to the analysis of a graph through the random-walk-on-a-graph model provide realistic subgraph drawings?

4.1 Analyzing the Effect of Stochastic Complementation on a Newsgroups Data Set

The real-world data set studied in this section is the newsgroups data set. It is composed of about 20,000 un-structured documents, taken from 20 discussion groups (newsgroups) of the Usernet diffusion list. For the ease of interpretation, we decided to limit the data set size by randomly sampling 150 documents out of three slightly correlated topics ("sport/baseball," "politics/mideast," and "space/general"; 50 documents from each topic). Those 150 documents are preprocessed as described in [21]. The resulting graph is composed of 150 document nodes, 564 term nodes, and three topic nodes representing the topics of the documents. Each document is connected to its corresponding topic node with a weight fixed to 1. Thus, the class (or topics) nodes are connected to document nodes of the corresponding topics, and each document is also connected to terms contained in the document. Drawing a parallel with our illustrative example (see Fig. 5), topic nodes correspond to c-nodes, document nodes to e-nodes, and terms to a-nodes. The goal of this experiment is to study the similarity between the terms and the topics through their connections with the document nodes. The reduced Markov chain is computed by setting S₁ to the nodes of the graph corresponding to the terms and the topics.

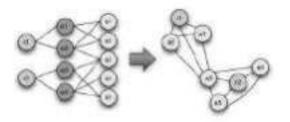


Fig. 2. Example illustrating stochastic implementation

The remaining nodes (documents) are rejected in the subgroup S_2 . The KDM PCA quickly provides the same mapping as the basic diffusion map when increasing the value oft. However, it can be observed on the embedding with t = 1 that several nodes are rejected outside the "triangle," far away from the other nodes of the graph. A new mapping, where the terms corresponding to each node are also displayed (for the visualization convenience, only terms cited by 25 documents or more are shown) for KDM PCA with t = 1.It can be observed that a group of terms are stuck near each topic nodes, denoting their relevance to this topic (i.e., "player," "win," "hit," and "team" for the sport topic). We also observe that terms lying in-between two topics are also commonly used by both topics ("human," "nation," and "European" seem to be words used in discussions on politics as well as on space),or centered in the middle of the three topics (common terms without any specificity, such as "work," "Usa," or "make"). Globally, the projection provides a good representation of the use of the terms in the different discussion groups for both the basic diffusion map and the KDM PCA. Terms rejected outside the "triangle" are often only

cited by few documents and seem to have few links with other terms. They are probably out of topic as, for instance, the series of terms on printer in the outlier group on the left of the sport topic ("printer," "Packard," or "Hewlett").

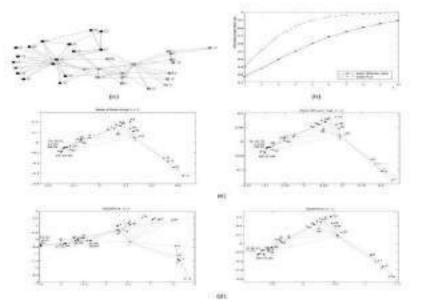


Fig. 2. Zachary Karate Social Network

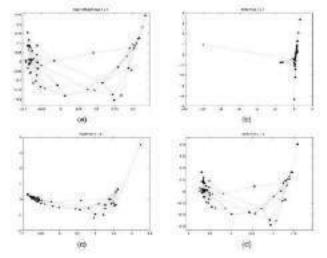


Fig 3. The graph mapping obtained by the diffusion map

4.2 DISCUSSION OF THE RESULTS

Let us now come back to our research questions. As a first observation, we can say that the two-step procedure (stochastic complementation followed by a diffusion map projection) provides an embedding in a low-dimensional subspace from which useful information can be extracted. Indeed, the experiments show that highly related elements are displayed close together while poorly related elements tend to be drawn far apart. This is quite similar to correspondence analysis to which the procedure is closely related. Second, it seems that stochastic complementation reasonably preserves proximity information, when combined with a diffusion map (KDM PCA) or an ISOMAP projection (MDS). For the diffusion map, this is normal, since both stochastic complementation and the diffusion map distance are based on a Markov chain

STOCHASTIC COMPLEMENTATION OF ANALYZING THE RELATIONSHIP BETWEEN TWO SET OF NODES WITH RESPECT TO NETWORKING

model—stochastic complementation is the natural technique allowing to censor states of a Markov chain. On the contrary, stochastic complementation should not be combined with a Laplacian Eigenmap, a curvilinear component analysis, or a Sammon nonlinear mapping—the resulting mapping is not accurate. Finally, the KDM PCA provides exactly the same results as the basic diffusion map when t is large. However, when the parameter t is low, the resulting projection tends to highlight the outlier nodes and to magnify the relative differences between nodes. It is therefore recommended to display a whole range of mappings for several different values of t.

[5] CONCLUSIONS AND FURTHER WORK

This work proposed to apply stochastic complementation in the analysis of a graph through the random-walk-on-a-graph model and provides realistic subgraph drawings. Here, the network is viewed as a graph, where the nodes correspond to the elements contained in the sets and the links correspond to the relations between the sets. It seems that stochastic complementation reasonably preserves proximity information, On the contrary, stochastic complementation should not be combined with a Laplacian Eigenmap, a curvilinear component analysis, or a Sammon nonlinear mapping—the resulting mapping is not accurate. Further work will be devoted to the combining stochastic implementation with a Laplacian Eigenmap, a curvilinear component analysis, or a Sammon nonlinear mapping—and produce the resulting mapping is accurate.

REFERENCES

[1]P. Baldi, P. Frasconi, and P. Smyth, Modeling the Internet and the Web: Probabilistic Methods and Algorithms. John Wiley & Sons, 2003.

[2]P. Carrington, J. Scott, and S. Wasserman, Models and Methods in Social Network Analysis. Cambridge Univ. Press, 2006.

[3]S. Chakrabarti, Mining the Web: Discovering Knowledge from Hypertext Data. Elsevier Science, 2003.

[4] F.R. Chung, Spectral Graph Theory. Am. Math. Soc., 1997.

[5]D.J. Cook and L.B. Holder, Mining Graph Data. Wiley and Sons, 2006.

[6] N. Cressie, Statistics for Spatial Data. Wiley, 1991.

[7]P. Domingos, "Prospects and Challenges for Multi-Relational Data Mining," ACM SIGKDD Explorations Newsletter, vol. 5, no. 1,pp. 80-83, 2003.

[8] Introduction to Statistical Relational Learning, L. Getoor and B. Taskar, eds. MIT Press, 2007.

[9] J. Gower and D. Hand, Biplots. Chapman & Hall, 1995.

[10] M.J. Greenacre, Theory and Applications of Correspondence Analysis. Academic Press, 1984.

[11]T. Hastie, R. Tibshirani, and J. Friedman, The Elements of Statistical Learning: Data Mining, Inference, and Prediction, second ed. Springer-Verlag, 2009.

[12]R. Johnson and D. Wichern, Applied Multivariate Statistical Analysis, sixth ed. Prentice Hall, 2007.

[13]R. Kimball and M. Ross, The Data Warehouse Toolkit: The Complete Guide to Dimensional Modeling. John Wiley & Sons, 2002.

[14]A.N. Langville and C.D. Meyer, Google's PageRank and Beyond: The Science of Search Engine Rankings. Princeton Univ. Press, 2006.

[15]C.D. Meyer, "Stochastic Complementation, Uncoupling Markov Chains, and the Theory of Nearly Reducible Systems," SIAM Rev.,vol. 31, no. 2, pp. 240-272, 1989.

[16]M. Saerens and F. Fouss, "HITS Is Principal Component Analysis," Proc. 2005 IEEE/WIC/ACM Int'l Joint Conf. Web Intelligence, pp. 782-785, 2005.

[17]O. Schabenberger and C. Gotway, Statistical Methods for Spatial Data Analysis. Chapman & Hall, 2005.

[18]M. Tenenhaus and F. Young, "An Analysis and Synthesis of Multiple Correspondence Analysis, Optimal Scaling, Dual Scaling, Homogeneity Analysis and Other Methods for Quantifying Categorical Multivariate Data," Psychometrika, vol. 50, no. 1, pp. 91-119, 1985.

[19]M. Thelwall, Link Analysis: An Information Science Approach. Elsevier, 2004.

[20]S. Wasserman and K. Faust, Social Network Analysis: Methods and Applications. Cambridge Univ. Press, 1994.

[21] L. Yen, F. Fouss, C. Decaestecker, P. Francq, and M. Saerens, "Graph Nodes Clustering Based on the Commute-Time Kernel," Proc. 11th Pacific-Asia Conf. Knowledge Discovery and Data Mining (PAKDD '07), 2007.

[22] L. Yen, F. Fouss, C. Decaestecker, P. Francq, and M. Saerens, "Graph Nodes Clustering with the Sigmoid Commute-Time Kernel: A Comparative Study," Data and Knowledge Eng., vol. 68,pp. 338-361, 2009.

[23]H. Zha, X. He, C.H.Q. Ding, M. Gu, and H.D. Simon, "Bipartite Graph Partitioning and Data Clustering," Proc. ACM 10th Int'l Conf. Information and Knowledge Management (CIKM '01), pp. 25-32, 2001.

[24] J. Gower and D. Hand, Biplots. Chapman & Hall, 1995.

[25]J.M. Kleinberg, "Authoritative Sources in a Hyperlinked Environment," J. ACM, vol. 46, no. 5, pp. 604-632, 1999.

Author[s] brief Introduction

Corresponding Address-(Pin code and Mobile is mandatory)



MES'S INSTITUTE OF MANAGEMENT AND CAREER COURSES (IMCC) 131, MAYUR COLONY, KOTHRUD, PUNE 411038.

in collaboration with

SAVITRIBAI PHULE PUNE UNIVERSITY

CERTIFICATE OF APPRECIATION

AWARDED TO Dr. VIJOS M. Ghodki FOR PUBLISHING/PRESENTING/PARTICIPATION PAPER ENTITLED A Study OF Least Square-Support Vector Machine Based Intrusion Detection System To Defect Denial-of Service Attacks.

AT National Conference on Computer Technology , Management and it's Applications (NC2TMA-2018) March 27th - 28th, 2018

Dr. Santosh Deshpande Head of Department

Dr. Vikas Inamdar Director 05/12/2022, 17:57 A STUDY OF LEAST SQUARE-SUPPORT VECTOR MACHINE BASED INTRUSION DETECTION SYSTEM TO DETECT ...

editor@ijcea.com	I				About	Conference Support	Thesis Publications	Contact Us
Home	Authors	Call for Papers	Submission	Call for Editors	Editorial Board	Publications	Old Publications	
				Interview Section				

A STUDY OF LEAST SQUARE-SUPPORT VECTOR MACHINE BASED INTRUSION DETECTION SYSTEM TO DETECT DENIAL-OF SERVICE ATTACKS

Ms. Koustubha Madhavi B¹, Dr. Vilas M Ghodki², Dr. S.B.Kishor³

¹Gondwana Research Scholar, Gadchiroli, Maharashtra, India

² Associate Professor, Dept. of Computer Science , J.B. College, Wardha, Maharashtra, India

³Head, Dept. of Computer Studies and Research, S.P College, Chandrapur, Maharsashtra, India

ABSTRACT:

Attack detection is one of the most important issues for computer networks security. The presence of redundant and irrelevant features in data have always caused a long-term problems in network traffic classification. Such features, in addition to slowing down the process of classification they also prevent the classifier from achieving accuracy in making decisions while dealing with large amounts of data. The performance of network intrusion detection systems based on machine learning techniques largely depends on the selected features. In this paper, we study and explore a new feature selection algorithm called mutual information which analytically selects the optimal feature for classification. This algorithm is designed to handle features that are both linear and non-linear in nature. This new feature selection algorithm is used to construct an Intrusion Detection System (IDS). Experiments are carried out to determine the performance on three intrusion detection evaluation datasets, namely KDD Cup 99, NSL-KDD and Kyoto 2006+ dataset. The evaluation results show that our feature selection algorithm contributes more critical features for LSSVM-IDS to achieve better accuracy and lower computational cost compared with the state-of-the-art methods.

Keywords: Feature Selection , Intrusion Detection , DoS Attacks



By admin | May 7th, 2018 | Conference Publications, NC2TMA-18 | 0 Comments

Share This Story, Choose Your Platform!

🤊 🖬 🐨 t 🖗 🛚 🗹





A STUDY OF LEAST SQUARE-SUPPORT VECTOR MACHINE BASED INTRUSION DETECTION SYSTEM TO DETECT DENIAL-OF SERVICE ATTACKS

Ms. Koustubha Madhavi B¹, Dr. Vilas M Ghodki², Dr. S.B.Kishor³

¹Gondwana Research Scholar, Gadchiroli, Maharashtra, India ² Associate Professor, Dept. of Computer Science, J.B. College, Wardha, Maharashtra, India

³*Head, Dept. of Computer Studies and Research, S.P College, Chandrapur, Maharsashtra, India*

ABSTRACT:

Attack detection is one of the most important issues for computer networks security. The presence of redundant and irrelevant features in data have always caused a long-term problems in network traffic classification. Such features, in addition to slowing down the process of classification they also prevent the classifier from achieving accuracy in making decisions while dealing with large amounts of data. The performance of network intrusion detection systems based on machine learning techniques largely depends on the selected features. In this paper, we study and explore a new feature selection algorithm called mutual information which analytically selects the optimal feature for classification. This algorithm is designed to handle features that are both linear and non-linear in nature. This new feature selection algorithm is used to construct an Intrusion Detection System (IDS). Experiments are carried out to determine the performance on three intrusion detection evaluation datasets, namely KDD Cup 99, NSL-KDD and Kyoto 2006+ dataset. The evaluation results show that our feature selection algorithm contributes more critical features for LSSVM-IDS to achieve better accuracy and lower computational cost compared with the state-of-the-art methods.

Keywords: Feature Selection, Intrusion Detection, DoS Attacks

A STUDY OF LEAST SQUARE-SUPPORT VECTOR MACHINE BASED INTRUSION DETECTION SYSTEM TO DETECT DENIAL-OF-SERVICE ATTACKS

[1] INTRODUCTION

Intrusion detection system (IDS) is a complement of traditional networks protection techniques namely user authentication, data encryption, and firewall as the first line of defense for computer and networks security. IDS have been recognized as intense research area in the past decade owing to the rapid increase of sophisticated attacks on computer networks [1]. The objective of IDS is to detect any anonymous or unusual activity as an attempt of breaking the security policy of computer networks. There are three broad categories of detection approaches: 1) classification; 2) data clustering; and 3) anomaly-based approach. In the classification approach, we classify the given data set into different types of attacks. Data classification is a supervise machine learning technique. In the second data clustering approach, we categorize the given data set into different categories on the basis of similarity and dissimilarity. Data clustering is an unsupervised machine learning technique. The third anomaly-based approach identifies deviations from the normal usage behavior patterns to identify the intrusion. In general, anomaly-based approach is semi supervised machine learning technique. Each approach has own advantages and disadvantages over the other approaches [2]. This paper address the first approach i.e. data classification for building intrusion detection model. There are many issues and challenges in the existing data classification approaches. The first is called imbalance class problem, where the number of examples of attack class is very rare. That is the data set distribution reflects a significant majority of normal class and a minority of attack class [3]. The second is to identify the appropriate classifier for intrusion detection from a large number of existing classifiers [4]. The third is pre-processing the raw data so that processed data can be used as input for a classifier. Accuracy of a classifier depends upon the quality of input data set. The quality of data depends upon the quality feature vector of data set.

[2] FEATURE SELECTION

Feature selection is a technique for eliminating irrelevant and redundant features and selecting the most optimal subset of features that produce a better characterization of patterns belonging to different classes. Methods for feature selection are generally classified into filter and wrapper methods [12]. Filter algorithms utilize an independent measure (such as, information measures, distance measures, or consistency measures) as a criterion for estimating the relation of a set of features, while wrapper algorithms make use of particular learning algorithms to evaluate the value of features. In comparison with filter methods, wrapper methods are often much more computationally expensive when dealing with highdimensional data or large-scale data. In this study hence, we focus on filter methods for IDS. Due to the continuous growth of data dimensionality, feature selection as a pre-processing step is becoming an essential part in building intrusion detection systems [13]. Mukkamala and Sung [14] proposed a novel feature selection algorithm to reduce the feature space of KDD Cup 99 dataset from 41 dimensions to 6 dimensions and evaluated the 6 selected features using an IDS based on SVM. The results show that the classification accuracy increases by 1 percent when using the selected features. Chebrolu et al. [15] investigated the performance in the use of a Markov blanket model and decision tree analysis for feature selection, which showed its capability of reducing the number of features in KDD Cup 99 from 41 to 12 features. Chen et al. [16] proposed an IDS based on Flexible Neural Tree

(FNT). The model applied a pre-processing feature selection phase to improve the detection performance. Using the KDD Cup 99, FNT model achieved 99.19 percent detection accuracy with only 4 features. Recently, Amiri et al. [12] proposed a forward feature selection algorithm using the mutual information method to measure the relation among features. The optimal feature set was then used to train the LS-SVM classifier and to build the IDS. Horng et al. [17] proposed an SVM-based IDS, which combines a hierarchical clustering and the SVM. The hierarchical clustering algorithm was used to provide the classifier with fewer and higher quality training data to reduce the average training and test time and to improve the classification performance of the classifier. Experimented on the corrected labels KDD Cup 99 dataset, which includes some new attacks, the SVM-based IDS scored an overall accuracy of 95.75 percent with a false positive rate of 0.7 percent.

[3] INTRUSION DETECTION FRAMEWORK BASED ON LEAST SQUARE SUPPORT VECTOR MACHINE

The framework of the proposed intrusion detection system is depicted in Fig. 1. The detection framework is comprised of four main phases: (1) data collection, where sequences of network packets are collected, (2) data preprocessing, where training and test data are preprocessed and important features that can distinguish one class from the others are selected, (3) classifier training, where the model for classification is trained using LS-SVM, and (4) attack recognition, where the trained classifier is used to detect intrusions on the test data. Support Vector Machine is a supervised learning method. It studies a given labeled dataset and constructs an optimal hyperplane in the corresponding data space to separate the data into different classes. They named this new formulation the Least Squares SVM (LS-SVM). LS-SVM is a generalized scheme for classification and also incurs low computation complexity in comparison with the ordinary SVM scheme [4].

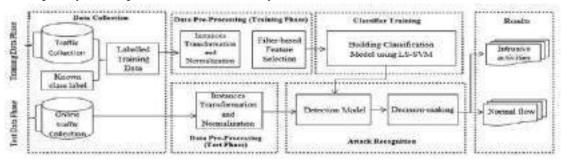


Figure 1: The framework for the LS-SVM based IDS

A. DATA COLLECTION

Data collection is the first and a critical step to intrusion detection. The type of data source and the location where data is collected from are two determinate factors in the design and the effectiveness of an IDS. To provide the best suited protection for the targeted host or networks, this study proposes a network-based IDS to test our proposed approaches. The proposed IDS runs on the nearest router to the victim(s) and monitors the inbound network traffic. During the training stage, the collected data samples are categorised with respect to the transport/Internet layer protocols and are labeled against the domain knowledge. However, the data collected in the test stage are categorised according to the protocol types only.

A STUDY OF LEAST SQUARE-SUPPORT VECTOR MACHINE BASED INTRUSION DETECTION SYSTEM TO DETECT DENIAL-OF-SERVICE ATTACKS

B. DATA PROCESSING

The data obtained during the phase of data collection are first processed to generate the basic features such as the ones in KDD Cup 99 dataset. This phase contains three main stages shown as follows.

C. Data Transferring

The trained classifier requires each record in the input data to be represented as a vector of real number. Thus, every symbolic feature in a dataset is first converted into a numerical value. For example, the KDD CUP 99 dataset contains

D. NSL-KDD Cup Dataset

The KDD Cup 1999 dataset is used as a benchmark for evaluating of IDS techniques [5]. The majority of examples in this data set have been extracted from the DARPA 1998 IDS evaluation [13]. KDD data set has a huge number of redundant examples. Duplicate examples may have a negative effect on the training process of machine learning classifiers. Through out in this empirical study, we used NSL-KDD Cup 99 dataset [7]. This data set is an improved version of KDD data set and publically available [7]. In this paper, KDD Train+ and KDD Test+ data sets that have 20% of the records of the entire NSL-KDD data set are used . The data set has 41 features. These features can be classified into four groups, namely basic features, content features, time-based features and host-based features. The details of these 41 features are presented in table I. Training data set consist of 29 different attacks. These attacks are further categorized into four different types and presented in Table 1. The brief introduction of four different types of attacks is as follows: 1) Denial of Service Attack (DoS): It is a class of attacks in which an attacker makes some computing or memory resource too busy or to handle legitimate requests, or denies legitimate users access to a machine. 2) Probing Attacks (Probe): It is a class of attacks in which an attacker scans a network of computers to gather information or find known vulnerabilities. An attacker with a map of machines and services that are available on a network can use this information to look for exploits. 3) Remote to Local Attacks (R2L): It is a class of attacks in which an attacker sends packets to a machine over a network but who does not have an account on that machine; exploits some vulnerability to gain local access as a user of that machine.

Do5	Probe	RIL	UIR
Neptune	Satan	Guess_Password	Buffer_overflow
Teardrop	Nmap	Warezmäster	Loadmodule
Land	Portsweep	Warenchent	rootkit
Smuf	IPSweep	Sendmail	
Pod		Multhop	
		flywrite	
		Imap	
		Spy	
		Phf	

Table 1: List of Attacks

E. Performance Evaluation

Several experiments have been conducted to evaluate the performance and effectiveness of the proposed LSSVM-IDS. For this purpose, the accuracy rate, detection rate, false positive rate and F-measure metrics are applied. The accuracy metric, detection rate and false positive rate are defined by

$$\begin{aligned} Accuracy &= \frac{TP + TN}{TP + TN + FN + FP}, \\ Detection \ Rate &= \frac{TP}{TP + FN}, \\ False \ Positive \ Rate &= \frac{FP}{FP + TN}, \end{aligned}$$

where True Positive (TP) is the number of actual attacks classified as attacks, True Negative (TN) is the number of actual normal records classified as normal ones, False Positive (FP) is the number of actual normal records classified as attacks, and False Negative (FN) is the number of actual attacks classified as normal records. The F-measure is a harmonic mean between precision p and recall r [44]. In other words, it is a statistical technique for examining the accuracy of a system by considering both precision and recall of the system. F-measure used in this paper assigns the same weights to both Precision Rate (PR) and Recall Rate (RR), and is given by

$$F\text{-measure} = \frac{2(Precision * Recall)}{Precision + Recall}$$

The precision (PR) is the proportion of predicted positives values which are actually positive. The precision value directly affects the performance of the system. A higher value of precision means a lower false positive rate and vice versa. The precision is given by

$$Precision = \frac{TP}{TP + FP}$$

Table 2 summarises the classification results of the different selection methods in regard to detection rates, false positive rates and accuracy rates. It shows clearly that the detection model combined with the FMIFS has achieved an accuracy rate of 99.79, 99.91 and 99.77 percent for KDD Cup 99, NSL-KDD and Kyoto 2006+, respectively, and significantly outperforms all other methods. In addition, the proposed detection model combined with FMIFS enjoys the highest detection rate and the lowest false positive rate in comparison with other combined detection models. The proposed feature selection algorithm is computationally efficient when it is applied to the LSSVM-IDS. Fig. 2 shows the building (training) and test times consumed by the detection model using FMIFS compared with the detection model using all features. The figure shows that the LSSVM-IDS + FMIFS performs better than LSSVM-IDS with all 41 features on all datasets. There are significant differences when performing experiments on KDD Cup 99 and NSL-KDD and a slight difference on Kyoto 2006+ dataset by comparison with the two aforementioned models.

A STUDY OF LEAST SQUARE-SUPPORT VECTOR MACHINE BASED INTRUSION DETECTION SYSTEM TO DETECT DENIAL-OF-SERVICE ATTACKS

		KOD Gp 示			381-410			Kato 2004		
2	19	鼎	kaniq	M	飛	Anny	M	RP	<i>inve</i> g	
LEWING	税准	1.8	税3	9.5	13	993	99.8	旧	99.77	
LSEVMEDS + MIPS () = 1.0	93	0.73	.901	藝術	13	69	98,9	曲	99.22	
LSEVIATES + MIRS(J=1)	93	03	96	63	16	16.75	9830	15	912	
LSEWHIDS + RLOS	85	(H	第日	625	14	装纸	983	182	989	
LSEVIALIS + All initiats	99J6	197	903	虹	13	15%	939	13	9.2	

Table 2 : Performance Classification for All Attacks Based on the Three Datasets

[4] CONCLUSION

Recent studies have shown that two main components are essential to build an IDS. They are a robust classification method and an efficient feature selection algorithm. In this paper, a supervised filter-based feature selection algorithm has been proposed, namely Flexible Mutual Information Feature Selection (FMIFS). FMIFS is an improvement over MIFS and MMIFS. FMIFS suggests a modification to Battiti's algorithm to reduce the redundancy among features. FMIFS eliminates the redundancy parameter b required in MIFS and MMIFS. This is desirable in practice since there is no specific procedure or guideline to select the best value for this parameter. FMIFS is then combined with the LSSVM method to build an IDS. LSSVM is a least square version of SVM that works with equality constraints instead of inequality constraints in the formulation designed to solve a set of linear equations for classification problems rather than a quadratic programming problem. The proposed LSSVM-IDS + FMIFS has been evaluated using three well known intrusion detection datasets: KDD Cup 99, NSL-KDD and Kyoto 2006+ datasets. The performance of LSSVMIDS + FMIFS on KDD Cup test data, KDDTest+ and the data, collected from Kyoto dataset has exhibited better classification performance in terms of classification accuracy, detection rate, false positive rate and Fmeasure than some of the existing detection approaches. In addition, the proposed LSSVM-IDS + FMIFS has shown comparable results with other state-of the-art approaches when using the Corrected Labels sub-dateset of the KDD Cup 99 dataset and tested on Normal, DoS, and Probe classes; it outperforms other detection models when tested on U2R and R2L classes. Furthermore, for the experiments on the KDD 21 dataset, LSSVM-IDS + FMIFS produces the best classification accuracy compared with other detection systems tested on the same dataset. Finally, based on the experimental results achieved on all datasets, it can be concluded that the proposed detection system has achieved promising performance in detecting intrusions over computer networks. Overall, LSSVM-IDS + FMIFS has performed the best when compared with the other stateof-theart models. Although the proposed feature selection algorithm FMIFS has shown encouraging performance, it could be further enhanced by optimising the search strategy

REFERENCES

- [1] S. Pontarelli, G. Bianchi, and S. Teofili, "Trafficaware design of a high-speed FPGA network intrusion detection system," IEEE Trans. Comput., vol. 62, no. 11, pp. 2322–2334, Nov. 2013.
- B. Pfahringer, "Winning the KDD99 classification cup: Baggedboosting," SIGKDD Explorations Newslett., vol. 1, no. 2, pp. 65–66, 2000.
- [3] I. Levin, "KDD-99 classifier learning contest: Llsoft's results overview," SIGKDD Explorations, vol. 1, no. 2, pp. 67–75, 2000.
- [4] D. S. Kim and J. S. Park, "Network-based intrusion detection with support vector machines," in Proc. Inf. Netw., 2003, vol. 2662, pp. 747–756.
- [5] A. Chandrasekhar and K. Raghuveer, "An effective technique for intrusion detection using neuro-fuzzy and radial SVM classifier," in Proc. Comput. Netw. Commun., 2013, vol. 131, pp. 499–507.
- [6] S. Mukkamala, A. H. Sung, and A. Abraham, "Intrusion detection using an ensemble of intelligent paradigms," J. Netw. Comput. Appl., vol. 28, no. 2, pp. 167–182, 2005.
- [7] A. N. Toosi and M. Kahani, "A new approach to intrusion detection based on an evolutionary soft computing model using neurofuzzy classifiers," Comput. Commun., vol. 30, no. 10, pp. 2201–2212, 2007.
- [8] Z. Tan, A. Jamdagni, X. He, P. Nanda, L. R. Ping Ren, and J. Hu, "Detection of denial-ofservice attacks based on computer vision techniques," IEEE Trans. Comput., vol. 64, no. 9, pp. 2519–2533, Sep. 2015.
- [9] A. M. Ambusaidi, X. He, and P. Nanda, "Unsupervised feature selection method for intrusion detection system," in Proc. Int.Conf. Trust, Security Privacy Comput. Commun., 2015, pp. 295– 301.
- [10] A. M. Ambusaidi, X. He, Z. Tan, P. Nanda, L. F. Lu, and T. U. Nagar, "A novel feature selection approach for intrusion detection data classification," in Proc. Int. Conf. Trust, Security Privacy Comput. Commun., 2014, pp. 82–89.
- [11] R. Battiti, "Using mutual information for selecting features in supervised neural net learning," IEEE Trans. Neural Netw., vol. 5, no. 4, pp. 537–550, Jul. 1994.
- [12] F. Amiri, M. RezaeiYousefi, C. Lucas, A. Shakery, and N. Yazdani, "Mutual information-based feature selection for intrusion detection systems," J. Netw. Comput. Appl., vol. 34, no. 4, pp. 1184–1199, 2011.
- [13] A. Abraham, R. Jain, J. Thomas, and S. Y. Han, "DSCIDs: Distributed soft computing intrusion detection system," J. Netw. Comput. Appl., vol. 30, no. 1, pp. 81–98, 2007.
- [14] S. Mukkamala and A. H. Sung, "Significant feature selection using computational intelligent techniques for intrusion detection," in Proc. Adv. Methods Knowl. Discovery Complex Data, 2005, pp. 285–306.
- [15] S. Chebrolu, A. Abraham, and J. P. Thomas, "Feature deduction and ensemble design of intrusion detection systems," Comput. Security, vol. 24, no. 4, pp. 295–307, 2005.

A STUDY OF LEAST SQUARE-SUPPORT VECTOR MACHINE BASED INTRUSION DETECTION SYSTEM TO DETECT DENIAL-OF-SERVICE ATTACKS

- [16] Y. Chen, A. Abraham, and B. Yang, "Feature selection and classification flexible neural tree," Neurocomputing, vol. 70, no. 1, pp. 305–313, 2006.
- [17] S.-J. Horng, M.-Y. Su, Y.-H. Chen, T.-W. Kao, R.J. Chen, J.-L. Lai, and C. D. Perkasa, "A novel intrusion detection system based on hierarchical clustering and support vector machines," Expert Syst. with Appl., vol. 38, no. 1, pp. 306–313, 2011.
- [18] G. Kim, S. Lee, and S. Kim, "A novel hybrid intrusion detection method integrating anomaly detection with misuse detection," Expert Syst. with Appl., vol. 41, no. 4, pp. 1690–1700, 2014.
- [19] P. Gogoi, M. H. Bhuyan, D. Bhattacharyya, and J. K. Kalita, "Packet and flow based network intrusion dataset," in Proc. 5thInt. Conf. Contemporary Comput., 2012, vol. 306, pp. 322–334.

In view of NEP, Syllabus of RTM Nagpur University

MATHEMATICS

B.Sc. Semester II (CBS)

Paper – I : Geometry, Differential & Difference Equations (M3) Paper – II : Vector Analysis (M4)



Vijay Soni
 Narendra Katre
 Rajendra Katre
 Ather Husain
 Edited by Dr. Rishi Agrawal

Himalaya Publishing House ISO 9001:2015 CERTIFIED

@ Author

No part of Mathematics B.Sc Semester - II (M3 & M4) may be reproduced, stored in a retrieval No part of Mathematics b.oc outries by any means, electronic, mechanical, photocopying, recording system, or transmitted in any form or by any means, electronic, mechanical, photocopying, recording and/or otherwise, without the prior written permission of the publishers.

Vijay Soni (Ch. 2, 3), Rishi Agrawal (Part of Ch. 6), Narendra Katre (Ch. 1, 4), Rajendra Katre (Ch. 5, 6), Ather Husain (Ch. 7, 8)

> First Edition : 2014, 2015, 2016, 2017, 2018, 2019, 2020 Revised Edition as Per New Syllabus : 2021

	-	
Published by		Mrs. Meena Pandey for Himalaya Publishing House Pvt. Ltd., "Ramdoot", Dr. Bhalerao Marg, Girgaon, Mumbai - 400 004. Phone: 022-23860170/23863863, Fax: 022-23877178 E-mail: himpub@bharatmail.co.in; Website: www.himpub.com
Branch Offices	3	
New Delhi	۰.	"Pooja Apartments", 4-B, Murari Lal Street, Ansari Road, Darya Ganj. New Delhi - 110 002. Phone: 011-23270392, 23278631; Fax: 011-23256286
Nagpur		Kundanial Chandak Industrial Estate, Ghat Road, Nagpur - 440 018. Phone: 0712-2738731, Telefax: 0712-2721215
Bengaluru	3	Plot No. 91-33, 2nd Main Road Seshdripuram, Behind Nataraja Theatre, Bengaluru-560020. Phone: 08041138821; Mob.: 9379847017, 9379847005.
Hyderabad	:	
Chennal	35	New No. 48/2, Old No. 28/2, Ground Floor, Sarangapani Street, T. Nagar. Chennai-600 012. Mobile: 09380460419
Pune	1	First Floor, "Laksha" Apartment, No. 527, Mehunpura, Shaniwarpeth (Near Prabhat Theatre), Pune - 411 030. Phone: 020-24496323/24496333; Mobile: 09370579333
Lucknow	:	House No 731, Shekhupura Colony, Near B.D. Convent School, Aliganj. Lucknow - 226 022, Mobile: 09307501549
Ahmedabad	3	114, "SHAIL", 1st Floor, Opp. Madhu Sudan House, C.G. Road, Navrang Pura, Ahmedabad - 380 009. Phone: 079-26560126; Mobile: 09377088847
Ernakulam	:	39/176 (New No: 60/251) 1st Floor, Karikkamuri Road, Ernakulam, Kochi - 682011, Phone: 0484-2378012, 2378016; Mobile: 09344199799 Plot No. 214/13/2/1550, 12-5
Cuttack	2	Plot No. 214/1342/1589, Budheswari Colony, Behind Durga Mandap, Cultack - 753 012, Orlinbo, Mahilian and State Sta
1227121012		Cultack - 753 012, Odisha; Mobile: 09338746007
Kolkata	1	Work, Dellanhate Main Deed to
DTPby	4	Kolkata - 700 010, Phone: 033-32449649, Mobile: 09883055590, 0743904030 HPH, Nagpur (Prasad)
Printed at	1	Geetanjali Press Pvt. Ltd., Nagpur, on behalf of HPH.
		NOW CONTRACTOR AND A CO

	法	CONTENTS	
		Paper - I : Geometry, Differential & Difference Equations (M3)	
¢	hapter - 1 5	SOLID GEOMETRY (Unit I)	Page
		Preliminaries	
		The Sphere	
		Equation of a Sphere in Different Forms	
	1.4.	Plane Section of a Sphere	
	1.5.	Intersection of Two Spheres	
	1.6.	Equation of a Circle	
	1.7.	Any Sphere through the Given Circle	
	1.8.	Intersection of a Sphere and a Line	
	1.9	Tangent Plane	
	1.10	Angle of Intersection of Two Spheres	
	1.11	The Cone	
	1.12	The Cylinder	
đ	Chapter - 2	FIRST ORDER DIFFERENTIAL EQUATIONS (Unit - II)	
	2.1.	Introduction	21
	2.2.	Family of curves	
	2.3.	Exact Differential Equation	24
	2,4.	Integrating Factor	27
10	2.5.	Linear Differential Equation	9.11
	2.6.	Reduction of Order	215
50	Chapter - 3	SECOND ORDER LINEAR DIFFERENTIAL EQUTIONS (Unit - III)	and a sector
	3.1.	Introduction	21
	3.2.	The General Solution of the Homogeneous Equation	
	3.3.	To Find Unknown Solution by using Known Solution	P.C
	3.4.	Homogeneous Equation with Constant Coefficients	
	3.5.	The Method of Undetermined Coefficients	2.14
	3.6.	Method of Variation of Parameters	2.14
	Chapter - 4	DIFFERENCE FOUNTIONS (Unit IN)	
	4.1.	Introduction	12.2
	4.2.	Some Basic Concepts	4.1
90	4.3.	principal contraction and a second seco	0.005
	4.4.	Formation of Difference Equations	
	4.5.	structure as a state of the sta	
	4.6.	Complementary Function (C.F.)	
	4.7.	The second	
	4.8	The particular integral (P.I.)	4.14
	4.9	First order non-homogeneous linear equations	4.14
	4.10	First order linear equation with variable coefficients	4.23
	4.11	First order linear equation with variable coefficients Equation homogeneous in $u(x)$	
	4.12	Equations Reducible to Linear Equations with Constant Coefficients	
		The officients and consume Coefficients	4.24

				the second	12.11.11.1.2
Paper -	2: V	ector	Anal	ysis	(1114)

	Paper - 2: Vector Analysis (1914)	
	VECTOR DIFFERENTIATION (Unit - I)	
Chapter - 5	VECTOR DIFFERENTIATION (Call Sy Preliminaries	
5.1.	Preliminaries Differentiation of Vectors	-
5.2.	Differential Geometry	-lines
5.3.	Differential Geometry	2114
5.4.	Vector Differential Operator	Personal
Chapter - 6	DOUBLE AND TRIPLE INTEGRAL, BETA-GAMMA FUNCTIONS (Unit - 11)	
6.1.	Double Integral	10
6.2.	Change of Order of Integration	n
6.3.	Transformation of Double Integral in Polar Form	122.2
6.4.	Evaluation of Area by Double Integral	mul
6.5.	Triple Integral	
6.6.	Gamma Function	
6.7.	Beta Function	
Chapter - 7	VECTOR INTEGRATION (Unit - III)	
7.1.	Integral of a Function	
7.2.	Ordinary Integral of Vector Function	
7.5.	Line Integrals	
7.4.	Conservative Fields	
7.5.	Surface Integrais	
7.6.	Volume Integral	
Chapter - 8	VECTOR THEOREMS (Unit - IV)	11.11
8.1.	Introduction	1
8.2.	Greens Theorem in the Plane	-
8.3.	Greens Theorem in the Plane	ani-j
8.4.	The Gauss Divergence Theorem	and a
	The Stokes Theorem	-12

res

I)	Model Question Papers (M3)
	y model Paper 1 with solutions
	ay model Paper 2
	ny wodel Paper 3
~J.	Model Question Paners (144)
	ii) Model Paper 3

ABOUT THE AUTHORS



Vijay M. Soni (MiSe., MISE) form Articleus freely for in Originithant of Mathematics, 2015. Science College, Gonda with 27 years of explainty experience in LIG level. The inferent free in simplifying Mathematics for novies. "Statistic are the least officie that a treasfer models into responsible citizens" in hit convintion. Theory of Relativity is hiteform. He class has extended in assemptibiling social work discogle various (NGO).



Nareatine T. Kette (M.Su, EDE), SET0 to working as an Artistian Professor and Head-Department of vibilitoriality, vibilitation desired, Keto). He has a traching experiment of 12 years at U() and 11 years of 00 level. He has been honored by (Galil Media) for resuring Prior Mark position to M.Su, extramation of Mitthansation fifth areas of indirect are Number Treesy, Integy Multimentics and Solarizing.



Rejenting T. Kenne (M.S., Balteli, SET) is working as an Architect Decision in Department of Mathematics, Bara Colloge of Sidence, Workhay He has a certaining experience of 17 years in the facility of Englishing and Science. The secured White Martip edition in M.Ss. Examination of Mathematics. The believes only in soft-dependency, hence mathematics studied cowards the same. He has an expert longing of Application Protoconful Department As a result of which, he has presented collidimentation research papers on Science and International Conferences.



Address Will Harverto (Miller, 600 https://www.dompar.com/assessment/factors/and/flood, Department of Mathematics, Miller, Morbill College Karania (Colf) having a methodogerpaniance of almost a describe at UC level. He record and their 20 mill, in 800 Historic June 2011. He mean of negative are Relativity. Cosmology and Moderial Theories of Gravitation. Also, teaching it a pression for him. He survey believes the "Truth always gives premiumon results". He charge individues students towards the real world and says flattice do the finant work smartly.

Edited by



Rishi K. Agrawal (M.Sc. NET, B.Ed. Br.D.) sorving at Histop College, Nugpar is an Associate Professor and Head, Department of Minimumoria has a reaching experience of 22 years at UG level. Since 2003, he has been working or a self-developed innovative teaching-learning method named 'GANBET DARSHAN'. With the same title, he is notive on YouTube. Working for popularization of Mathematics as a service to the energy with a strong belief that each student has his own mathematics laboratory in the form of his minti which is to be optical by a teacher

and les the students make proper use of it by thinking and visualising multernatios around them. He also believes that "Present main Present rate, to Eurore main Fature hat"

..... Jai Gangel *



	Rece S.S.S.K.R. (Control D	nt Trend Innani Mal cge with Potential for epartment of Vidya Bhar	OST Sponsored In s in Scien navidyalaya, Excellence Reaccredi In collaboration f Mathematic ati Mahavidy od Arts & Sci 22-23 March	Karanja (L Karanja (L Mad by NAAC at A'La s, SGBAU, A alaya, Amray ience College	echnology ad), Dist.Washin met CGP4-3.24) mravati vati &	m
	is is to certify tha	it Prof./Dr./Mr./				
	DEPARTMENT OF		and the second states and			
Tec	actively particip hnology held on UDY OF FUNGAL S	1 22 nd & 23 rd Ma	rch, 2018 and 1	as presented a	ent Trends in Science research paper entit RDHA CTTY(M.S)	& tled
duri A 41	ing technical sess	sion of the confe	erence.	The second	tt=f	dehil
Dr. J.P. Baxi Organizing Secretary	Dr. D.T. Dongare Organizing Secretary	Dr. D.R. Halwe Organizing Secretary	Dr. A.P. Charjan Co-Convener	Dr. S.D. Katore Co-Convener	Dr. F.C. Raghuwanshi Co-Conveaer	Dr. P.R. Rajput Convener





UGC (CPE) and DST Sponsored International Conference on Recent Trends in Science and Technology



22-23 March, 2018

PROCEEDINGS



Organized by Vidya Bharati Shaikshanik Mandal Amravati's

S.S.S.K.R.Innani Mahavidyalaya Karanja (Lad), Dist. Washim (M.S.)

Reaccredited at level 'A' grade by NAAC (CGPA 3.24) & Conferred CPE status by UGC,New Delhi. In collaboration with

Department of Mathematics & IQAC Sant Gadge Baba Amravati University, Amravati Vidya Bharati Mahavidyalaya, Amravati & Shri Dr.R.G.Rathod Science College, Murtizapur

Venue: S.S.S.K.R. Innani Mahavidyalaya Campus, Karanja (Lad) Email: iertst2018@gmail.com Website : www.ssskrimv.org.in Contact No.: 07256-222148/222171

Contents

Life Sciences

Sr. No.	Name of Author(s)	Title of Research Article	Page No.
LS1	Gawali, V.R., Deshmukh, S.D., Tayade, S.N. & Dabhade, D.S.	A Check List Of Some Soil Invertebrates Of Washim, Maharashtra, India	1
LS2	Sawsundar, A.S., Deshmukh, S.D., Tayade, S.N. & Dabhade, D.S.	Study Of Groundwater Quality Of Washim Town	6
LS3	Raut, N.M. & Deshmukh, U.S.	Diversity And Abundance Of Spiders (Aranae) In Salbardi Forest (Satpura Range), Maharashtra, India.	10
LS4	Bhagat, V.B. & Hingankar, A.P.	Comparative Study Of RBC Morphology With Peripheral Blood Smear Of Iron Deficient Anemia In Akot Region	15
LS5	Chaudhari, P.W. & Deshmukh, U.S.	Study Of Salticidae Fauna From Upper Wardha Dam, Dist. Amravati, Maharashtra, India.	18
LS6	Garode, A.M. & Bhusari, M.R.	Bacteriological Examination Of Filtered Water From Chikhali Municipal Corporation Water Filter At Various Stages.	22
LS7	Morey, C.D. & Tantarpale, V.T.	Water Quality Assessment Of Motala River And Nalganga Reservoir, Nalgangapur Dist. Buldana (M.S.) India	25
LS8	Mahajan, V.S. & Harney, N.V.	Macrophytes Diversity Of Mohabala Lake Near Bhadrawati, District - Chandrapur (Ms), India.	35
LS9	Ingole, A.B.	Serum Electrolytes(Na&K) Percentage Observed In Tribes Of Chikhaldara Region. (Melghat) Dist. Amravati (M.S.)	38
LS10	Wath, M.R. & Lande, P.M.	Assessment Of Anthelmintic Efficacy And Preliminary Phytochemical Investigations Of Invasive Weed Cassia Uniflora Mill.	41
LS11	Wakode, A.V. & Dhore, M.M.	Leafing, Flowering And Fruiting Pattern Of Some Species Of Fabaceae From Washim District (M.S.)	47
LS12	Deshmukh, A.S.	Spent Mushroom Substrate : Animal Feed	50
LS13	Sharma, R.R. & Thakare, P.V.	Isolation Of Bioactive Components From Chlorophytum Kolhapurens And Chlorophytum Bharuchae And Its Evaluation For Antioxidant Activity	54
LS14	Morey, J.P.	A Study Of Climate Changing Pattern In Vidharbha Region	61
LS15	Babare, A., Mishra, V. & Babare, M.	"Studies On Alternative Inanimal Dissection With E-Alternative Resources From Website"	64
LS16	Charjan, A.P., Joshi, P.S., Thakare, V.G. & Virani, R.S.	Effect Of Nimesulide On Hatching Rate Of Clarias Batrachus (Linnaeus, 1758)	67
LS17	Makode, P.M., Pandharikar, S.D., Bhise, J.V. & Gulhane, R.A.	Effects Of Dietary Onion On Behaviour Of The Fresh Water Fish Clarias Batrachus (Linnaeus, 1758)	69
LS18	Tantarpale, V.T., Puri, S.D., Manekar, S.P. & Gijare, S.S.	Anuran Inventory Of Vidarbha Region, Maharashtra, India	72
LS19	Pundlik, A.D., Katole, K.G., Raut, A.V. & Budhalkar, S.B.	Study On Physico-Chemical Parameters Of Lonar Lake Water	75

LS121	Jadhav, R.N.	Screening Of Azotobacter Sp From Rhizosphere Of Soybean Cultivated In Latur District Area For Phosphate Solubilization And Antifungal Activity	523
LS122	Thakur, S.B. & Kothale, K.V.	Preliminary Phytochemical Analysis Of Fruit Extracts Of Medicinally Important Plant Trichosanthes Cucumerina Linn.	527
LS123	Tippat, S.K.	Ecological Diversity Of Climbing Plants In Tropical Dry Deciduous Forest Of Melghat, Maharashtra, India	532
LS124	Kothale, K.V. & Deshmukh, M.K.	Anatomical And Priliminary Phytochemical Investigation Of Crotalaria Retusa L.	536
LS125	Solanke, A.K.	Toxicity Of Heavy Metal Mercury On Some Biochemical Parameters In Fresh Water Fish, Catla Catla	539
LS126	Joshi, P. & Mishra, R.	An Effect Of Irradiation On Water Bugs- Qualitative Study	542
LS127	Wanjare, P.D. & Anasane, P.Y.	Antimicrobial Activity And Phytochemical Screening Of Ailanthus Excelsa Bract	545
LS128	Shirsat, R., Jasutkar, J. & Aher, S.	Chemoprofiling And Antioxidant Potential Of A Lesser Known Leucasstricta	548
LS129	Wanjari, A.J.	Fish Diversity Of Saikheda Reservoir, Maharashtra State, India	552
LS130	Wankhade, M.R. & Kothale, K.V.	Preliminary Phytochemical Analysis Of Cordia Sebestena L.	555
LS131	Ade, P.P.	Feeding Efficiency Of Male And Female Poecilia Reticulata In Different Water Sample	558
LS132	Khadse, P.M. & Deshmukh, V.R.	Qualitative Phytochemical Analysis And Pharmacological Studies Of Acacia Arabica (Lamk.) Willd	566
LS133	Rewaskar, N.	Health, Food And Nutrition Of Adolescents	569
LS134	Sangole A.A. & Sangole M.T.	Monitoring Airborne Fungal Mycoflora In College Laboratory	571
LS135	Gupta, S.S.	Physico-Chemical Status Of Water Quality From Water Source At Drug Dam, Taluka Kalamb, District Yavatmal, Maharastra	574
LS136	Wanjari, H.V. & Somatkar J.R.	Diversity Of Avifauna In And Around Ekburji Reservoir, Washim M.S. India	577
LS137	Kadam V. & Deosthale S.	Screening Of Antidiabetic Ethnomedicinal Plants From Painganga Forest Range Umarkhed, District- Yavatmal, Maharashtra	586
LS138	Kalode, S. & Dalal, L.	Study Of Fungal Air Abundance, Diversity In Indoor Environment Of College Library	589
LS139	Tapase, B.S. & Nimbarte, S.R.	Chemi-Microbiological Characterisation Of Potable Water Of Nagpur City, Dist. Nagpur, Maharashtra.	593
LS140	Tirpude J.	Infrastructure Of The Amnion And Scanning Electron Microscope Appearances In The Megachiropteron Bat, Rousettusleschenaulti (Desmerest) At Term Pregnancy	596
LS141	Yeole, S.M.	Diversity Of Avifauna In And Around Parbhani Town, Maharashtra, India	601
LS142	Patil, P.S. & Tayade, S.N.	Lecanid Rotifers (Rotifera: Monogononta: Lecanidae) From Ditches Of Washim (MS)	605

STUDY OF FUNGAL SPORES FROM OUTDOOR ENVIRONMENT OF WARDHA CITY (M.S)

Kalode, S. & Dalal, L.

Department of Botany, J.B.College of Science, Wardha swatimkamdi@gmail.com

ABSTRACT

Air is the main source of fungi and they are abundantly present in the outdoor environment. Airborne fungi are the most common organisms in nature. Many physical, Chemical and biological factors are responsible for the growth of fungi. Fluctuation of temperature which gives variations with number of fungal spores. The present study was carried out in the year 2014-15 using gravity petriplate exposure method containing PDA media for trapping the fungal spores. The present study showed that number of fungal spores present in the vegetable market and studying their percentage contribution. The study shows a the percentage of Aspergillus spp and cladosporium spp was more dominant and other fungi like Penicillium ,Alternaria, fusarium Curvularia which are predominant fungi and Cunninghamella, Stachybotrytis spp was less no of counted.

Keywords: Aeromycoflora, Vegetable market, PDA medium

INTRODUCTION

Aeromycology is the study of dispersal and distribution of fungal spores and their mycelia in the air; and their dependence on meteorological conditions like temperature, rainfall, humidity, wind speed etc. The prevalence of airborne fungal spores are depended upon many biotic and abiotic factors, thus the airborne microorganism of any environment is specific in nature. Number and type of fungi vary with time of day, weather and seasonal fluctuations, condition of the surrounding areas, climatic conditions and the presence of a local source of spores (Pepeliniak and Segvic Klaric, 2003). Airborne microbe is a component of our environment and is of potential economic and health implications (Gregory, 1961; Molds Hashimoto, 1986). are frequent contaminant of fresh vegetables. Airborne fungi are also considered as a key factor and as indicator of the level of air pollution. Occurance of Aeromycoflora in vegetable and fruit market were studied. (Sharma also and Bhattacharjee,2001;Medhi and Sharma 2010).The aim of this study was to monitor the occurrences and seasonal distribution of the airborne cultivable fungi in the air of a Vegetable market associated environments at outdoor in order to evaluate.

MATERIALS AND METHODS

The Vegetable market was situated in the centre of wardha city and surrounded by railway station and Bus stand. For isolation of aeromycoflora, PDA culture media were used. Aeromycoflora of the Vegetable market of Wardha city was observed by gravity petriplate exposure method containing

PDA medium. This method also used by Tiwari P. (2008) for survey of aeromycoflora. The 10 cm diameter size Petriplate were exposed twice in a day. The exposed Petriplate were brought into the laboratory and incubated at $28 \pm 1^{\circ}$ C. for 4-5 days. At the end of incubation period the fungal colonies were counted, isolated and identified with the help of available literature (Barnett, 1969; Nigmani et al. 2006). For fungal spores identification, slides were prepared with the help of glycerine jelly as mounting media and lactophenol cotton blue as the standard stain. Temperature and Humidity were recorded in the Vegetable market during the sampling period using a Hygrometer. For ecological studies, percentage frequency and percentage contribution of individual species during the survey period was calculated using the standard formula (Tiwari, 1999)

Percentage contribution= <u>Total no of indivisual</u> <u>spp</u>

Total no of all spp \times 100

RESULT AND DISCUSSION

The present work deals with the Aeromycoflora of vegetable market from Wardha city. Kakde and Kakde (2012) was also shows air borne fungal spores in the vegetable market. In the present study total 831 fungal colonies belonging to 50 fungal forms, represent a group of a genus Ascomycotina ,Zygomycotina And Deuteromycotina. Most of the fungal colonies which represent the deuteromycotina. The Aspergillus and Cladosporium spp was more dominant fungi at the time of study period. The



UGC (CPE) and DST Sponsored International Conference on **Recent Trends in Science and Technology** S.S.S.K.R. Innani Mahavidyalaya, Karanja (Lad), Dist.Washim

(College with Potential for Excellence Reaccredited by NAAC at 'A' Level CGPA- 3.24) In collaboration with



Department of Mathematics, SGBAU, Amravati Vidya Bharati Mahavidyalaya, Amravati & Shri Dr. R.G. Rathod Arts & Science College, Murtizapur 22-23 March, 2018

CERTIFICATE



This is to certify that Prof./Dr./Mr./Ms. P.A. SAUDAGAR of JANKIDEVI BAJAJ COLLEGE OF SCIENCE, WARDHA

has actively participated in the International Conference on Recent Trends in Science & Technology held on 22nd & 23rd March, 2018 and has presented a research paper entitled ANTIBACTERIAL ACTIVITY OF COPPER (II) OXIDE NANOSTRUCTURE SYNTHESIZED BY MICROWARE IRRADIATION

during technical session of the conference.



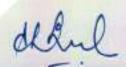












Dr. J.P. Baxi Organizing Secretary

Dr. D.T. Dongare Organizing Secretary

Dr. D.R. Halwe Organizing Secretary

Dr. A.P. Charjan

Dr. S.D. Katore Co-Convener

Dr. F.C. Raghuwanshi Co-Convener

Dr. P.R. Rajput Convener





UGC (CPE) and DST Sponsored International Conference on Recent Trends in Science and Technology



22-23 March, 2018

PROCEEDINGS



Organized by Vidya Bharati Shaikshanik Mandal Amravati's

S.S.S.K.R.Innani Mahavidyalaya Karanja (Lad), Dist. Washim (M.S.)

Reaccredited at level 'A' grade by NAAC (CGPA 3.24) & Conferred CPE status by UGC,New Delhi. In collaboration with

Department of Mathematics & IQAC Sant Gadge Baba Amravati University, Amravati Vidya Bharati Mahavidyalaya, Amravati & Shri Dr.R.G.Rathod Science College, Murtizapur

अनुक्रमणिका Humanities (मराठी विभाग)

अ. क्र.	लेखकाचे नाव	विषय	पृष्ठ क्र.
१	शा. बा. भोसले त.एस. राउत	अमरावती खिलाड़ी महिला कर्मचारिओं की जीवनषैली का हिमोग्लोबिन घटक के साथ सहसम्बंध	२६६२
२	रं. जिवने	मानवी शरीरावरावर ऑक्सिडनविरोधी द्रव्ये उपकारक तर मुक्तमुलके हानीकारक — एक अभ्यास	२६६५
ર	क.पी. कोरडे	आहार पोषणातील घटक आणि आजार, उपचार	२६६७
Х	व्हि. भि. कोल्हे	पर्यावरण संतुलनात मानवाचा सहभाग	२६७०
ų	एस.एम. लांबाडे	राष्ट्रसंत तुकडोजी महाराजांचा पर्यावरणीय दृष्टिकोन	२६७४
ह्	डी.एन. राठोड	भारतीय समाजातील अध्दश्रध्दा व वैज्ञानिक दृष्टीकोन	२६७९
9	अ. र. बोके	बालगृहातील किशोरवयीन मुले—मुलींच्या बौध्दिक गुणांकाचे तुलनात्मक अध्ययन	२६८४
۷	अ. सु. जोशी	कवी नीलकृष्ण देशपांडे यांच्या 'पापण्यांचे काठ ओले' कवितासंग्रहातील वेदना आणि सौंदर्य	२६८७
۶	अ. अ. नांदगावकर	आधुनिक युगातील मानवतावाद म्हणजेच शिक्षण विचार	२६८९
१०	शि. वै. भागवतकर	विज्ञान व तंत्रज्ञानाच्या युगात स्त्री शिक्षणात झालेले परिवर्तन एक अध्ययन	२६९२
११	सु. म. लाबडे	शाण्डिल्योपनिषद् प्रत्याहार का समायोजन, मानसिक स्वास्थ्य तथा बलात् जैवधारिता पर प्रभाव(महाविद्यालयीन छात्राओं के संदर्भ में)	२६९५
१२	प्रि. व. देशमुख साहित्यातून पर्यावरण दर्शन : कविता		२७००
१३	टि. डी. राजगुरे	इतिहास लेखनात विज्ञान व तंत्रज्ञानाचा उपयोग एक अभ्यास	२७०३
१४	वि. ढेंगळे	जागतीक तापमानवाढ आणि आंतरराष्ट्रीय राजकारण Global Warming & International Politics	२७०७
१५	रू. न. कऱ्हाडे	मराठी भाषा विषयाच्या अध्यापनात मल्टिमिडीया तंत्रज्ञानाचा व इतर नाविन्यपूर्ण साधनांचा उपयोग : एक संशोधनात्मक दृष्टिक्षेप	२७१०
१६	ज्ञा.धो. आघम	तरूणाईचे नवे विचारच देशाला नवी उंची गाठुन देतील	२७१४

ABSTRACTS SECTION

.

		Late Submission	
1	G.V. Lakhotiya, P.V. Tekade, P.A. Saudagar, N. G. Belsare, A.D. Rangari	Antibacterial Activity of copper (II) Oxide Nanostructure Synthesized by Microwave Irradiation	2777
2	S.S. Dhabekar & C. Patil	Ecofriendly Mosquito Repellent: Recent Trend in Environmental Sciences- A Review	2782
3	P.T. Agrawal	Synthesis and Characterization of Hepta-O-Benzoyl-	2787
4	G.D. Deshmukh	Waders Diversity in the Korambi Talav ofGhodazari Sanctuary, Maharashtra, India	2789

ANTIBACTERIAL ACTIVITY OF COPPER (II) OXIDE NANOSTRUCTURE SYNTHESIZED BY MICROWAVE IRRADIATION

G.V. Lakhotiya^{1*}, P.V. Tekade¹, P.A. Saudagar¹, N. G. Belsare², A.D. Rangari²

¹ Jankidevi Bajaj College of Science, Wardha- 442001, India ³ Brijlal Biyani Science College, Amravati- 444605, India *Corresponding author E-mail: <u>lakhotiya.govinda@gmail.com</u>

ABSTRACT

The multi-functionality of cupric oxide (CuO) nanostructure leads to fascinating applications in various fields. Herein, we report a simple, template-free, and surfactant-less wet-chemical microwave irradiated synthesis of CuO nanomaterials having ellipsoidal morphology. The morphology, structural, and optical properties of as-synthesized CuO nanostructures are investigated in detail. CuO nanoellipsoids are found to possess high surface area around $30m^2/g$ with the band gap of 1.85 eV and hence used to study the antibacterial activity. The antibacterial study was performed with gram positive and gram negative bacterial strains of Staphylococcus aureus and Escherichia coli, respectively.

Keywords: Nanoellipsoidal CuO, antibacterial activity, microwave synthesis, S. aureus, E. coli.

1. INTRODUCTION

CuO is one of the simplest members of the Cu family which shows its multi-functionality as a ptype semiconductor with narrow band gap of 1.24 eV.¹ It is a well-known material for the properties proliferating its applications in various fields viz. photovoltaics,² superconductors,³ lithium batteries,⁴ gas sensors,⁵ magnetic storage media,⁶ emission,⁷ methanol field synthesis.⁸ electrochemical sensing,⁹ water gas shift reaction,¹⁰ spin dynamics,¹¹ supercapacitors,¹² and heterogeneous catalysis.¹³ Besides the inherent multi-functional properties arising from heteronanomorphologies, CuO nanomaterials have also emerged out as a potential candidate for antibacterial and antimicrobial activity.¹⁴ The properties of CuO nanomaterials are closely related to its morphologies and crystallite size.^{15, 16} Various morphologies of CuO viz. micro-spheres,¹⁷ nanoplatelets,¹⁸ nanoparticles,¹⁹ nanorods,²⁰ nanowires,²¹ nanoneedles,²² nanoparticles,¹⁹ nanoneedles,²² nanoshuttles,²⁵ nanosheets,²⁴ nanoribbons,²³ nanobundles²⁶ etc. have attracted considerable attention due to their fundamental importance in above mentioned effective applications. Till date, various methods have been adopted to synthesize CuO, including solid-state reaction, sol-gel, electrochemical, sono-chemical, hydrothermal/solvothermal, microwave assisted hydrothermal, precipitation-pyrolysis, metal organic decomposition, and thermal decomposition.²⁷⁻³⁵ green Moreover, some biological routes have also been emerged.³⁶ All

these techniques are found to be efficient in producing single crystalline materials of diverse morphologies. However, most of these techniques either take long duration to carry out the reactions or not cost effective.

In addition to the need of short reaction duration, microwave synthesis also evenly suppresses side reactions, and thus enhances the yield with best reproducibility.³⁷ Microwave assisted hydrothermal technique has produced several hierarchical nanostructures of CuO including nanoflowers and nanopetals.^{32,38,39} However, there are not many reports on the microwave-irradiated synthesis of CuO using commercial microwave oven.⁴⁰⁻⁴³ Herein, we have adopted the same simple microwave-irradiated route for the wetchemical surfactant-less synthesis of copper oxide (CuO) nanostructures under ambient condition. The as-synthesized CuO nanostructures were characterized by X-ray diffraction (XRD), field emission scanning electron microscopy (FESEM), Brunauer-Emmett-Teller surface (BET) area analyzer, UV-vis. spectrophotometer for its structural, morphological, surface and properties. The as-synthesized nanostructure is found to possess good crystallinity, uniform morphology, and high purity and its antibacterial activity is tested against the gram positive bacterial strains of Staphylococcus aureus (NCIM 2127) and the gram bacterial strains of Escherichia coli negative (NCIM 2065) using the disc diffusion assay method.

2. EXPERIMENTAL 2.1 CHEMICALS

Copper sulphate (CuSO₄·5H₂O, 99.95%), sodium hydroxide (NaOH), ethanol (C₂H₅OH),.Bacterial strains of Staphylococcus aureus (NCIM 2127) and Escherichia coli (NCIM 2065) were obtained from National Chemical Laboratory (NCL), Pune, India. Muller–Hinton agar medium was used for the growth of microorganisms. All the chemicals received were of analytical grade and were used without any further purification.

2.2 SYNTHESIS OF CUO NANOMATERIAL

In the present study, microwave irradiated synthesis of CuO nanomaterial was carried out using the microwave irradiated technique as reported elsewhere with some modifications ⁴⁴. The commercial microwave chamber was used with the reaction conditions of 500 W for 10 min. During the reaction, color of the solution changed initially from blue to colorless and then slowly turns black. The black colored colloidal solution was centrifuged to separate out the precipitates. These precipitates were then washed using double distilled water, absolute ethanol, and acetone in sequence. This procedure was repeated several times. Finally, black colored powder was dried at 60° C for 4 h and used for further characterization.

2.3 CHARACTERIZATION

The crystallographic properties of the assynthesized samples were studied using PANalytical high resolution X-ray diffraction (PW 3040/60) operated at 40 kV and 30 mA using Cu Kα X-rays (1.54Å). The surface morphology of asprepared copper oxide nanostructures were analyzed using Carl Zeiss SUPRA 40 fieldemission scanning electron microscope (FESEM). The effective Brunauer-Emmett-Teller (BET) surface area of the as-synthesized nanomaterials was measured using a Ouantachrome ChemBET TPR/TPD analyzer. Optical property of the CuO nanostructures was studied using UV-vis. absorption spectrophotometer (Schimadzu 1800).

2.5 ANTIBACTERIAL ACTIVITY

Antibacterial activity of CuO nanostructure was tested against gram positive bacterial strains of Staphylococcus aureus (NCIM 2127) and gram negative bacterial strains of Escherichia coli (NCIM 2065) using the disc diffusion assay method with impregnated disks of as-synthesized CuO nanopetals. Approximately, 25.0 mL of molten and cooled nutrient agar media was poured in the sterilized petri dishes and was kept overnight at room temperature to ensure any contamination. The bacterial test organism S. aureus and E. Coli was grown in nutrient broth for 24 h at 37 °C. Bacterial lawns were prepared by using a 100 µL nutrient broth culture of each bacterial organism. Four dilutions of assynthesized CuO nanopetals was prepared for testing viz. 5 mg/mL, 2.5 mg/mL, 2 mg/mL and 1.5 mg/mL in ethanol. Nanostructured impregnated discs were then placed on the bacterial lawn. These plates were incubated at 37°C for 24 h.

3. RESULTS AND DISCUSSION 3.1 STRUCTURAL PROPERTY

X-ray diffraction (XRD) analysis was carried out to identify the crystal structure and phase purity of samples. Figure 1 illustrates the XRD pattern of as synthesized material. The material shows the diffraction peaks that match with JCPDS card no. 01-080-1916 indicating single monoclinic phase of CuO. The average crystallite size calculated by Scherrer formula is found to be \sim 12 nm.

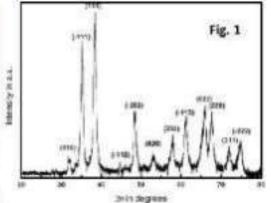


Figure 1. XRD pattern of as synthesized material 3.2 MORPHOLOGY

Figure 2 shows FESEM images of as synthesized CuO nanostructures which resemble ellipsoid-like morphology in nanodimension. There is a crucial role of concentration of NaOH to decide the preferential growth direction of nanostructure which supports the formation of ellipsoidal geometry of the nanostructure. ⁴⁵ Figure 3 depicts the energy dispersive absorption spectrum which confirms the stoichiometry of Cu:O as 1:1.

Aayushi International Interdisciplinary Research Journal (AIIRJ) ISSN 2349-638x Impact Factor 4.574 Special Issue No. 26 UGC Approved Sr.No.64259 Website :- www.aiirjournal.com Email id:- <u>aiirjpramod@gmail.com</u>

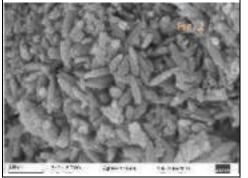


Figure 2. FESEM images of CuO nanostructures

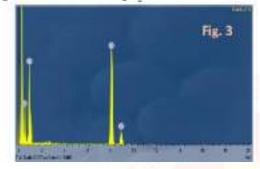


Figure 3. EDAX spectra of as synthesized nanostructure

3.3 SURFACE AREA

The Brunauer–Emmett–Teller (BET) technique was used to measure the surface area, pore radius, and pore volume of the as synthesized CuO nanostructure. Figure 3 depicts the nitrogen adsorption-desorption plots for the same. The effective specific surface area of the nanostructure was measured to be 30 m²/g. This larger surface area in this system could be well utilized to study the multi-functionality of CuO in particular the surface phenomenon viz. catalysis, antibacterial properties.

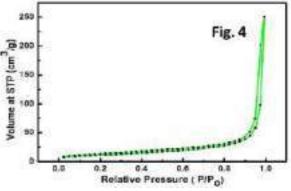


Figure 4. Nitrogen adsorption-desorption isotherm for CuO nanoellipsoids measured at 77 K

3.4 ANTIBACTERIAL ACTIVITY

Antibacterial activity of as synthesized CuO nanostructure has been studied by many groups relating to the role of different parameters viz. size effect, morphology, dissolution of copper ions in different medium etc.^{14, 46-48}. Thus, as-synthesized nanostructure of CuO has been treated for its antibacterial activity against both gram-positive and gram-negative bacteria. Figure 5 shows the image of zone of inhibition with different concentrations of CuO nanoellipsoids as an antibacterial agent against E. Coli and S. aureus.

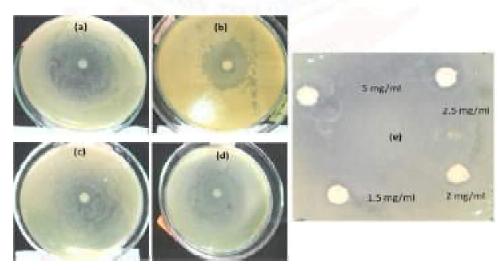


Figure 5. Zone of Inhibition of CuO nanopetals against E. Coli concentration (a) 1.5 mg/mL (b) 2 mg/mL (c) 2.5 mg/mL (d) 5 mg/mL (e) Zone of inhibition against S. aureus with different concentration of CuO nanostructure

Concentration of CuO	Zone of inhibition (in mm) against	
nanoellipsoids	E. Coli	S.
(mg/mL)		Aureus
5	25	18
2.5	23	15
2	20	12
1.5	18	06

Table 1. Zone of inhibition using CuO nanopetals against E.Coli and S.aureus

The diameter of inhibition zone reflects the magnitude of susceptibility of microbes (Table 1). E. Coli which exhibited a larger zone of inhibition than that of S. aureus is found to be more susceptible to CuO nanopetals. Table 1 also highlights the role of concentration of CuO nanopetals in the inhibition zone measurements and relates direct proportion between them. Our observations are in good agreement with Azam et al.⁵⁸ who also observed the same trend in case of CuO nanoparticles.

The metabolisms, differences in the cell structure, physiology or degree of contact of organisms with CuO nanomaterial are possible parameters that affect the antibacterial activity. The antibacterial activity of CuO nanoellipsoids can be attributed to the adhesion of bacterial cells over the surface of CuO nanoellipsoids, which releases the copper ions. These copper ions get attached with negatively charged bacterial cell wall leading to rupturing of the bacterial cell wall, thereby protein denaturation and ends with cell death. The effectiveness of CuO nanostructures in antibacterial activity is also known to be due to the

indirect effects or changes in the surrounding charge environment of bacteria.⁴⁸

4. CONCLUSIONS

In present work, we successfully synthesized CuO nanoellipsoids in absence of any surfactant and/or template using wet chemical microwave irradiation technique. The as-synthesized nanostructures were found to possess larger The as-synthesized surface area. CuO nanostructure is demonstrated for the antibacterial activity which gives excellent result against the bacteria E. coli and S. aureus.

ACKNOWLEDGMENTS

GL thanks Principal, Jankidevi Bajaj College of Science, Wardha (M.S.), India for providing facilities for carrying out this work and also acknowledges Director, Center for materials for electronics technology (C-MET), Pune for providing characterization facilities.

REFERENCES

- 1. S. Anandan and S. H. Yang, J Exp Nanosci, 2007, 2, 23-56.
- A. E. Rakhshani, Solid State Electron, 1986, 29, 7-17.
- 3. Y. Tokura, H. Takagi and S. Uchida, Nature, 1989, 337, 345-347.
- 4. T. Iijima, Y. Toyoguchi, J. Nishimura and H. Ogawa, J Power Sources, 1980, 5, 99-109.
- 5. X. P. Li, Y. Wang, Y. Lei and Z. Y. Gu, Rsc Adv, 2012, 2, 2302-2307.
- R. V. Kumar, Y. Diamant and A. Gedanken, Chem Mater, 2000, 12, 2301-2305.
- Y. W. Zhu, T. Yu, F. C. Cheong, X. J. Xui, C. T. Lim, V. B. C. Tan, J. T. L. Thong and C. H. Sow, Nanotechnology, 2005, 16, 88-92.

- 8. C. Baltes, S. Vukojevic and F. Schuth, J Catal, 2008, 258, 334-344.
- J. F. Ping, S. P. Ru, K. Fan, J. A. Wu and Y. B. Ying, Microchim Acta, 2010, 171, 117-123.
- E. Colbourn, R. A. Hadden, H. D. Vandervell, K. C. Waugh and G. Webb, J Catal, 1991, 130, 514-527.
- 11. F. Onufrieva and J. Rossatmignod, Phys Rev B, 1995, 52, 7572-7603.
- 12. S. K. Shinde, D. P. Dubal, G. S. Ghodake and V. J. Fulari, Rsc Adv, 2015, 5, 4443-4447.
- 13. A. Nezamzadeh-Ejhieh and S. Hushmandrad, Appl Catal a-Gen, 2010, 388, 149-159.
- 14. G. G. Ren, D. W. Hu, E. W. C. Cheng, M. A. Vargas-Reus, P. Reip and R. P. Allaker, Int J Antimicrob Ag, 2009, 33, 587-590.

- 15. G. M. Whitesides and B. Grzybowski, Science, 2002, 295, 2418-2421.
- 16. Y. Y. Xu, D. R. Chen and X. L. Jiao, J Phys Chem B, 2005, 109, 13561-13566.
- Y. Zhang, X. L. He, J. P. Li, H. G. Zhang and X. G. Gao, Sensor Actuat B-Chem, 2007, 128, 293-298.
- K. B. Zhou, R. P. Wang, B. Q. Xu and Y. D. Li, Nanotechnology, 2006, 17, 3939-3943.
- 19. R. K. Bedi and I. Singh, Acs Appl Mater Inter, 2010, 2, 1361-1368.
- C. Yang, X. T. Su, F. Xiao, J. K. Jian and J. D. Wang, Sensor Actuat B-Chem, 2011, 158, 299-303.
- 21. M. Kaur, K. P. Muthe, S. K. Despande, S. Choudhury, J. B. Singh, N. Verma, S. K. Gupta and J. V. Yakhmi, J Cryst Growth, 2006, 289, 670-675.
- 22. M. A. Dar, Y. S. Kim, W. B. Kim, J. M. Sohn and H. S. Shin, Appl Surf Sci, 2008, 254, 7477-7481.
- 23. C. L. Zhu, C. N. Chen, L. Y. Hao, Y. Hu and Z. Y. Chen, J Cryst Growth, 2004, 263, 473-479.
- 24. C. Yang, F. Xiao, J. D. Wang and X. T. Su, Sensor Actuat B-Chem, 2015, 207, 177-185.
- 25. D. Chen, G. Z. Shen, K. B. Tang and Y. T. Qian, J Cryst Growth, 2003, 254, 225-228.
- H. Chen, D. W. Shin, J. H. Lee, S. M. Park, K. W. Kwon and J. B. Yoo, J Nanosci Nanotechno, 2010, 10, 5121-5128.
- 27. W. Z. Wang, Y. J. Zhan, X. S. Wang, Y. K. Liu, C. L. Zheng and G. H. Wang, Mater Res Bull, 2002, 37, 1093-1100.
- H. W. Qin, Z. L. Zhang, X. Liu, Y. J. Zhang and J. F. Hu, J Magn Magn Mater, 2010, 322, 1994-1998.
- 29. G. Q. Yuan, H. F. Jiang, C. Lin and S. J. Liao, J Cryst Growth, 2007, 303, 400-406.
- 30. S. Anandan, G. J. Lee and J. J. Wu, Ultrason Sonochem, 2012, 19, 682-686.
- 31. Z. Y. Zhong, V. Ng, J. Z. Luo, S. P. Teh, J. Teo and A. Gedanken, Langmuir, 2007, 23, 5971-5977.

- 32. D. P. Volanti, M. O. Orlandi, J. Andres and E. Longo, Crystengcomm, 2010, 12, 1696-1699.
- 33. H. M. Fan, L. T. Yang, W. S. Hua, X. F. Wu, Z. Y. Wu, S. S. Xie and B. S. Zou, Nanotechnology, 2004, 15, 37-42.
- 34. A. Galembeck and O. L. Alves, Synthetic Met, 1999, 102, 1238-1239.
- 35. F. Bakhtiari and E. Darezereshki, Mater Lett, 2011, 65, 171-174.
- 36. A. V. Singh, R. Patil, A. Anand, P. Milani and W. N. Gade, Curr Nanosci, 2010, 6, 365-369.
- 37. I. Bilecka and M. Niederberger, Nanoscale, 2010, 2, 1358-1374.
- 38. J. Zhu and X. F. Qian, J Solid State Chem, 2010, 183, 1632-1639.
- 39. G. H. Qiu, S. Dharmarathna, Y. S. Zhang, N. Opembe, H. Huang and S. L. Suib, J Phys Chem C, 2012, 116, 468-477.
- 40. A. Jung, S. Cho, W. J. Cho and K. H. Lee, Korean J Chem Eng, 2012, 29, 243-248.
- 41. H. Wang, J. Z. Xu, J. J. Zhu and H. Y. Chen, J Cryst Growth, 2002, 244, 88-94.
- 42. H. C. Song, S. H. Park and Y. D. Huh, B Korean Chem Soc, 2007, 28, 477-480.
- 43. L. Xu, H.Y. Xu, F. Wang, F. J. Zhang, Z.D. Meng, W. Zhao and W.C. Oh, J Korean Ceramic Society, 2012, 49, 151-154.
- 44. G. Lakhotiya, S. Bajaj, A.K. Nayak, D Pradhan, P Tekade, A. Rana, Beilstein journal of nanotechnology 8, 1167.
- 45. Xu, L.; Xu, H.Y.; Wang, F.; Zhang, F.J.; Meng, Z.D.; Zhao, W.; Oh, W.C. Journal of the Korean Ceramic Society, **2012**, 49, 2,151-154
- 46. S. Meghana, P. Kabra, S. Chakraborty and N. Padmavathy, Rsc Adv, 2015, 5, 12293-12299.
- 47. A. Azam, A. S. Ahmed, M. Oves, M. S. Khan, S. S. Habib and A. Memic, Int J Nanomed, 2012, 7, 6003-6009.
- M. Ahamed, H. A. Alhadlaq, M. A. M. Khan, P. Karuppiah and N. A. Al-Dhabi, J Nanomater, 2014, 2014, 637858-637861.



Scanned by CamScanner

HOME

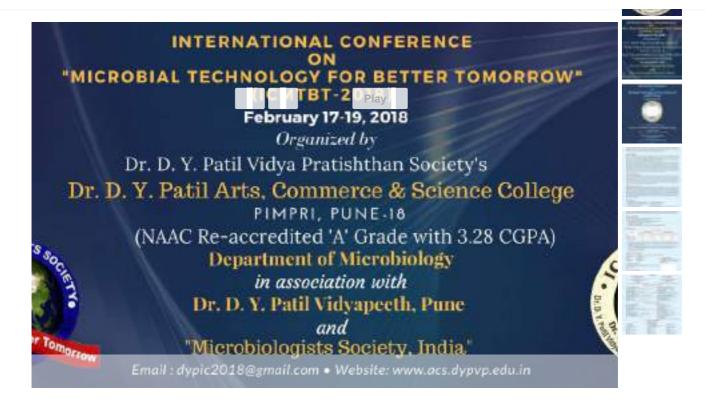
AIM AND SCOPE

\$

SPECIAL ISSUE 01

AUTHOR INSTRUCTIONS

MORE... LOG IN



Special Issue of 2018 International Conference on Microbial Technology for Better Tomorrow ICMTBT,

held @D.Y. Patil College of Arts, Commerce and Science, Pimpri, Pune,

17-19 February, 2018

Sl.No	Title and Author Name	
01	CHARACTERIZED IDENTIFICATION AND LOW-COST PRESERVATION OF MALASSEZIA sppENABLING FUTURE POSSIBILITY FOR CONTROL Komal S. Gomare and D.N. Mishra	01-10
02	<u>EFFICACY OF SEQUENTIAL SPRAYS OF DIFFERENT FUNGICIDES</u> <u>AGAINST EARLY BLIGHT ALTERNARIA SOLANI (ELLIS AND MARTIN)</u>	11-15

HOME

AIM AND SCOPE

SPECIAL ISSUE 01

AUTHOR INSTRUCTIONS

MORE... LOG IN

Ļ

	Alisha D. Malik and Irene J. Furtado	
05	MICROBIAL QUALITY ASSESSMENT OF SOME BRANDS OF COSMETICS PRODUCTS SOLD IN INDIAN MARKETS M F Mansuri, C D Afuwale, M A Upadhyay, A N Chopda and N V Patel	29-36
06	STUDIES ON SEMEN PRESERVATION AND EXPRESSION OF HSPA8 IN ENDANGERED INDIAN LEOPARD (PANTHERA PARDUS FUSCA) SPERMATOZOA BY USING QUANTITATIVE PCR (Q-PCR) PGS Manjusha and Sadanand D. Sontakke	37-43
07	ESTIMATION OF ANTIOXIDANT AND PHENOLIC CONTENT OF ORGANICALLY AND CONVENTIONALLY GROWN TOMATO CULTIVAR (Solan Lalima) UNDER MID-HILL CONDITIONS OF HIMACHAL PRADESH Nitika Thakur	44-52
08	TRANSFORMATION OF RHIZOBIUM WITH PLASMID FROM PSEUDOMONAS SPP. A 1113 TO DEGRADE DIMETHOATE S.R Shinde, M.V. Bhailume and V.S Hamde	53-56
09	PARTIAL CHARACTERIZATION AND PURIFICATION OF A NOVEL CLASS II A PEDIOCIN PRODUCED BY PEDIOCOCCUS PARVULUS STRAIN MF233 ISOLATED FROM BATTER OF IDLI A TRADITIONAL FERMENTED FOOD OF SOUTH INDIA FOR MANAGEMENT OF FOOD BORNE PATHOGENS S. S. Khandare and S.D Patil	57-65
10	BIOREMEDIATION OF TEXTILE AZO DYE ORANGE F2R BY BACTERIAL ISOLATES Amarja Harishchandra Bhosale and R. M. Khobragade	66-69
11	PRODUCTION OF PENICILLIN ACYLASE BY BACILLUS SUBTILIS Afshaan Naaz Khaleed Shaikh*, Mankale K.K and Dixit P.P.	70-74



ł

International Journal of Pharmacy and Biological Sciences

ISSN: 2321-3272 (Print), ISSN: 2230-7605 (Online) IJPBS | Volume 8 | Special Issue 1- ICMTBT | 2018 | 57-65 International Conference on Microbial Technology for Better Tomorrow ICMTBT Held at D.Y. Patil College of Arts, Commerce and Science, Pimpri, Pune, 17-19 February





| Conference Proceedings | Research Article | Biological Sciences |Open Access |MCI Approved|

UGC Approved Journal

PARTIAL CHARACTERIZATION AND PURIFICATION OF A NOVEL CLASS II A PEDIOCIN PRODUCED BY *PEDIOCOCCUS PARVULUS* STRAIN MF233 **ISOLATED FROM BATTER OF IDLI A TRADITIONAL FERMENTED FOOD OF SOUTH INDIA FOR MANAGEMENT OF FOOD BORNE PATHOGENS**

S. S. Khandare^{*1}, and S.D Patil²

¹ Dept. Of Microbiology, J. B. College of Science, Wardha, 442001, Maharashtra, (INDIA), ² Former Head, Dept. Of Microbiology and Biotechnology, Shri Shivaji Science College, Amravati 44603, Maharashtra, (INDIA)

*Corresponding Author Email: ksuhas21@gmail.com

ABSTRACT

Pediococcus parvulus MF 233 isolated from batter of idli a traditional fermented food of south India, identified based on its 16S rRNA gene sequence, produced pediocin that had broad spectrum of inhibition against Gram positive and Gram negative antibiotic resistant pathogenic, food spoilage organisms, Salmonella typhi, Escherichia coli., Pseudomonas aeruginosa and Staphylococcus aureus. Pediocin (designated as pediocin MF233) was purified by ammonium sulphate precipitation, dialysis (MWCO 1000D) and ion exchange chromatography (DEAE Cellulose) showed increased specific activity from 3.66 to 9.52, 27.50 and 216, with 2.6, 2.88 and 7.87-fold increase in purification of protein respectively. The molecular weight of pediocin MF233 was 3000 Da. Pediocin MF233 found to be most heat stable and showed full activity at 121 $^{\circ}$ C for 10 minutes and maintained full stability after storage for 60 days at -20 °C and for 40 days at 4°C, partial stability at 37°C for 20 and 40 days while stability decreased slowly thereafter and lossed completely by 60 days. Pediocin MF233 showed stability at pH 2 to 6. Active principle of Pediocin MF233 was proteinaceous in nature since it was inactivated by proteolytic enzymes, but not by nonproteolytic enzymes. UV radiation did not affect the activity of Pediocin MF233. The studies concluded that the ability of Pediocin MF233 produced by Pediococcus parvulus MF233 in inhibiting a wide-range food pathogenic and spoilage bacterium, is of potential interest for food safety and may have future applications as food preservative.

KEY WORDS

Pediocin MF233, partial purification, characterization, foodborne pathogens, inhibitory activity.

INTRODUCTION:

The empirical use of microorganisms and their natural products for the preservation of foods (biopreservation) have been a common practice in the history of mankind.¹ The lactic acid bacteria generally considered as food grade organisms since they are involved in numerous food fermentations, known to man for

millennia, do not pose any health risk to man and are designated as generally regarded as safe (GRAS) status.² The inhibition of food spoilage microbes could be attributed to the production of antimicrobial compounds including organic acids, hydrogen peroxide, antibiotics and bacteriocins.³ Many species of Lactobacillus, used in the manufacture of fermented dairy products, inhibit the growth of other bacteria

Substituted diethyl 4-(substituted-phenyl)-2,6-dimethyl-1,4-dihydropyridine-3,5 dicarboxylate found to be effective as active biological compound. The physico-chemical properties of the transition metal complexes of these newly synthesized N-heterocyclics with certain metals are reported here. The proton-ligand and metal-ligand stability constants of complexes of diethyl 4-(substituted-phenyl)-2,6-dimethyl-1,4-dihydropyridine-3,5dicarboxylate with Ni(II), Cu(II), and Co(II) were determined in mixed solvents (70% dioxane). Presence of OH/NH group(s) in these compounds confirmed at 0.1 M ionic strength in 70% dioxane-water mixture pH metrically Ni(II), Cu(II), and Co(II) metal ions formed 1: 1 and 1: 2 complex with all the three ligands. The result shows that the ratio of logK1/logK2 is positive in all cases. This indicates that there is little or no steric hindrance to the addition of secondary molecules.



Sonal D. Bajaj Pradip V. Tekade

Dr. PRADIP V, TEKADE , M.Sc., Ph.D., NET, SET, GATE, Associate Professor, Department of Chemistry, Jankidevi Bajaj College of Science, Wardha He has published 50 research papers in Journals and 6 Books.Dr. SONAL D. BAJAJ , M.Sc., Ph.D., B.Ed. She has published 14 research papers in various national and international Journals and two books.

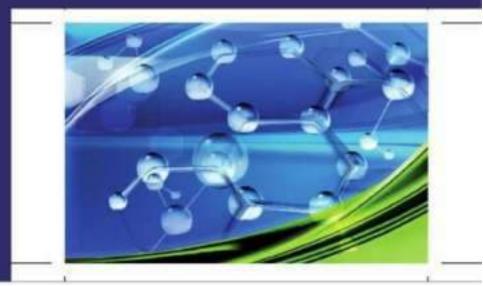
Proton/Metal– Ligand Stability Constants of Complexes of N-Heterocycle

pH-Metric Study of Stability Constant of First Transition Metal Ions with Novel Bio-inspired 1,4-Dihydropyridine Ligands





The ultrasound welocity measurements are helpful to study the intermolecular interactions and thermodynamical properties of pure components and their mixtures. Therefore, in the present book, ultrasonic studies of substituted propenamide studied in ethanolic solutions of various concentrations at different temperatures 303, 308 and 313 K with a view to understand molecular interactions in these solutions. Moreover, the complex formation in the solution of some N-heterocyclic compounds studied with some metal ions by determination of stability constant using spectrophotometry. The spectrophotometric study of chelating properties of some newly substituted heterocycles viz 2:44,5-dihydro-1,2-oxazol-5 yllphenol-N-methylaniline, 4-(1H-benzimdazole-2-yllphenol and Ethyl 4-44-hydroxyphenyl)-6-methyl-2-oxo-1,2,3,4-tetrahydropyramidine-5-carboxylate with Ni (II) and Cu(II) metal ions by Job's variation method reported here. The study of thermo-acoustical parameters and study of stability constant of complexes by spectrophotometric study gives an important information regarding the presence of molecular interaction. It has an application in drug absorption and transmission.



Sonal Dilip Bajaj Pradip V. Tekade

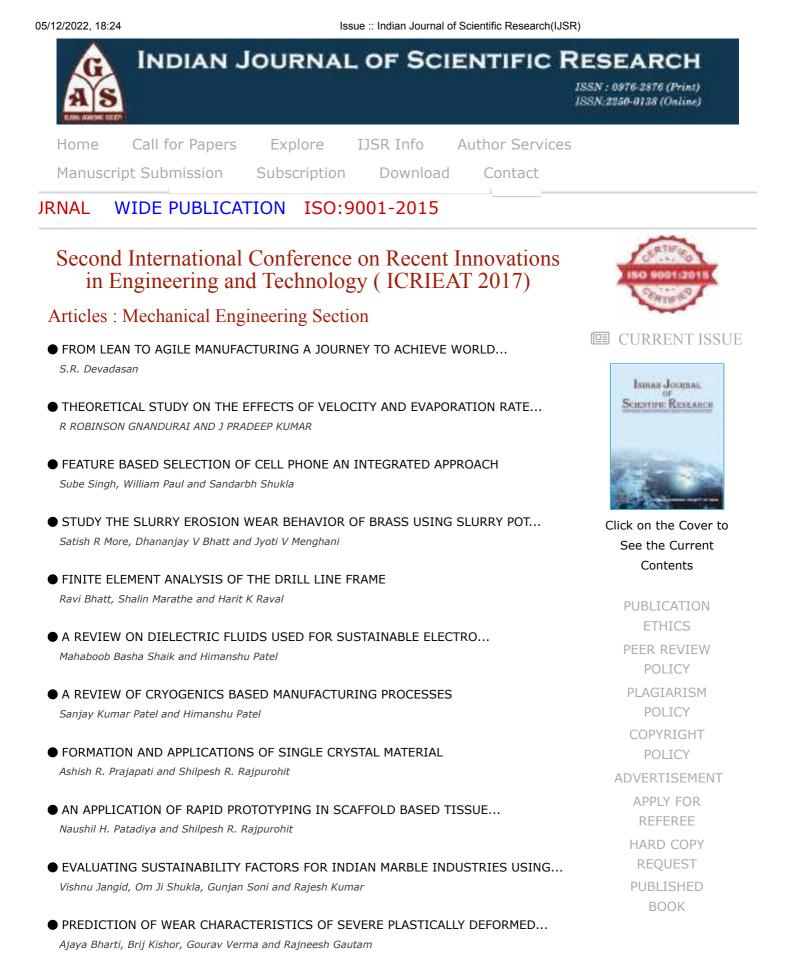
Dr. Pradip V. Tekade : M.Sc., Ph.D., NET , SET , GATE. Assistant Professor in Deptt. of Chemistry, J.B.C.S. Wardha. He has published 45 research papers in journals and 6 books. Dr. Sonal D. Bajaj; M.Sc., Ph.D., 8.Ed. Assistant Professor in Deptt. of Chemistry, J.B.C.S. Wardha. She has published 13 research papers in journals and one book.

Acoustical and Spectrophotometric Study of Substituted N-Heterocyclics

Determination of Specific Molecular Interaction and Stability Constant of Substituted N-Heterocyclics







- FREE VIBRATION ANALYSIS AND WEIGHT OPTIMIZATION OF COMPOSITE DRIVE... Anamika and Anindya Bhar
- APPLICATION OF BOX-BEHNKEN DESIGN FOR OPTIMIZATION OFDRILLING... Ajith Arul Daniel S, Sakthivel M and Sudhagar S

- ENHANCING THE SECURE FILE-LEVEL AND BLOCK-LEVEL AUTHORIZED DATA... Pingili Sravya
- IMPLICIT ASCERTAIN FOR PLAGIARISM DETECTION AND TEXT CLASSIFICATION Nilesh Channawar and S.B. Kishor
- HELPING HANDS ANTHROPOMORPHIC ROBOT HAND S. Kanagaraj, G. Prema Arokia Mary and M. S. Hema
- DETECTING FORGED IMAGES USING EDBTC FEATURE
 P.C. Chandrika, Y. Suganya and R. Arthie Reena
- UNIVERSAL PLATFORM FOR HETEROGENEOUS MOBILE APPLICATIONS TO... R. Kalaiselvi, V. P. Sumathi and M. S. Hema
- CROP HAND-AN ANDROID BASED CROP AND FERTILIZER ADVISOR G. Sundrameenakshi, A. Jayasuriya, G. Srioviya and V.P. Sumathi
- A STRUCTURE FOR ANALYZING AND DETRACTING NEGATIVE EMOTIONAL... A. Lokesh, M.S. Hema and Nageswara Guptha
- A STUDY OF FEATURE SELECTION IN COMPRESSED MEDICAL IMAGES USING... Shaik Jakeer Hussain and R. Kiran Kumar
- A SURVEY ON RELEVANT FEATURE EXTRACTION USING TEXT MINING IN DM... K. Kathirvel and C. Ramachandran
- A SURVEY ON INTERNET OF AGRICULTURE M.C.S. Geetha, M. Amala Jayanthi, R. Lakshmana Kumar and I. Elizabeth Shanthi
- REVIEW OF VARIOUS TECHNIQUES IN BRAIN TUMOUR SEGMENTATION USING MRI Jayashree Shedbalkar, K. Prabhushetty and Abhay Kumar Inchal
- AN EFFECTIVE IMAGE RETRIEVAL USING GENETIC ALGORITHM BASED ON... Aslam Y. Suriya, Sumedh J. Manwatkar and Varsha Choudhary
- SECURITY ASPECTS OF REST WEB SERVICES Amar Pimpalkar and Manish Jivtode
- PREDICTION OF SOIL TEXTURE USING FEED FORWARD NEURAL NETWORKS
 P. Parameswari and M. Manikantan
- A COMPARATIVE STUDY ON PERFORMANCE ANALYSIS OF CRYPTOGRAPHIC C. Sruthi, T. Abirami and G. Prema Arokia Mary
- NOISELESS DATA COMPRESSION TECHNIQUE BY USING BURROWS-WHEELER... Karuna Khobragade, Shruti Malame and Pratiksha Kapse
- SENTICOMPOSITE COMPLAINT PORTAL USING SUPERVISED APPROACH M. Suryia, P. Valarmathi and Radhika Devi
- DATAANALYSIS PROCEDURE FOR DISCOVERING RELATIONSHIPS IN A GRAPH WITH...

Chandrasekaran K. Arun, Vilas M. Ghodki and S.B. Kishor

0-8.03.86647.85.6

DATAANALYSIS PROCEDURE FOR DISCOVERING RELATIONSHIPS IN A GRAPH WITH RESPECT TO NETWORKING

¹Chandrasekaran K Arun, ²Dr, Vilas M Ghodki, ⁴Dr.S.B.Kishor ¹Research Scholar, Gondwana University, Mahaharashtra, India ²Associate Professor, Dept. of Computer Science, J.B. College, Wardha, Maharashtra, India ³Head, Dept. of Computer Studies and Research. S.P College, Chandrapur, Maharsashtra, India E-Mail :¹bmk_17@yahoo.com, ³vilasghodki@gmail.com,⁷s.b.kishor.spc@gmail.com

Abstract - This paper precisely proposes a link-analysis based technique allowing to discover relationships existing between nodes in a computer network or, more generally, a graph. More specifically, this work is based on a random-walk through the database defining a Markov chain having as many states as nodes in the computer network. Suppose, for instance, we are interested in analyzing the relationships between nodes in a computer network, a two-step procedure is developed in analyzing the relationships. First, a much smaller, reduced. Markov chain, only containing the nodes but preserves the main characteristics of the initial chain, is extracted by stochastic complementation. For extracting the reduced Markov by stochastic complementation, an efficient algorithm is proposed. Secondly, the reduced chain is analyzed by, for instance, projecting the states in the subspace spanned by the right eigenvectors of the transition matrix called the basic diffusion map, or by computing a kernel principal-component analysis on a diffusion-map distance, extending the basic diffusion map to directed graphs, is introduced.

Keywords - Diffusion Map , Stochastic complementation, Feature Redundancy

I. INTRODUCTION

Wireless sensor networks (WSNs) are being used for diverse applications such as low cost area monitoring. environment monitoring, industrial and machine health monitoring, structural monitoring and military surveillance [1], [2]. In these applications, WSNs generate a large amount of data in the form of streams. In recent times, data mining techniques have been used to extract useful knowledge from WSN data [3], through discovering relationships among the sensor nodes which are known as behavioral patterns [4]. More recently, research has been focused to mine different types of behavioral patterns, e.g., sensor association rules [5], [6], [9] from stored (static) sensor data, context association rules [10] from sensor data stream, associated sensor patterns [7] and regularly frequent sensor patterns [8] from static as well as stream data. Traditional statistical, machine learning, pattern recognition, and data mining approaches [28] usually assume a random sample of independent objects from a single relation. Many of these techniques have gone through the extraction of knowledge from data, almost always leading, in the end, to the classical double-entry tabular format, containing features for a sample of the

population. These features are therefore used in order to learn from the sample, provided that it is representative of the population as a whole. However, real-world data coming from many fields such as World Wide Web, marketing, social networks, or biology [16] are often multi relational and interrelated. The work recently performed in statistical relational learning [22], aiming at working with such data sets, incorporates research topics, such as link analysis [36] web mining [1].[9], social network analysis [8], or graph mining[11]. All these research fields intend to find and exploit links between objects which could be of various types and involved in different kinds of relationships. On the other hand, when dealing with a starschema database , this two-step procedure reduces to The multiple correspondence analysis. proposed methodology therefore extends correspondence analysis to the analysis of a relational database. In short, this paper has three main contributions:A two-step procedure for analyzing weighted graphs or relational databases is proposed ...

 It is shown that the suggested procedure extends correspondence analysis. ISBN: 978-93-86647-85-6

 A kernel version of the diffusion map distance, applicable to directed graphs, is introduced.

The paper is organized as follows: the basic diffusion map distance and its natural kernel on a graph in Section II. In Section IV we some experimental results involving several data sets.

ILTHE DIFFUSION MAP DISTANCE AND ITSNATURAL KERNEL MATRIX

In this section, the basic diffusion map distance [24] is briefly reviewed and some of its theoreticaljustifications are detailed. Then, a natural kernel matrix isderived from the diffusion map distance, providing ameaningful similarity measure between nodes.

A. The Diffusion Map Distance

In our two-step procedure, a diffusion map projection, based on the so-called diffusion map distance, will beperformed after stochastic complementation. Now, since the original definition of the diffusion map distance dealsonly with undirected, aperiodic, Markov chains, it will first be assumed in Section 2 that the reduced Markov chain, obtained after stochastic complementation, is indeed undirected, aperiodic, and connected—in which case the corresponding random walk defines an irreducible reversible Markov chain. Notice, that it is not required that the

original adjacency matrix is irreducible and reversible; theseassumptions are only required for the reduced after obtained adjacencymatrix complementation. The original derivation of the diffusion map, introducedindependently by Nadler et al., and Pons and Latapy [22],[13], is detailed in interpretations of this mapping appeared in the literature For an application of the basicdiffusion map to dimensionality reduction, see [35].Since P is aperiodic, irreducible, and reversible, it is wellknown that all the eigenvalues of P are real and theeigenvectors are also real [7]. Moreover, allits eigenvalues $\in [-1, +1]$, and the eigenvalue 1 has multiplicityone [7]. With these assumptions, Nadler et al. andPons and Latapy [42], [43], [46]. [47] proposed to use asdistance between states i and j

$$d_{ij}^{2}(t) = \sum_{k=1}^{n} \frac{(x_{ik}(t) - x_{jk}(t))^{2}}{\pi_{k}}$$
(1)

since, for a simple random walk on an undirected graph, the entries of the steady-state vector $\boldsymbol{\pi}$ are proportional (the

 \propto sign) to the generalized degree of each node (the total of the elements of the corresponding row of the adjacencymatrix [28]). This distance, called the diffusion mapdistance, corresponds to the sum of the squared differencesbetween the probability distribution of being in any stateafter t transitions when starting (i.e., at time t = 0) from two different states, state i and state j. In other words, two nodes are similar when they diffuse through the network—and thus influence the network—in a similar way. This is anatural definition which quantifies the similarity between two states based on the evolution of the states' probability distribution. Of course, when $i = j, d_{xj}(t) = 0$ Nadler et al. [22] showed that this distance measure a simple expression in terms of the right eigenvectors of P:

$$d_{ij}^{2}(t) = \sum_{k=1}^{n} \lambda_{k}^{2t} (u_{ki} - u_{kj})^{2},$$
 (3)

where $u_{ki} = [\mathbf{u}_k]_i$ is component *i* of the *k*th right eigenvector, \mathbf{u}_k , of P and λ_k is its corresponding eigenvalue. Assusual, the λ_k are ordered by decreasing modulus, so that the contributions to the sum in (3) are decreasing with *k*. On the other hand, $\mathbf{x}_i(t)$ can easily be expressed in the space spanned by the left eigenvectors of P, the ∇k .

$$\mathbf{x}_{i}(t) = (\mathbf{P}^{T})^{t} \mathbf{e}_{i} = \sum_{k=1}^{n} \lambda_{k}^{t} \mathbf{v}_{k} \mathbf{u}_{k}^{T} \mathbf{e}_{i} = \sum_{k=1}^{n} (\lambda_{k}^{t} u_{k}) \mathbf{v}_{k}, \quad (4)$$

where ei is the ith column of I,

e, = $[0, ..., 0, 1, 0, ..., 0]^T$ with the single 1 in position *i*. The resulting mapping aims to represent each state *i* in andimensional euclidean space with coordinates $(1\lambda_i^t)u_{0i}(\lambda_i^t)u_{0i}(\dots, |\lambda_n^t|u_{ni})$ as in (4). Dimensions

$$|\lambda_2^t| u_{2i}, |\lambda_3^t| u_{3i}, \dots, |\gamma_n| = u^t$$
, as in (4). Dimensions $|\lambda_k^t|$. This original

areordered by decreasing modulus, $[^{-k_1}]$. This original mappingintroduced by Nadler and coauthors will be referred to asthe basic diffusion map in this paper, in contrast with the diffusion map kernel (KDM) that was introduced in Section II. The weighting factor, D^{-1} , in (2) is necessary to obtain(3), since the Vk are not orthogonal. Instead, it can easily be shown that we have ISBN 978-93-38647-85-6

P1

product as
$$(x, y) = x^T D^{-1} y$$
, where the metric of the space is D^{-1} [7] Notice also that there is a close relationship between the space is $(x, y) = x^T D^{-1} y$.

relationship betweenspectral clustering (the mapping provided by the normalizedLaplacian matrix; see, for instance, [15], [35]) andthe basic diffusion map. Indeed, a common embedding of he nodes consists of representing each node by the coordinates of the smallest nontrivial eigenvectors (correspondingto the smallest eigenvalues) of the normalizedLaplacian matrix, $\tilde{\mathbf{L}} = \mathbf{D}^{-1/2} \tilde{\mathbf{L}} \mathbf{D}^{-1/2}$,

More precisely, if \mathbf{u}_k isthe kth largest right eigenvector of the transition matrix Pand L is the kth smallest nontrivial eigenvector of thenormalized Laplacian matrix

A subtle, still important, difference between this mappingand the one provided by the basic diffusion map concerns theorder in which the dimensions are sorted, Indeed, for the basic iffusion map, the eigenvalues of the transition matrix P areordered by decreasing modulus value. For this spectralclustering model, the eigenvalues are sorted by decreasingvalue (and not modulus), which can result in a differentrepresentation if P has large negative eigenvalues. Thisshows that the mappings. provided by spectral clustering andby the basic diffusion map are closely related. Notice that at least three other justifications of this eigenvector-based mapping appeared before in the literature, and are briefly reviewed here. It has been shownthat the entries of the subdominant right eigenvector of thetransition matrix P of an aperiodic, irreducible, reversible, Markov chain can be interpreted as a relative distance to its"stationary distribution" . This distance may be regarded as an indicatorof the number of iterations required to reach this equilibriumposition, if the system starts in the state from whichthe distance is being measured. These quantities are onlyrelative, but they serve as a means of comparison among thestates [30]. The same embedding can be obtained byminimizing the criterion

$$\sum_{i=1}^{n} \sum_{j=1}^{n} a_{ij} (z_i - z_j)^2 = \mathbf{z}^{\mathrm{T}} \mathbf{L} \mathbf{z}$$

Here, z_i is the coordinate of node i on the axis and the vector z contains the z_i . Theproblem sums up in finding the smallest nontrivial eigenvector of (I-P), which is the same as the secondlargest eigenvector of P, and this is once more similar to thebasic diffusion map. Notice that this mapping has beenrediscovered and reinterpreted by Belkin and Niroyi [2], [3]in the context of nonlinear

dimensionality reduction. Thelast justification of the basic diffusion map, introduced in[15], is based on the concept of two-way partitioning of agraph. Minimizing a normalized cut enterion whileimposing that the membership vector is centered withrespect to the metric D leads to exactly the same embeddingas in the previous interpretation. Moreover, some authors

showed that applying a specific cut criteria to bipartitegraphs leads to simple correspondence analysis. More generally, these mappings are, of course, alsorelated nonlinear and embedding graph 10 dimensionalityreduction, which have been highly studied topics in recentyears, especially in the manifold learning community (see,i.e., [21], [30], [37] for recent surveys or developments).Experimental comparisons with popular dimensionalityreduction techniques are nonlinear presented in the following section.

IV. EXPERIMENT AND ANALAYSIS

A. Graph Reduction Influence and Embedding

Comparison

The objective of this experiment is twofold. The first aim isto study the influence of stochastic complementation ongraph mapping. The second one is to compare five populardimensionality reduction methods, namely, the diffusionmap kernel PCA (KDM PCA or simply KDM), the LaplacianEigenmap (LE) [3], the Curvilinear Component Analysis(CCA) [14], Sammon's nonlinear Mapping (SM) [25], andthe classical Multidimensional Scaling [6], [12], based ongeodesic distances (MDS). For CCA, SM, and MDS, thedistance matrix is given by the shortest path distancecomputed on the reduced graph whose weights are set tothe inverse of the entries of the adjacency matrix obtainedby stochastic complementation. Notice that the MDSmethod computed from the geodesic distance on a graphis also known as the ISOMAP method after [6]. Providedthat the resulting reduced Markov chain is usually dense, the time complexity of each algorithm is as follows: ForKDM PCA, LE, and MDS, the problem is to compute the dominant eigenvectors of a square matrix since the graphis mapped on a d-dimensional space, which is

 $O(d \tau n_1^2)$, where n is the number of nodes of interest

being displayed and au is the number of iterations of the power method. ForSM and CCA, the complexity is about $O(\tau n_1^2)$, where τ is the number of iterations (these 1.6

algorithms are iterative byrecorded. On the other hand, computing the shortest pathdistances matrix takes $O(\alpha_1^2 \log(\alpha_1))$ Thus, each algorithmhas a time complexity between $O(n_1^2)$ and $O(n_1^3)$. In this experiment, we address the task of classification of unlabeled nodes in labeloi graphs. that is semisupervised classification on a graph . Notice thatthe goal of this experiment is not to design a state-of-theartsemisupervised classifier, rather it is to study theperformance of the proposed method, in comparison withother embedding methods Three graphs are investigated. The first graph isconstructed from the well-known Iris data set [4]. Theweight (affinity) between nodes representing samples isprovided by

$$w_{ij} = exp[-d_{ij}^2/\sigma^2]$$
, where d_0 is the cuclidean

distance in the feature space and of is simply the samplevariance. The classes are the three iris species. The secondgraph is extracted from the IMDb movie database [37] The last graph, extracted from the CORA data set , is composed ofscientific papers from three topics. A citation graph is builtupon the data set, where two papers are linked if the firstpaper cites the second one. The tested graph contains1,410 nodes divided into three classes representing machinelearning research topics For each of these three graphs, extra nodes are added torepresent the class labels (called the class nodes). Each classnode is connected to the graph nodes of the correspondingelass. Moreover, in order to define cross-validation folds, these graph nodes are randomly split into training sets andtest sets (called the training nodes and the test nodes, respectively), the edges between the test nodes and the classnodes being removed. The graph is then reduced to the testnodes and to the class nodes by stochastic complementation(the training nodes are rejected in the S2 subset, and thus, censored), and projected into a 2D space by applying one of he projection algorithms described before. Terms and topic nodes are displayed jointly.

between the test nodes and the class nodes is accuratelyreconstructed in the reduced graph, these nodes from the testset should be projected close to the class node of their corresponding class. We report the classification accuracy forseveral labeling rates, i.e., portions of unlabeled nodes which constitute the test set. The proportion of the test nodes varies between 50 percent of the graph nodes (rwofold crossvalidation) to 10 percent (10-fold cross validation). This means that the proportion

of training nodes left apart(censored) by stochastic complementation increases withthe number of folds. The whole cross-validation procedure isrepeated 10 times (10 runs) and the classification accuracyaveraged on these 10 runs is reported, as well as the95 percent confidence interval.For classification, the assigned label of each test node issimply the label provided by the nearest class node, in termsof cuclidean distance in the 2D embedding space. This willpernnt to assess if the class information is correctly preservedduring stochastic complementation and 2D dimensionalityreduction. The parameter t of theKDM PCA is set to 5, in viewof our preliminary experiments.Figs. 1a, 1b, and 1c show the classification accuracy, aswell as the 95 percent confidence interval, obtained on thethree investigated graphs for different training test setpartitioning (folds). The x-axis represents the number offolds, and thus, an increasing number of nodes left apart(censored) by stochastic complementation (from 0, 50, . . . , upto 90 percent). As a baseline, the whole original graphcorresponding to one single fold and referred to as 1-fold) isalso projected without removing any class link and withoutperforming a stochastic complementation; this situation represents the ideal case, since all the class information iskept. All the methods should obtain a good accuracy score inthis setting-this is indeed what is observed First, we observe that, although obtaining very goodperformance when projecting the original graph (1-fold), CCA and SM perform poorly when the number of folds, and thus, the amount of censored nodes, increases. On theother hand, LE is quite unstable, performing poorly on theCORA data set. This means that stochastic complementationcombined with CCA, SM, or LE does not workproperly. On the contrary, the performance of KDM PCAand MDS remains fairly stable; for instance, the averagedecrease of performance of KDM PCA is around 10 percent, in comparison with the mapping of the original graph(from 1-fold to 2-fold-50 percent of the nodes arecensored), which remains reasonable. MDS offers a goodalternative to KDM PCA, showing competitive performance;however, it involves the computation of the all-pairsshortest path distance. These results are confirmed when displaying the mappings.Figs. Ia, 1b, and 1c show a mapping example offhe test nodes, as well as the class nodes (the white markers)of the CORA graph, for the 10-fold cross-validation setting. Thus, only 10 percent of the graph nodes are unlabeled andprojected 90 the stochastic complementation of after percentremaining nodes. It can be observed that the LaplacianEigenmap managed toseparate the different

ISBN: 978-93-86647-85-6

classes, but mostly in terms of angularsimilarity. On the KDM PCA mapping (Fig. 8d), the classnodes are welllocated, at the center of the set of nodeshelonging to the class. On the other hand, the mappingsprovided by CCA and SM after stochastic complementationdo not accurately preserve the class information.

Figure 1(a): Classification accuracy obtained by the five compared projection methods for the Iris ((a), three classes), IMDb

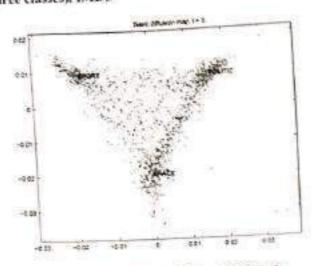


Figure 1 (b) : KDM PCA, or KDM), the LaplacianEigenmap ((e), LE), the Curvilinear **Component Analysis**

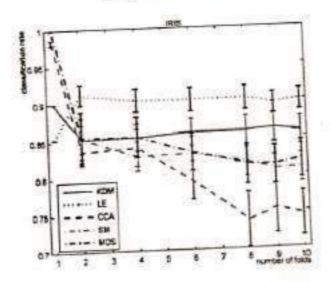
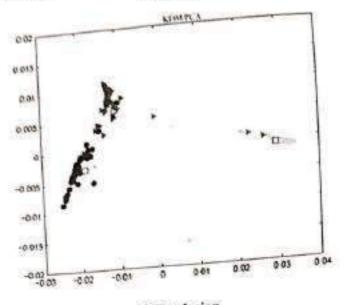


Figure 1(c) : The mapping of 10 percent of the Cora graph (10-folds setting) obtained by the five projection methods



V Conclusion

Let us now come back to our research questions. As a firstobservation, we can say that procedure(stochastic complementation followed by a diffusion mapprojection) provides an embedding in a lowdimensionalsubspace from which useful information can be extracted.Indeed, the experiments show that highly related elementsare displayed close together while poorly related elementstend to be drawn far apart. This is quite similar tocorrespondence analysis to which the procedure is closelyrelated. Second, it seems that stochastic proximity preserves complementationreasonably information, when combined with a diffusion map (KDM PCA) or an ISOMAPprojection (MDS). For the diffusion map, this is normal, since both stochastic complementation and the diffusionmap distance are based on a Markov chain model-stochasticcomplementation is the natural technique allowingto censor states of a Markov chain. On the contrary,stochastic complementation should not be combined with LaplacianEigenmap, a curvilinear component analysis, ora Sammon nonlinear mapping-the resulting mapping isnot accurate. Finally, the KDM PCA provides exactly thesame results as the basic diffusion map when t is large. However, when the parameter t is low, the resultingprojection tends to highlight the outlier nodes and tomagnify the relative differences between nodes. It istherefore recommended to display a whole range ofmappings for several different values of t.

280

ISBN: 978-93-86647-85-6

Second International Conference on Recent Innovations in Engineering and Technology (ICRIEAT-2017)

References

- P. Baldi, P. Frasconi, and P. Smyth, Modeling the Internet and theWeb: Probabilistic Methods and Algorithms. John Wiley & Sons, 2003.
- [2] M. Belkin and P. Niyogi, "LaplacianEigenmaps and SpectralTechniques for Embedding and Clustering," Advances in NeuralInformation Processing Systems, vol. 14, pp. 585-591, MIT Press, 2001.
- [3] M. Belkin and P. Niyogi, "LaplacianEigenmaps for DimensionalityReduction and Data Representation," Neural Computation, vol. 15, pp. 1373-1396, 2003.
- [4] C. Blake, E. Keogh, and C. Merz, "UCI Repository of MachineLearning Databases," Univ. California, Dept. of Information andComputer Science, <u>http://www.ics.uci.edu/~mlearn/MLRepository.htm</u> 1, 1998.
- [5] J. Blasius, M. Greenacre, P. Groenen, and M. van de Velden, "Special Issue on Correspondence Analysis and Related Methods," Computational Statistics and Data Analysis, vol. 53, no. 8,pp. 3103-3106, 2009.
- [6] I. Borg and P. Groenen, Modern Multidimensional Scaling: Theoryand Applications. Springer, 1997.
- [7] P. Bremaud, Markov Chains: Gibbs Fields, Monte Carlo Simulation, and Queues. Springer-Verlag, 1999.
- [8] P. Carrington, J. Scott, and S. Wasserman, Models and Methods inSocial Network Analysis Cambridge Univ. Press, 2006.
- [9] S. Chakrabarti, Mining the Web: Discovering Knowledge fromHypertext Data, Elsevier Science, 2003.
- [10] F.R. Chung, Spectral Graph Theory. Am. Math. Soc., 1997.
- [11] D.J. Cook and L.B. Holder, Mining Graph Data. Wiley and Sons,2006.
- [12] T. Cox and M. Cox, Multidimensional Scaling, second ed. Chapmanand Hall, 2001.
- [13] N. Cressie, Statistics for Spatial Data. Wiley, 1991.

- [14] P. Demartines and J. Herault, "Curvilinear Component Analysis: A Self-Organizing Neural Network for Nonlinear Mapping ofData Sets," IEEE Trans. Neural Networks, vol. 8, no. 1, pp. 148-154 Jan. 1997.
- [15] C. Ding, "Spectral Clustering," Tutorial presented at the 16thEuropean Conf. Machine Learning (ECML '05), 2005.
- [16] P. Domingos, "Prospects and Challenges for Multi-Relational DataMining," ACM SIGKDD Explorations Newsletter, vol. 5, no. 1,pp. 80-83, 2003.
- [17] F. Fouss, A. Pirotte, J.-M. Renders, and M. Saerens, "Random-Walk Computation of Similarities between Nodes of a Graph, with Application to Collaborative Recommendation," IEEE Trans Knowledge and Data Eng., vol. 19, no. 3, pp. 355-369, Mar. 2007.
- [18] E. Fouss, J.-M. Renders, and M. Saerens, "Links betweenKleinberg's Hubs and Authorities, Correspondence Analysis andMarkov Chains," Proc. Third IEEE Int'l Conf. Data Mining (ICDM), pp. 521-524, 2003.
- [19] F. Fouss, L. Yen, A. Pirotte, and M. Saerens, "An ExperimentalInvestigation of Graph Kernels on a Collaborative RecommendationTask," Proc. Sixth Int'l Conf. Data Mining (ICDM '06), pp. 863-868, 2006.
- [20] F. Geerts, H. Mannila, and E. Terzi, "Relational Link-BasedRanking," Proc. 30th Very Large Data Bases Conf. (VLDB), pp. 552-563, 2004.
- [21] X. Geng, D.-C.Zhan, and Z.-H. Zhou, "Supervised NonlinearDimensionality Reduction for Visualization and Classification,"IEEE Trans. Systems, Man, and Cybernetics, Part B: Cybernetics, vol. 35, no. 6, pp. 1098-1107, Dec. 2005.
- [22] Introduction to Statistical Relational Learning, L. Getoor and B. Taskar, eds. MIT Press, 2007.
- [23] J. Gower and D. Hand, Biplots. Chapman & Hall, 1995.
- [24] M.J. Greenacre, Theory and Applications of Correspondence Analysis Academic Press, 1984.

1

DATAANALYSIS PROCEDURE FOR DISCOVERING RELATIONSHIPS IN A GRAPH WITH RESPECT TO NETWORKING

¹Chandrasekaran K Arun, ²Dr. Vilas M Ghodki, ³Dr.S.B.Kishor
 ¹Research Scholar, Gondwana University, Mahaharashtra, India
 ²Associate Professor, Dept. of Computer Science, J.B. College, Wardha, Maharashtra, India
 ³Head, Dept. of Computer Studies and Research, S.P College, Chandrapur, Maharashtra, India
 E-Mail: ¹bmk_17@yahoo.com, ²vilasghodki@gmail.com, ³s.b.kishor.spc@gmail.com

Abstract - This paper precisely proposes a link-analysis based technique allowing to discover relationships existing between nodes in a computer network or, more generally, a graph. More specifically, this work is based on a random-walk through the database defining a Markov chain having as many states as nodes in the computer network. Suppose, for instance, we are interested in analyzing the relationships between nodes in a computer network, a two-step procedure is developed in analyzing the relationships. First, a much smaller, reduced, Markov chain, only containing the nodes but preserves the main characteristics of the initial chain, is extracted by stochastic complementation. For extracting the reduced Markov by stochastic complementation, an efficient algorithm is proposed. Secondly, the reduced chain is analyzed by, for instance, projecting the states in the subspace spanned by the right eigenvectors of the transition matrix called the basic diffusion map, or by computing a kernel principal-component analysis on a diffusion-map kernel computed from the reduced graph and visualizing the results. Indeed, a valid graph kernel based on the diffusion-map distance, extending the basic diffusion map to directed graphs, is introduced.

Keywords - Diffusion Map, Stochastic complementation, Feature Redundancy

I. INTRODUCTION

Wireless sensor networks (WSNs) are being used for diverse applications such as low cost area monitoring, environment monitoring, industrial and machine health monitoring, structural monitoring and military surveillance [1], [2]. In these applications, WSNs generate a large amount of data in the form of streams. In recent times, data mining techniques have been used to extract useful knowledge from WSN data [3], through discovering relationships among the sensor nodes which are known as behavioral patterns [4]. More recently, research has been focused to mine different types of behavioral patterns, e.g., sensor association rules [5], [6], [9] from stored (static) sensor data, context association rules [10] from sensor data stream, associated sensor patterns [7] and regularly frequent sensor patterns [8] from static as well as stream data. Traditional statistical, machine learning, pattern recognition, and data mining approaches [28] usually assume a random sample of independent objects from a single relation. Many of these techniques have gone through the extraction of knowledge from data, almost always leading, in the end, to the classical double-entry tabular format, containing features for a sample of the

population. These features are therefore used in order to learn from the sample, provided that it is representative of the population as a whole. However, real-world data coming from many fields such as World Wide Web, marketing, social networks, or biology [16] are often multi relational and interrelated. The work recently performed in statistical relational learning [22], aiming at working with such data sets, incorporates research topics, such as link analysis [36] web mining [1],[9], social network analysis [8], or graph mining[11]. All these research fields intend to find and exploit links between objects which could be of various types and involved in different kinds of relationships. On the other hand, when dealing with a starschema database, this two-step procedure reduces to multiple correspondence analysis. The proposed methodology therefore extends correspondence analysis to the analysis of a relational database. In short, this paper has three main contributions:A two-step procedure for analyzing weighted graphs or relational databases is proposed. .

• It is shown that the suggested procedure extends correspondence analysis.





• This A. C. Mark, Ph.D. SET, rendering on text of DB Science Colleges of the subary experiments of 15 years of CO and Science at PO 16(4) Recognitive process. In: Ph.D. In Physics, under Science Fractiny of RTM Support Dictory processing the Ph.D. In Physics, under Science Hashering published insepating process, and second structure of the State second in telephone and and entropy conference. He has second in telephone and an internal conference. He has second in telephone and and entropy conference. He has second in telephone and an internal conference. He has second in telephone and and entropy of the State.



De Automatiques de Science Colly, Schwitzberg in au Ansochter Professor to M.I. Pare Arts, Commerce & Science Colly, Schwitzberg and a Science Willieur, and contribution bestand subject descelapingentebre vertichte as minisher of Board of Science (Physics of ICDA National dissensity for three neuros. Recognized Ph.D. Supervisity (Physics in Opperate and many rescapit) papers in International & National Jourgal & Condensities at his credit Visited Officer University, Religions in Europe to present Condensities at his credit Visited Officer University, Religions in Europe to present







S18664

530 CHO

ISBN 978-93-5142-293-8

PSP 154

As per the New Syllabus of RTM Nagpur University, Nagpur

PHYSICS

Paper - I (201): (Oscillations, Kinetic Theory of Gases and Thermodynamics) Paper - II (202): (Gravitation, Astrophysics, Magnetism and Magnetostatics)

Dr. Dilip S. Choudhary Dr. Prakash D. Wankar Dr. Fulchand M. Nirwan Dr. Mahesh Y. Salunkhe Dr. Pramod K. Sakharka